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**Comparison of post-lethality thermal treatment conditions on the reduction of *Listeria monocytogenes* and sensory properties of vacuum packed ricotta salata cheese**

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**Abstract**

Ricotta salata is a whey protein cheese produced in Sardinia that in the last decades has been linked to several recalls and in 2012 to a severe human listeriosis outbreak. Contamination of ricotta salata with *L. monocytogenes* mainly occurs during post-process handling and generally originates from the processing environment. The application of water bath heat treatment in vacuum packed ricotta salata is a possible strategy to control *L. monocytogenes* superficial contamination. The objective of the present study was to select a heat treatment able to inactivate *L. monocytogenes* count of at least 5 log. Nine temperature time combinations, 75 °C, 85 °C and 90 °C applied for 15 min, 25 min and 40 min each were tested in ricotta wheels artificially contaminated with a mixture of 5 *L. monocytogenes* strains. Inactivation was assessed respectively one day and 30 days after heat treatment. The efficacy of treatments was evaluated based on the reduction in *L. monocytogenes* counts, on the impact on sensory properties and on the cost of the treatment. Two out of nine treatment combinations, i.e. 85 °C for 40 min and 90 °C for 40 min, were effective in reducing *L. monocytogenes* contamination level of 5 log. No significant difference was observed in sensory

properties after the heat treatments. Therefore both combinations are eligible to conduct a successive study aimed to extend the shelf-life of ricotta salata up to several months.

Keywords: *Listeria monocytogenes*, whey cheese, post-lethality treatment, sensory properties.

## 1. Introduction

Ricotta salata is a traditional whey protein cheese obtained in Sardinia (Italy) through the heat coagulation of the whey remaining after the production of sheep's milk cheeses. The main phases of production technology of ricotta salata are described as follows. Traditionally is manufactured using the whey remaining after the production of hard sheep's milk cheese, usually Pecorino Romano PDO (protected denomination of origin), which is stored in a silo at 45 °C until use. The whey is filtered and preheated at 60-70 °C using a plate heat exchanger. The whey is then transferred in large open kettles with approximately 1,200-1,500 liters capacity, added with 1% by weight of sodium chloride and heated to temperature above 80 °C for 30 minutes. As a result of heating, curd start floating on the top of liquid, this is collected using perforated scoops and transferred into plastic molds. The so called ricotta "*Toscanella*" is formed into cylindrical shapes and pressed to enhance drainage. The curd is salted either by dry-salting (5% w/v) or by brine-salting and dried for about 10 days in cold rooms at 10-12 °C. The manufacturing process result in cheese wheels weighing approximately 3 kg with a pH of 6.1-6.9,  $a_w$  of 0.940-0.970, moisture of 50-60% (< 50% if intended for grating), fat of 28-33% and protein of 14-23% (Spanu, Scarano, Spanu, Penna, Viridis, & De Santis, 2012; Spanu, Spanu, Pala, Viridis, Scarano, & De Santis, 2013). The final product is individually packed in vacuum bags and stored at refrigeration temperature with a set shelf-life which differs from three weeks up to several months, depending on the food business operators. Packaging of ricotta salata depends on the final use of the product, being ricotta salata wheels vacuum packed as a whole in shrinking bags if intended to be consumed grated, for mixing with other cheeses or as an ingredient, or cut into wedges before packaging if consumed plain. No preservatives are used for shelf life extension. In recent years contamination of ricotta salata with *Listeria monocytogenes* leaded voluntary recalls by international companies importing the product from Sardinia. In 2008 the European Commission documented a case of *L. monocytogenes* infection associated with the consumption of ricotta salata cheese (RASFF, 2008). The most recent and severe episode occurred

in the USA where a multistate outbreak of listeriosis linked to ricotta salata imported from Italy caused 20 hospitalization and 4 deaths (CDC, 2012). Heat treatments, such as thermization and pasteurization, applied to milk during cheese making and to whey during ricotta production inactivate *Listeria* cells to levels of approximately 3 to 6 log<sub>10</sub> cfu (Buazzi, Johnson, & Marth, 1992; Casadei, Esteves de Matos, Harrison, & Gaze, 1998; ICMSF, 1996; Villani, Pepe, Mauriello, Moschetti, Sannino, & Coppola, 1996). Contamination of whey cheeses with *L. monocytogenes* origins from the processing environment and is localized almost exclusively on the rind, with a reported prevalence in ricotta salata of approximately 20% (Pintado & Malcata, 2000; Lioliou, Litopoulou-Tzanetaki, Tzanetakis, & Robinson, 2001; Ibba, Cossu, Spanu, Viridis, Spanu, Scarano, & De Santis, 2013; Spanu, Scarano, Ibba, Spanu & De Santis, 2015). The intrinsic properties of ricotta salata support the growth of *L. monocytogenes*, once onto the product, to level as high as 7.0 log<sub>10</sub> cfu g<sup>-1</sup> of rind, potentially harmful to human health (Spanu, Scarano, Spanu, Penna, Viridis, & De Santis, 2012). Ricotta salata produced in Sardinia is mainly exported in North America and in other European countries. However, international health authorities accept different health risk for *L. monocytogenes*, leading to an absence in 25 g recommended by FDA and 10<sup>2</sup> cfu g<sup>-1</sup> criteria at the time of consumption set by European Commission (EC) Regulation No. 2073/2005. Even with a strict application of good hygienic practices during production, superficial contamination of ricotta salata could not be totally avoided, but only reduced (Tompkin, Scott, Bernard, Sveum, & Gombas, 1999). Therefore, the application of alternative control strategies should be applied if the product is exposed to environmental contamination after the lethality treatment (e.g., cooking) and before packaging (FSIS, 2014). In order to reduce *L. monocytogenes* contamination in ready to eat food, a number of post-package decontamination methods have been proposed, such as thermal pasteurization, irradiation and high-pressure. The efficacy of these decontamination technologies in different ready to eat products have been reviewed (Zhu, Du, Cordray, & Ahn, 2005). The final choice of the treatment to apply in ready to eat food stays on the food business operator based on scientific evidences on the efficacy, but is certainly cost-oriented. Heat post-lethality treatments (i.e. hot water bath and steam pasteurization) are widely used in the food industry due to their effectiveness in reducing the load of pathogenic microorganisms (Arnoldi, 2002; Orta-Ramirez & Smith, 2002). The effectiveness of a thermal treatment is influenced by several factors such as temperature-time

ratio, food composition, size and weight of the product and microorganism characteristics (Doyle, Mazzotta, Wang, Wiseman, & Scott, 2001; Ray, 2004; Sofos, 2002; Yen, Sofos, & Schmidt, 1991). Hence, the validation of a post-lethality treatment should be designed around the product, taking into account the formulation, packaging and the expected storage and use conditions. Therefore, results obtained on a specific product cannot be extended on another product, even if similar. Previous research demonstrated the efficacy of the immersion of vacuum packed ricotta salata wheels in water bath at 85 °C for 90 min in reducing *L. monocytogenes* counts of 6 log<sub>10</sub> cfu g<sup>-1</sup> of rind (Spanu, Spanu, Pala, Virdis, Scarano, & De Santis, 2013). However, the effect on sensory characteristics of such treatment was not investigated. Any technological interventions that negatively affect the sensory quality of a product become useless for a commercial purpose. For this reason sensory evaluation play a non negligible role in this type of investigation. The objective of the present study was to compare 9 different temperature-time conditions for the superficial treatment of whole ricotta salata wheels. The efficacy will be evaluated taking into account the extent of reduction and survival of artificially inoculated *L. monocytogenes* and the impact on sensory properties. The results will be used to select the temperature-time ratio to perform a further study aimed to extend the shelf-life of ricotta salata up to 180 days.

## 2. Materials and methods

### 2.1. Ricotta salata samples

A total of 465 vacuum packed ricotta salata wheels were provided by a local cheese-making plant using sheep milk. Samples were randomly selected from 3 different batches (155 ricotta wheels for each batch) and stored in a cold room at 4±2 °C until the experiment was performed. Immediately after their arrival samples were labeled according to their use for the experiment. Experimental Units (EUs) were defined ricotta salata wheels artificially contaminated with *L. monocytogenes* and successively submitted to heat treatment. Positive Controls (PCs) were defined ricotta salata wheels artificially contaminated with *L. monocytogenes*. Blank Samples (BLs) were defined the units not inoculated and used to evaluate the level of natural contamination of ricotta salata with *L. monocytogenes*. Sensory Units (SEs) were defined the not inoculated

samples used to evaluate sensory properties either after heat treatment (SEt) or as control with no treatment (SEc). Composition Units (CUs) were defined not inoculated samples used for the determination of intrinsic properties (pH and  $a_w$ ) and composition (moisture, fat and proteins) after heat treatments.

## 2.2. Artificial inoculation

The Technical Guidance document prepared by the EU Community Reference Laboratory (CRL) for *L. monocytogenes* (Beaufort, Cornu, Bergis, Lardeux, & Lombard, 2014) was used for the experiment designing. A mixture of 5 *L. monocytogenes* strains was used to artificially contaminate EUs and PCs ricotta salata wheels. Of the strains that composed the inoculum one was the reference strain ATTC 19111 (serovar 1/2a) obtained from American Type Culture Collection (Manassas, VA, USA), while the other four were wild-type strains (respectively serotypes 1/2a, 1/2b, 1/2c and 4b), previously recovered from the cheese-making plant environments or from ricotta salata. The wild-type strains were selected in order to be representative of the main serotypes associated with foodborne listeriosis. All the strains were stored at -80 °C in Brain Heart Infusion (BHI) broth (Oxoid, Basingstoke, UK) with glycerol (15% v/v). The inoculum level was aimed to demonstrate a reduction in *L. monocytogenes* level, or Performance Criterion (PC), of 5  $\log_{10}$  cfu g<sup>-1</sup> of rind, considered to suffice to attain a Food Safety Objective (FSO) of 10<sup>2</sup> cfu g<sup>-1</sup> throughout the entire storage period under refrigeration. Previous experiments were conducted to standardize the preparation of inoculum according to the indications contained in the Guidelines for conducting *Listeria monocytogenes* challenge testing of foods (Scott, Swanson, Frier, Pruett jr., Sveum, Hall, Smoot, & Brown, 2005). In order to prepare cells in the same physiological state (late exponential or early stationary phase) each strain was separately inoculated into tubes containing BHI broth and cultured overnight at 30 °C in a shaking water bath (100 rev min<sup>-1</sup>). To adapt cultures at refrigeration temperatures, cells were then subcultured into 10 mL of BHI and incubated at 4±2 °C for approximately 15 days. A “mixed working culture” was obtained by transferring equal volumes of each individual culture into a sterile flask. The concentration was adjusted to ca. 10<sup>7</sup> cfu mL<sup>-1</sup> using sterile saline solution (0.85% NaCl). Plate count on Trypticase Soy Agar (TSA, Biolife, Milan, Italy) was used to confirm concentrations. The whole surface of ricotta salata wheels was evenly sprayed with 2 mL of *L. monocytogenes* mixed culture using an atomizer. A

holding period of 15 min at room temperature was allowed to inoculated samples in order to let the suspension attach, after which ricotta salata were individually vacuum packed in shrink bags (Criovac Cook-In HT-3000, Sealedair Ltd., St Neots, UK) and stored at refrigeration temperature until further use.

### 2.3. Heat treatment and experimental design

The experiment was conducted in three independent trials, one for each batch, conducted one month apart. Heat treatment was performed by immersion of vacuum packed ricotta salata wheels in hot water bath. Nine different temperature- time conditions were tested: 75 °C, 85 °C and 90 °C applied for 15 min, 25 min and 40 min each. The number and the types of ricotta salata samples used for each treatment condition are reported in table 1. Immediately after the heat treatment ricotta salata wheels were immersed in a tank containing iced water for approximately 2 hours and then stored at  $4 \pm 2$  °C until analysis. The analysis points or testing times (T) were: the day of inoculum and heat treatment, defined as  $T_0$ ; 24 hours after heat treatment, defined as  $T_1$  and 30 days after heat treatment defined as  $T_{30}$ .  $T_1$  was performed the day subsequent the heat treatment to avoid false negative caused by the presence of sub-lethally injured *L. monocytogenes* cells that may survive the heat treatment but are not immediately culturable. The PCs were analysed at  $T_0$ , 6 hours after inoculation to assess if the level of contamination was effectively  $10^5 \log_{10} \text{cfu g}^{-1}$ . The BLs were also examined at  $T_0$ , to account for eventual natural contamination of ricotta salata with *L. monocytogenes*. The EUs and CUs were submitted to heat treatment and analysed at  $T_1$  and  $T_{30}$ . Part of SE units were treated (SEt) and part, used as negative controls (SEc), were not treated (Table 1). The sampling plan with sample units, testing times and related analysis is summarized in table 2. The effective temperature obtained on ricotta salata surface during each heat treatment was monitored using an additional ricotta salata wheel where a data logger (KT 20T, Kimo, Montpon Ménéstérol, France) was placed 1.5 cm below the surface and the temperature recorder during the treatment.

### 2.4. Microbiological analysis

Detection and enumeration of *L. monocytogenes* (ISO 11290-1:1996/Amd 1:2004; ISO 11290-2:1998/Amd 1:2004) and enumeration of aerobic mesophilic bacteria (ISO 4833:2003) were conducted on 25 g of ricotta

salata aseptically collected cutting the rind up to 2 cm. In order to detect the presence of sublethally injured cells that may survive in the product but may not be cultured on selective media, on heat treated samples the enumeration of *L. monocytogenes* was also conducted using the Thin Agar Layer (TAL) method. The TAL method consists in the overlay of a nonselective agar medium onto agar plates containing a selective medium that combines the ability to enumerate and to differentiate heat injured cells (Kang and Fung, 1999; Wu and Fung, 2001). From each positive sample, 5 suspected colonies of *L. monocytogenes* were submitted to phenotypic identification. Multiplex PCR was carried out to confirm identification and to separate the major serovars (1/2a, 1/2b, 1/2c and 4b) into distinct serogroups (Doumith, Buchrieser, Glaser, Jacquet, & Martin, 2004). The *prs* gene, specific for *Listeria* spp. was used as internal amplification control. A selection of the strains recovered from EUs at T<sub>30</sub> was submitted to pulsed-field gel electrophoresis (PFGE) to confirm that the strains recovered were the same that were inoculated. From each of the 3 replicate were selected up to five strains for each temperature-time combination. In order to capture as much variability as possible a preliminary screening of isolates to submit to PFGE was conducted based on the serogroups. PFGE was carried out using the protocol proposed by Graves & Swaminathan (2001). The obtained restriction profiles were analysed by visual examination to distinguish inoculated strains among each other and from strains originating by natural contamination.

## 2.5. Physico-chemical properties and composition

Intrinsic properties and chemical composition of ricotta salata were determined to account for possible interaction with *L. monocytogenes* survival and growth. PH and a<sub>w</sub> were measured using pH meter GLP22 (Crison Instruments SA, Barcelona, Spain) and water activity meter Aqualab 4TE (Decagon, Pullman, WA, USA), respectively. Near infrared transmittance (NIT) compositional analyzer (FOSS, Eden Prairie, MN, USA) was used for the analysis of fat, moisture, protein and total solids. Differences in intrinsic properties and composition ( $\bar{x} \pm SD$ ) of ricotta salata cheese between the different temperature-time combinations used for the heat treatment and over time (T<sub>1</sub> and T<sub>30</sub>) were compared using Fisher's least significant difference (LSD) test. Statistical analysis was performed with Statgraphics Centurion XVI software (StatPoint Technologies, Warrenton, VA, USA).



190

191 *2.6. Sensory analysis*

192 The "Difference from control test" was applied to highlight sensory differences between heat treated samples  
 193 (SEt) and the negative control (SEc). This test is very helpful to determine difference between one or more  
 194 samples against the control and, if the difference is significant, to measure its size (Meilgaard, Civille, &  
 195 Carr, 1999). On the other hand this test can cause a sensory fatigue when many samples have to be taken into  
 196 account because, during each session, the control sample as reference and as blind sample must be served. In  
 197 order to avoid the sensory fatigue only five out nine temperature-time combination were evaluated: 75 °C  
 198 and 90 °C treated for 15 min and 40 min, and 85 °C treated for 25 min. Thirty judges (14 females and 16  
 199 males, aged 25-50 years) specialized in dairy products, previously selected for their sensitivity and after  
 200 attending a course of 60 hours in sensory analysis (ISO 8586-1: 1993), evaluated the samples against an  
 201 untreated control on a numerical category scale (0 = no difference and 9 = very large difference). Ricotta  
 202 salata samples were kept at 4-6 °C until sensory assessments. Before analysis the ricotta samples were  
 203 portioned extracting two opposing slices. The slices were further portioned into parallelepiped pieces (5 x 1.5  
 204 x 1.5 cm) and served, at room temperature, in odorless plastic containers marked with a random three-digit  
 205 number (Meilgaard, Civille, & Carr, 1999). Judges were also provided by a tray containing an unsalted  
 206 cracker and a glass of water. The evaluation was carried out in a randomized and balanced order (Macfie,  
 207 Bratchell, Greenhoff, & Vallis, 1989). Statistical analysis was performed with Statgraphics Centurion XVI  
 208 software (StatPoint Technologies) by the one-way ANOVA (factor: samples) and the Fisher's LSD.

209

210 *2.7. Validation of heat treatment*

211 The experiment was conducted in three independent trials for each of the 9 temperature-time combinations.  
 212 Samples used in each trial belonged to three different production batches (batch A, B and C). Analyses were  
 213 conducted at two different sampling times ( $T_1$  and  $T_{30}$ ). For each heat treatment combination and sampling  
 214 time were analyzed three samples. To account for a margin of safety, the effectiveness of heat treatment, i.e.  
 215 performance standard ( $\Delta$ ) was considered in the worst conditions, i.e. the minimum level of  $\log_{10} \text{cfu g}^{-1}$   
 216 reduction in *L. monocytogenes* counts. Reduction obtained as consequence of heat treatments was calculated

independently for each batch, by computing the minimum difference between the concentration ( $\log_{10}$  cfu g<sup>-1</sup>) before the treatment ( $T_0$ ) and after the treatment ( $T_1$  and  $T_{30}$ ) observed in the triplicate samples.

## 2.8. Statistical analysis

Mean mesophilic bacteria counts ( $\log_{10}$  cfu g<sup>-1</sup>), intrinsic properties ( $\bar{x} \pm \text{SD}$ ) and composition ( $\% \pm \text{SD}$ ) between the different temperature-time combinations at  $T_1$  and  $T_{30}$  were compared using Fisher's least significant difference (LSD) test. All statistical analyses were performed with Statgraphics Centurion XVI software (StatPoint Technologies, Warrenton, VA, USA).

## 3. Results

### 3.1. *L. monocytogenes* contamination and background microflora

Natural contamination of ricotta salata rind with *L. monocytogenes* occurred in 6 out of 18 BLs (30.0%) all originating from the first batch. Enumeration of *L. monocytogenes* was possible in five BLs, showing a level of contamination of  $2.68 \pm 0.51 \log_{10}$  cfu g<sup>-1</sup> ( $\bar{x} \pm \text{SD}$ ). The mean level of artificial contamination expressed as  $\log_{10}$  cfu g<sup>-1</sup> ( $\bar{x} \pm \text{SD}$ ) obtained on the rind of PCs units at  $T_0$  was  $4.82 \pm 0.43$ ,  $5.5 \pm 0.04$  and  $5.36 \pm 0.09$  in the first, second and third replicate, respectively. Enumeration of aerobic mesophilic bacteria was conducted on 18 BLs, 9 PCs and 162 EUs. In BLs aerobic mesophilic population ( $\bar{x} \pm \text{SD} \log_{10}$  cfu g<sup>-1</sup>) was  $7.41 \pm 0.47$ ,  $7.45 \pm 1.04$ ,  $7.83 \pm 1.00$  while in PCs was  $7.21 \pm 0.31$ ,  $7.78 \pm 0.68$  and  $8.83 \pm 0.11$  in the first, second and third batch respectively. The mean  $\log_{10}$  cfu g<sup>-1</sup> reduction in total bacterial counts observed at  $T_1$  ranged between ca. 1.0 and 3.0 for ricotta treated at 75 °C, 4.0 and 5.5 for treatment at 85 °C and from 5 to 6 for 90 °C. After 30 days of refrigerated storage the microbial population increased of less than 1.0  $\log_{10}$  cfu g<sup>-1</sup> in ricotta salata treated at 75 °C and of ca. 1.0 and 2.0  $\log_{10}$  cfu g<sup>-1</sup> in samples treated respectively at 85 °C and 90 °C. Pair-wise comparison of aerobic mesophilic bacteria counts between ricotta salata samples submitted to the 9 treatment combinations and between samples analysed at  $T_1$  and  $T_{30}$  are reported in table 3.

### 3.2. Inactivation and survival of *L. monocytogenes*

The lethal effect was evaluated on 9 EUs (3 for each replicate) for each temperature-time combination at  $T_1$  and  $T_{30}$ . The minimum differences in *L. monocytogenes* counts between  $T_1$  and  $T_0$  ( $\Delta_1$ ) and  $T_{30}$  and  $T_0$  ( $\Delta_{30}$ ) for each of the 9 treatments are reported in table 4. At  $T_1$  three out of nine combinations, i.e. 85 °C for 40 min, 90 °C for 25 min and 90 °C for 40 min, were effective either with the enumeration and detection methods. However, the 90 °C for 25 min combination showed the survival of *L. monocytogenes* after storage at 4 °C for 30 days. The complete description of the effect of each treatment at  $T_1$  and  $T_{30}$  is reported in table 5 and figure 1.

Overall, 334 strains were confirmed as *L. monocytogenes* by molecular identification. Twenty-seven strains (90.0%) isolated from BLs were serogroup 1/2a, while 3 (10.0%) were serogroup 1/2c. Of the 49 strains isolated from PCs, 17 (34.7%) were serogroup 1/2a, 15 (30.6%) serogroup 1/2b, 11 (22.4%) serogroup 1/2c and 6 (12.2%) serogroup 4b. From EUs at  $T_1$  were isolated 95 strains which were grouped as follows: 32 (33.7%) serogroup 1/2a, 12 (12.6%) serogroup 1/2b, 16 (16.8%) serogroup 1/2c and 35 (36.8%) serogroup 4b. From EUs at  $T_{30}$  were isolated 160 strains which were grouped as follows: 52 (32.5%) serogroup 1/2a, 23 (14.4%) serogroup 1/2b, 41 (25.6%) serogroup 1/2c and 44 (27.5%) serogroup 4b. Of *L. monocytogenes* recovered from EUs at  $T_{30}$  were submitted to PFGE 19, 13 and 30 strains from batch A, B and C respectively. Strains recovered showed the same PFGE profile of the inoculated strains and belonged exclusively to the wild type: 18 (29.0%) were 1/2a, 12 (19.4%) were 1/2b, 13 (21.0%) were 1/2c and 19 (30.6%) were 4b.

### 3.3. Temperature monitoring

The initial temperature of ricotta salata rind before the immersion in hot water was  $6.0 \pm 0.8$  °C. Figure 2 shows the temperature profile recorded on the rind of ricotta salata during water bath heat treatments for the three temperatures.

### 3.4. Sensory features

The results of sensory analysis (average values  $\pm$ SD) are shown in table 7. The blind control allowed estimating the *placebo* effect, produced by asking to find a difference when in fact no differences exist.

Statistic inferences are estimates by comparing the samples and the blind control. All the heat treated samples are significantly different from the untreated one (blind control) with the exception of that treated at lowest temperature-time combination (75 °C for 15 min). However the difference size goes from 1.3 to 1.8 points that converted in the verbal scale correspond to “slight/moderate difference”. The heat treated samples were not different between each other, meaning that heating from 75 to 90 °C for a time ranging between 15 and 40 min do not significantly change the sensory characteristics of ricotta salata.

### 3.5. Ricotta salata composition

Physico-chemical characteristics were determined on untreated samples (18 BLs) and heat treated samples (162 CUs). Intrinsic properties values ( $\bar{x} \pm \text{SD}$ ) for untreated samples were  $6.42 \pm 0.09$  for pH and  $0.963 \pm 0.01$  for  $a_w$ . Composition values ( $\% \pm \text{SD}$ ) were respectively of  $55.35 \pm 2.09$  for moisture,  $21.75 \pm 2.42$  for fat,  $14.55 \pm 1.37$  for proteins and  $4.56 \pm 1.38$  for salt. Differences in intrinsic properties and composition between the 9 treatment combinations at  $T_1$  and  $T_{30}$  are reported in table 6. PH and  $a_w$  were always within limits for *L. monocytogenes* growth and no significant difference was observed between values of ricotta salata submitted to heat treatment with different temperature-time conditions ( $P > 0.05$ ).

## 4. Discussion

In the last decades ricotta salata has been associated with several recalls due to *L. monocytogenes* contamination and more recently even with foodborne listeriosis outbreaks. Contamination of ricotta salata with *L. monocytogenes* mainly origins from food processing environment and is localized almost exclusively on the rind (Pilo, Marongiu, Corgiolu, Viridis, Scarano, & De Santis, 2007). Whole ricotta salata wheels are generally intended to be consumed grated including the rind. Post-process control strategies are needed in order to comply with international health authorities limits. Hot water bath treatment in vacuum packed ricotta salata has been previously evaluated, demonstrating to be an effective and economic method to inactivate surface contamination of ricotta salata cheese (Spanu, Spanu, Pala, Viridis, Scarano, & De Santis, 2013). However, optimization of the process was needed in order to account for the level of reduction in *L. monocytogenes* counts ( $\log_{10} \text{cfu g}^{-1}$ ), changes in sensory properties and cost of the treatment. In the present study was compared the listericidal effect of nine temperature-time combinations for the treatment of

artificially contaminated whole ricotta salata wheels. The recovery of *L. monocytogenes* from not inoculated blank samples, confirms that natural contamination of ricotta salata is not a rare finding. The prevalence of contamination of ricotta salata produced in Sardinia is estimated around 20% (Ibba, Cossu, Spanu, Viridis, Spanu, Scarano, & De Santis, 2013; Spanu, Scarano, Ibba, Spanu & De Santis, 2015). This level can result, when ricotta is stored at refrigeration temperatures for up to two months, in concentration of the pathogens of approximately  $10^7 \log_{10} \text{ cfu g}^{-1}$ , potentially harmful to human health (Spanu, Scarano, Spanu, Penna, Viridis, & De Santis, 2012). The present study was aimed to validate the temperature-time combinations able to reduce *L. monocytogenes* concentration of  $5 \log \text{ cfu g}^{-1}$ , considered sufficient to comply with the food safety objective of  $<100 \text{ cfu g}^{-1}$  for the products placed on the market during their shelf-life (EC, 2005).

Enumeration of *L. monocytogenes* in positive control units confirmed that the desired level of contamination of  $10^5 \text{ cfu g}^{-1}$  was obtained. Out of nine temperature-time combinations only two,  $85^\circ\text{C}$  for 40 min and  $90^\circ\text{C}$  for 40 min, showed to be effective in reducing *L. monocytogenes* to undetectable levels either at  $T_0$  and  $T_{30}$ . These combinations allowed to reach respectively  $56.3 \pm 1.5^\circ\text{C}$  and  $57.7 \pm 1.4^\circ\text{C}$  on ricotta surface, effective in killing *L. monocytogenes*. On one hand, little or no efficacy was observed for treatment conducted at  $75^\circ\text{C}$ , regardless of the time of application, while on the other hand no efficacy was demonstrated for treatment conducted for 15 min, regardless of the temperature used. Despite an initial inactivation of up to 5.0 log at  $T_0$  as consequence of treatments performed at  $75^\circ\text{C}$  (i.e. 25 and 40 min), *L. monocytogenes* was still culturable with the detection method. The survival and successive growth during storage at refrigeration temperature for 30 days resulted in counts as high as ca. 7 log. Failure of heat treatment at  $75^\circ\text{C}$  to inactivate *L. monocytogenes* could be explained with the difference between water temperature and the maximum temperature obtained on ricotta rind ( $47.5^\circ\text{C}$ ). Treatment conducted at  $85^\circ\text{C}$  for 25 min resulted in a temperature on ricotta rind of  $49.8 \pm 1.5^\circ\text{C}$  which allowed *L. monocytogenes* survival to concentrations of up to 1 log at day zero. As consequence the microorganism grew during the successive storage to level as high as 5 log. Although *L. monocytogenes* was not countable with the enumeration method after 30 days in samples treated at  $90^\circ\text{C}$  for 25 minutes, it was still detectable with the qualitative method. No guarantee can be provided that the pathogen will not growth to levels potentially dangerous to human during ricotta salata shelf-life. The highest temperature detected on ricotta salata rind after 25 min was  $52.5$

°C for the treatment at 90 °C. This could explain the presence of heat injured cells, which recovered after the subsequent storage at refrigeration temperature for 30 days. Strains capable to survive and growth after heat treatment belonged to the wild type, suggesting that they are characterized by a greater resistance as compared with reference strains. As far as the gap between the temperature of water during treatments and the temperature recorded on the ricotta salata rind it should be noted that temperatures were detected 1.5 cm below the ricotta surface, which may underestimate the effective temperature reached on the interface between packaged ricotta and water. The heat transfer is a function of the thermal properties of foods, which depend, among other factors, by chemical composition and temperature. However, due to the complexity of heat transfer calculations, specific experiments should be conducted in order to define the specific thermal properties of ricotta salata. Changes in sensory properties of heat treated ricotta salata were observed with respect of untreated samples, but no differences were among treatments. This indicates the feasibility of using more protective treatments with no negative implication for ricotta salata sensory profile.

## 5. Conclusion

Contamination of Ricotta salata with *L. monocytogenes* can effectively be controlled by the application of water bath heat treatment applied after packaging of the product. Treatments performed at 85 °C for 40 min or 90 °C for 40 min can be effectively used to obtain a reduction of 5 log of the pathogen. No significant difference was observed in the sensory properties between the treatments. Although treatments at 85 °C for 40 min might gather food processors favors as compared to 90 °C for 40 min, the latter may provide a greater safety of the product when is stored for periods of time longer than 30 days. Treatments applied for 40 min, either at 85 °C and 90 °C, are eligible as combination of choice to be used in a further study to assess the efficacy as post-lethality treatment aimed to extend ricotta salata shelf-life.

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## References

- Arnoldi, A. (2002). Thermal processing and nutritional quality. In: Henry, C.J.K., Chapman, C. (Eds.), The Nutrition Handbook for Food Processing. Woodhead, Cambridge, England, pp. 265-286.
- Beaufort, A., Cornu, M., Bergis, H., Lardeux, A.L., Lombard, B. (2014). EURL Lm Technical guidance document for conducting shelf-life studies on *Listeria monocytogenes* in ready-to eat foods. Version 3. Community Reference Laboratory for *Listeria monocytogenes*. Available at: [http://www.fsai.ie/uploadedFiles/EURL%20Lm\\_Technical%20Guidance%20Document%20Lm%20shelf-life%20studies\\_V3\\_2014-06-06%20\(2\).pdf](http://www.fsai.ie/uploadedFiles/EURL%20Lm_Technical%20Guidance%20Document%20Lm%20shelf-life%20studies_V3_2014-06-06%20(2).pdf) Accessed October 2 2014.
- Buazzi, M. M., Johnson, M. E., & Marth, E. H. (1992). Fate of *Listeria monocytogenes* during the manufacture of mozzarella cheese. *Journal of Food Protection*, 55, 80-83.
- Casadei, M. A., Esteves de Matos, R., Harrison, S. T., & Gaze, J. E. (1998). Heat resistance of *Listeria monocytogenes* in dairy products as affected by the growth medium. *Journal of Applied Microbiology*, 84(2), 234-239.
- CDC. (2012). Multistate Outbreak of Listeriosis Linked to Imported Frescolina Marte Brand Ricotta Salata Cheese (Final Update). Available at: <http://www.cdc.gov/listeria/outbreaks/cheese-09-12/>. Accessed 10 May 2014
- Doumith, M., Buchrieser, C., Glaser, P., Jacquet, C., & Martin, P. (2004). Differentiation of the major *Listeria monocytogenes* serovars by multiplex PCR. *Journal of Clinical Microbiology*, 42, 3819-3822.
- Doyle, M. E., Mazzotta, A. S., Wang, T., Wiseman, D. W., & Scott, V. N. (2001). Heat Resistance of *Listeria monocytogenes*. *Journal of Food Protection*, 64(3), 410-429.
- EC. (2005). Commission Regulation (EC) No. 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. <http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2005:338:0001:0026:EN:PDF> Accessed 28.06.14.



Food Safety and Inspection Service. (2014). Compliance Guideline: Controlling *Listeria monocytogenes* in post-lethality exposed ready-to-eat meat and poultry products. United States Department of Agriculture. Available at: [http://www.fsis.usda.gov/wps/wcm/connect/d3373299-50e6-47d6-a577-e74a1e549fde/Controlling\\_LM\\_RTE\\_Guideline\\_0912?MOD=AJPERES](http://www.fsis.usda.gov/wps/wcm/connect/d3373299-50e6-47d6-a577-e74a1e549fde/Controlling_LM_RTE_Guideline_0912?MOD=AJPERES). Accessed 20 March 2014.

Graves, L. M., & Swaminathan, B. (2001). PulseNet standardized protocol for subtyping *Listeria monocytogenes* by macrorestriction and pulsed-field gel electrophoresis. *International Journal of Food Microbiology*, 65, 55-62.

Ibba, M., Cossu, F., Spanu, V., Viridis, S., Spanu, C., Scarano, C., & De Santis, E. P. L. (2013). *Listeria monocytogenes* contamination in dairy plants: evaluation of *Listeria monocytogenes* environmental contamination in two cheese-making plants using sheeps milk. *Italian Journal of Food Safety*, 2, 109-112.

ICMSF. (1996). Microorganisms in Foods 5: Microbiological Specification of Food Pathogens. International Commission on Microbiological Specifications for Foods (ICMSF). London: Blackie Academic & Professional, (Chapter 8).

ISO 8586-1:1993. Sensory analysis-General Guidance for The Selection, Training and Monitoring of Assessors. Part 1: Selected assessors. International Organization for Standardization, Geneva, Switzerland.

ISO 11290-1:1996/Amd 1:2004. Microbiology of food and animal feeding stuffs-Horizontal method for the detection and enumeration of *Listeria monocytogenes*. Part 1: Detection method (1st ed.), International Organization for Standardization, Geneva, Switzerland.

ISO 11290-2:1998/Amd 1:2004. Microbiology of food and animal feeding stuffs-Horizontal method for the detection and enumeration of *Listeria monocytogenes*. Part 2: Enumeration method (1st ed.), International Organization for Standardization, Geneva, Switzerland.

ISO 4833:2003. Microbiology of food and animal feeding stuffs-Horizontal method for the enumeration of microorganisms: Colony-count technique at 30 °C. International Organization for Standardization, Geneva, Switzerland.



- Kang, D. H., Fung, D. Y. (1999). Thin agar layer method for recovery of heat-injured *Listeria monocytogenes*. *Journal of Food Protection*, 62(11), 1346-1349.
- Lioliou, K., Litopoulou-Tzanetaki, E., Tzanetakis, N., & Robinson, R. K. (2001). Changes in the microflora of manouri, a Greek whey cheese, during storage. *International Journal of Dairy Technology*, 54(3), 100-106.
- Macfie, H. J. H., Bratchell, N., Greenhoff, K., & Vallis, L. V. (1989). Designs to balance the effect of order of presentation and first– order carry–over effects in hall tests. *Journal of Sensory Studies*, 4, 2, 129-148.
- Meilgaard, M., Civille, G. V., & Carr, B. T. (1999). *Sensory Evaluation Techniques*, pp. 86-91 CRC Press, Boca Raton, 1991.
- Orta-Ramirez, A., & Smith, D. M. (2002). Thermal inactivation of pathogens and verification of adequate cooking in meat and poultry products. *Advances in Food and Nutrition Research*, 44, 147-194.
- Pilo, A. L., Marongiu, P., Corgiolu, G., Viridis, S., Scarano, C., & De Santis, E. P. L. (2007). *Listeria monocytogenes* contamination sources in sheep cheese processing plants and strains virulence genes typing. *In proceeding of: 5th IDF International Symposium On The Challenge to Sheep and Goats Milk Sectors*, Alghero, Italy.
- Pintado, M. E., & Malcata, F. X. (2000). Optimization of modified atmosphere packaging with respect to physicochemical characteristics of Requeijão. *Food Research International*, 33, 821-832.
- RASFF. (2008). The Rapid Alert System for Food and Feed Annual Report. [http://ec.europa.eu/food/food/rapidalert/report2008\\_en.pdf](http://ec.europa.eu/food/food/rapidalert/report2008_en.pdf) (accessed 10.03.2014).
- Ray, B. (2004). *Fundamental food microbiology*. Boca Raton: CRC Press LLC.
- Scott, V. N., Swanson, K., Frier, T. A., Pruett jr., W. P., Sveum, W. H., Hall, P.A., Smoot, L. A., Brown, D. G. (2005). Guidelines for conducting *Listeria monocytogenes* challenge testing of foods. *Food Protection Trends*, 25(11), 818-825.
- Sofos, J. N. (2002). Approaches to pre-harvest food safety assurance. In F. J. M. Smulders & J. D. Collins (Eds.), *Food safety assurance and veterinary public health: Vol. 1* (pp. 23-48). Wageningen, Netherlands: Wageningen Academic Publishers.

- Spanu, C., Scarano, C., Spanu, V., Penna, C., Viridis, S., & De Santis, E. P. L. (2012). *Listeria monocytogenes* growth potential in Ricotta salata cheese. *International Dairy Journal*, 24, 120-122.
- Spanu, C., Spanu, V., Pala, C., Viridis, S., Scarano, C., & De Santis, E. P. L. (2013). Evaluation of a post-lethality treatment against *Listeria monocytogenes* on Ricotta salata cheese. *Food Control*, 30(1), 200-205.
- Spanu, C., Scarano, C., Ibba, M., Spanu, V., & De Santis, E. P. L. (2015). Occurrence and traceability of *Listeria monocytogenes* strains isolated from sheep's milk cheese-making plants environment. *Food Control*, 47, 318-325.
- Tompkin, R. B., Scott, V. N., Bernard, D. T., Sveum, W. H. & Gombas, K. S. (1999). Guidelines to prevent post-processing contamination from *Listeria monocytogenes*. *Dairy, Food and Environmental Sanitation*, 19, 551-562.
- Villani, F., Pepe, O., Mauriello, G., Moschetti, G., Sannino, L., & Coppola, S. (1996). Behavior of *Listeria monocytogenes* during the traditional manufacture of water-buffalo Mozzarella cheese. *Letters in Applied Microbiology*, 22, 357-360.
- Wu, V. C. H., Fung, D. Y. C. (2001). Evaluation of thin agar layer method for recovery of heat-injured foodborne pathogens. *Journal of Food Science*, 66(4), 580-583.
- Yen, L. C., Sofos, J. N., & Schmidt, G. R. (1991). Effect of meat curing ingredients on thermal destruction of *Listeria monocytogenes* in ground pork. *Journal of Food Protection*, 54, 408-412.
- Zhu, M., Du, M., Cordray, J., & Ahn, D.U. (2005). Control of *Listeria monocytogenes* contamination in ready-to-eat meat products. *Comprehensive Reviews in Food Science and Food Safety*, 4(2), 34-42.

1 Table 1. Temperature-time combinations used for water bath heat treatment of ricotta salata

Temperature-time condition		Ricotta salata samples					Total
Temperature	Minutes	BL <sup>1</sup>	PC <sup>2</sup>	EUs <sup>3</sup>	CUs <sup>4</sup>	SEs <sup>5</sup>	
No treatment	-	18	9	-	-	42	69
75 °C	15	-		18	18	12	48
	25	-		18	18	3	39
	40	-		18	18	12	48
85 °C	15	-		18	18	3	39
	25	-		18	18	12	48
	40	-		18	18	3	39
90 °C	15	-		18	18	12	48
	25	-		18	18	3	39
	40	-		18	18	12	48
Total		18	9	162	162	114	465

2 <sup>1</sup>BLs (Blank Samples): not inoculated units; <sup>2</sup>PCs (Positive Controls): samples inoculated with *L.*  
3 *monocytogenes*; <sup>3</sup>EUs (Experimental Units): samples inoculated with *L. monocytogenes* and successively  
4 heat treated; <sup>4</sup>CUs (Composition Units): heat treated units for physico-chemical analysis; <sup>5</sup>SEs (Sensory  
5 Units): samples used to assess the effect of heat treatment on sensory properties.

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15 Table 2. Number of ricotta salata wheels and analysis performed at each sampling time.

Analysis	Test units	Sampling time			
		T <sub>0</sub> <sup>a</sup>	T <sub>1</sub> <sup>b</sup>	T <sub>30</sub> <sup>c</sup>	Total
Detection and enumeration of <i>L. monocytogenes</i> and aerobic mesophilic bacteria	BLs <sup>1</sup>	18	-	-	-
	PCs <sup>2</sup>	9	-	-	9
	EUs <sup>3</sup>	-	81	81	162
	CUs <sup>4</sup>	-	81	81	162
Intrinsic properties and composition	BLs <sup>1</sup>	18	-	-	-
	CUs <sup>4</sup>	-	81	81	162
Sensory analysis	SEs <sup>5</sup>				38

16 Superscript letters are referred to the time between inoculation and analysis: <sup>a</sup> = day of inoculum and heat  
 17 treatment; <sup>b</sup> = 24 hours after heat treatment; <sup>c</sup> = 30 days after heat treatment. Superscript numbers are referred  
 18 to test units: <sup>1</sup>BLs (Blank Samples): not inoculated units; <sup>2</sup>PCs (Positive Controls): samples inoculated with  
 19 *L. monocytogenes*; <sup>3</sup>EUs (Experimental Units): samples inoculated with *L. monocytogenes* and successively  
 20 heat treated; <sup>4</sup>CUs (Composition Units): heat treated units for physico-chemical analysis; <sup>5</sup>SEs (Sensory  
 21 Units): samples used to assess the effect of heat treatment on sensory properties.

Table 3. Comparison of aerobic mesophilic bacteria counts ( $\log_{10}$  cfu  $g^{-1}$ ;  $\bar{x} \pm SD$ ) of heat treated ricotta salata with 9 different temperature-time combinations analyzed 24 h after the treatment ( $T_1$ ) and after storage at refrigeration temperature for 30 days ( $T_{30}$ ).

Treatment		Aerobic mesophilic bacteria			
Temperature	Minutes	+ve/n	$T_1$	+ve/n	$T_{30}$
75 °C	15	9/9	$6.94 \pm 0.62^A$	9/9	$7.06 \pm 0.80^A$
	25	9/9	$5.69 \pm 0.55^B$	9/9	$6.20 \pm 1.36^A$
	40	9/9	$5.12 \pm 0.42^B$	7/9	$6.50 \pm 1.28^A$
85 °C	15	9/9	$3.90 \pm 0.85^C$	9/9	$4.88 \pm 0.88^B$
	25	9/9	$3.71 \pm 0.94^C$	5/9	$4.58 \pm 1.14^B$
	40	9/9	$2.45 \pm 1.97^D$	4/9	$4.14 \pm 1.78^{BC}$
90 °C	15	9/9	$2.02 \pm 1.22^D$	6/9	$4.15 \pm 0.82^{BC}$
	25	9/9	$1.68 \pm 1.29^D$	3/9	$2.90 \pm 0.78^C$
	40	9/9	$2.01 \pm 1.12^D$	3/9	$3.67 \pm 1.62^{BC}$

Means in the same column on the same testing time ( $T_1$  or  $T_{30}$ ) with different capital letter are significantly different ( $P < 0.05$ ).

48 Table 4. *Listeria monocytogenes* reduction ( $\Delta$ )<sup>a</sup> on ricotta salata rind after water bath heat treatment

Temperature	Batch	15 min		25 min		40 min	
		$\Delta T_1$	$\Delta T_{30}$	$\Delta T_1$	$\Delta T_{30}$	$\Delta T_1$	$\Delta T_{30}$
75 °C	A	-0.23	4.18	-4.54	2.26	-3.24	2.57
	B	-2.59	0.48	-5.45	-5.45	-5.45	-5.45
	C	-1.78	-1.36	-5.27	-1.73	-5.27	-5.27
85 °C	A	-4.54	-4.54	-4.54	-4.54	-4.54	-4.54
	B	-5.45	-1.51	-5.45	-0.22	-5.45	-5.45
	C	-2.15	-2.67	-3.97	-5.27	-5.27	-5.27
90 °C	A	-4.54	-4.54	-4.54	-4.54	-4.54	-4.54
	B	-5.45	-5.45	-5.45	-5.45	-5.45	-5.45
	C	-3.97	-2.13	-5.27	-5.27	-5.27	-5.27

49 <sup>a</sup>Values are the difference between concentration ( $\log_{10}$  cfu g<sup>-1</sup>) the day of artificial inoculation ( $T_0$ ) and 24  
 50 hours ( $\Delta T_1$ ) and 30 days ( $\Delta T_{30}$ ) after treatment. For each batch and for each temperature-time combination  
 51 values are the minimum difference between the initial contamination level and the maximum count after the  
 52 treatment in the triplicate samples.

53 Table 5. Enumeration and detection of *L. monocytogenes* in ricotta salata artificially contaminated and heat treated with different temperature-time combinations  
 54 and relative compliance with Regulation CE limits evaluated 24 h (T<sub>1</sub>) and 30 days (T<sub>30</sub>) after the heat treatment.

Treatment			<i>L.monocytogenes</i>						
			T <sub>1</sub>			T <sub>30</sub>			
Temperature	Minutes	Batch	Enumeration		Detection in 25 g	Enumeration		Detection in 25 g	
			+ve/n	log <sub>10</sub> cfu/g	+ve /n	+ve/n	log <sub>10</sub> cfu/g	+ve /n	
75 °C	15	A	2/3	3.67±0.89	3/3 <sup>1</sup>	3/3	8.20±0.48 <sup>2</sup>	3/3	
		B	1/3	2.86±0.00	1/3 <sup>1</sup>	1/3	5.93±0.00 <sup>2</sup>	3/3	
		C	1/3	3.49±0.00	3/3 <sup>1</sup>	3/3	3.47± 0.48 <sup>2</sup>	3/3	
	25	A	0/3	0.00±0.00	1/3 <sup>1</sup>	2/3	6.39±0.58 <sup>2</sup>	3/3	
		B	0/3	0.00±0.00	1/3 <sup>1</sup>	0/3	0.00±0.00 <sup>2</sup>	0/3	
		C	0/3	0.00±0.00	1/3 <sup>1</sup>	1/3	3.54±0.00 <sup>2</sup>	2/3	
	40	A	2/3	1.30±0.00	3/3 <sup>1</sup>	3/3	6.87±0.34 <sup>2</sup>	3/3	
		B	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		C	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
	85 °C	15	A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	1/3
			B	0/3	0.00±0.00	0/3	2/3	2.82±1.58 <sup>2</sup>	2/3
			C	1/3	3.12± 0.00	2/3 <sup>1</sup>	1/3	2.60 ± 0.00 <sup>2</sup>	3/3
25		A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		B	0/3	0.00±0.00	0/3	1/3	5.23±0.00 <sup>2</sup>	1/3	
		C	1/3	1.30± 0.00	1/3 <sup>1</sup>	0/3	0.00±0.00	1/3	
40		A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		B	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		C	0/3	0.0±0.00	0/3	0/3	0.00±0.00	0/3	
90 °C	15	A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	2/3	
		B	0/3	0.00±0.00	0/3	0/3	0.00±0.00	1/3	
		C	1/3	1.30± 0.00	3/3 <sup>1</sup>	2/3	2.92±0.22 <sup>2</sup>	2/3	
	25	A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		B	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		C	0/3	0.00±0.00	0/3	0/3	0.00±0.00	2/3	
	40	A	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		B	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	
		C	0/3	0.00±0.00	0/3	0/3	0.00±0.00	0/3	

55 Compliance are intended as follows: <sup>1</sup>not compliant with the Regulation CE 2073/2005 detection limits before the food has left the immediate control of the food  
56 business operator ( $T_1$ ); <sup>2</sup>not compliant with the Regulation CE 2073/2005 enumeration limits for the products placed on the market during their shelf-life ( $T_{30}$ ).

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Table 6. Intrinsic properties ( $\bar{x} \pm SD$ ) and composition ( $\% \pm SD$ ) of ricotta salata submitted to 9 different heat treatment combinations and analyzed 24 h ( $T_1$ ) and 30 days ( $T_{30}$ ) after storage at refrigeration temperature.

Treatment		pH		$a_w$		Moisture %		Fat %		Proteins %		NaCl %	
Temperature	Min	$T_1$	$T_{30}$	$T_1$	$T_{30}$	$T_1$	$T_{30}$	$T_1$	$T_{30}$	$T_1$	$T_{30}$	$T_1$	$T_{30}$
75 °C	15	6.41±0.12 <sup>A</sup>	6.32±0.06 <sup>A</sup>	0.959±0.01 <sup>A</sup>	0.952±0.01 <sup>A</sup>	54.96±1.44 <sup>A</sup>	54.59±1.67 <sup>A</sup>	20.97±1.88 <sup>A</sup>	20.87±2.29 <sup>A</sup>	15.19±3.38 <sup>AB</sup>	14.75±1.35 <sup>AB</sup>	5.02±0.84 <sup>A</sup>	5.50±1.33 <sup>A</sup>
	25	6.42±0.11 <sup>A</sup>	6.36±0.05 <sup>A</sup>	0.954±0.01 <sup>A</sup>	0.951±0.01 <sup>A</sup>	54.90±2.08 <sup>A</sup>	54.94±1.36 <sup>A</sup>	21.73±2.01 <sup>AB</sup>	22.70±3.01 <sup>A</sup>	15.56±2.63 <sup>B</sup>	14.05±1.09 <sup>A</sup>	5.04±0.72 <sup>A</sup>	5.57±0.79 <sup>A</sup>
	40	6.41±0.13 <sup>A</sup>	6.35±0.07 <sup>A</sup>	0.950±0.01 <sup>A</sup>	0.654±0.01 <sup>A</sup>	55.09±2.23 <sup>A</sup>	53.53±1.93 <sup>A</sup>	20.90±3.11 <sup>A</sup>	22.71±2.32 <sup>A</sup>	14.44±1.29 <sup>ABC</sup>	14.56±0.93 <sup>AB</sup>	5.18±0.30 <sup>A</sup>	5.20±0.74 <sup>AB</sup>
85 °C	15	6.41±0.12 <sup>A</sup>	6.36±0.09 <sup>A</sup>	0.952±0.01 <sup>A</sup>	0.953±0.01 <sup>A</sup>	54.53±2.04 <sup>A</sup>	54.17±2.16 <sup>A</sup>	21.31±2.26 <sup>AB</sup>	21.65±2.52 <sup>A</sup>	14.20±2.32 <sup>ABC</sup>	15.01±1.54 <sup>AB</sup>	5.26±1.12 <sup>A</sup>	4.98±0.95 <sup>AB</sup>
	25	6.41±0.12 <sup>A</sup>	6.33±0.08 <sup>A</sup>	0.956±0.01 <sup>A</sup>	0.956±0.01 <sup>A</sup>	55.40±2.03 <sup>A</sup>	54.07±2.74 <sup>A</sup>	21.23±1.73 <sup>AB</sup>	22.29±3.13 <sup>A</sup>	13.97±1.04 <sup>ABC</sup>	15.37±1.12 <sup>AB</sup>	5.24±0.84 <sup>A</sup>	4.63±0.59 <sup>B</sup>
	40	6.38±0.10 <sup>A</sup>	6.33±0.11 <sup>A</sup>	0.953±0.01 <sup>A</sup>	0.956±0.01 <sup>A</sup>	54.72±1.81 <sup>A</sup>	53.91±2.12 <sup>A</sup>	21.65±2.21 <sup>AB</sup>	22.48±2.09 <sup>A</sup>	14.95±1.78 <sup>ABC</sup>	14.94±0.97 <sup>AB</sup>	5.16±0.76 <sup>A</sup>	5.09±0.76 <sup>AB</sup>
90 °C	15	6.40±0.11 <sup>A</sup>	6.33±0.09 <sup>A</sup>	0.953±0.01 <sup>A</sup>	0.954±0.01 <sup>A</sup>	54.30±1.64 <sup>A</sup>	53.61±1.62 <sup>A</sup>	23.07±0.88 <sup>B</sup>	22.75±2.15 <sup>A</sup>	13.33±0.56 <sup>C</sup>	14.48±1.50 <sup>AB</sup>	5.36±0.66 <sup>A</sup>	5.18±0.44 <sup>AB</sup>
	25	6.38±0.12 <sup>A</sup>	6.35±0.15 <sup>A</sup>	0.955±0.01 <sup>A</sup>	0.954±0.01 <sup>A</sup>	54.78±1.66 <sup>A</sup>	54.24±2.24 <sup>A</sup>	21.74±2.01 <sup>AB</sup>	22.52±2.36 <sup>A</sup>	14.38±2.00 <sup>ABC</sup>	14.33±0.82 <sup>AB</sup>	5.12±0.90 <sup>A</sup>	5.32±0.79 <sup>AB</sup>
	40	6.35±0.11 <sup>A</sup>	6.35±0.12 <sup>A</sup>	0.954±0.01 <sup>A</sup>	0.955±0.01 <sup>A</sup>	54.58±1.37 <sup>A</sup>	54.22±2.02 <sup>A</sup>	23.03±1.08 <sup>B</sup>	21.80±2.26 <sup>A</sup>	13.70±0.54 <sup>AC</sup>	15.50±2.66 <sup>B</sup>	5.19±0.87 <sup>A</sup>	5.03±0.93 <sup>AB</sup>

Each data point is the mean of three samples. For each parameter means in the same column on the same testing time ( $T_1$  or  $T_{30}$ ) with different capital letter are significantly different ( $P < 0.05$ ).

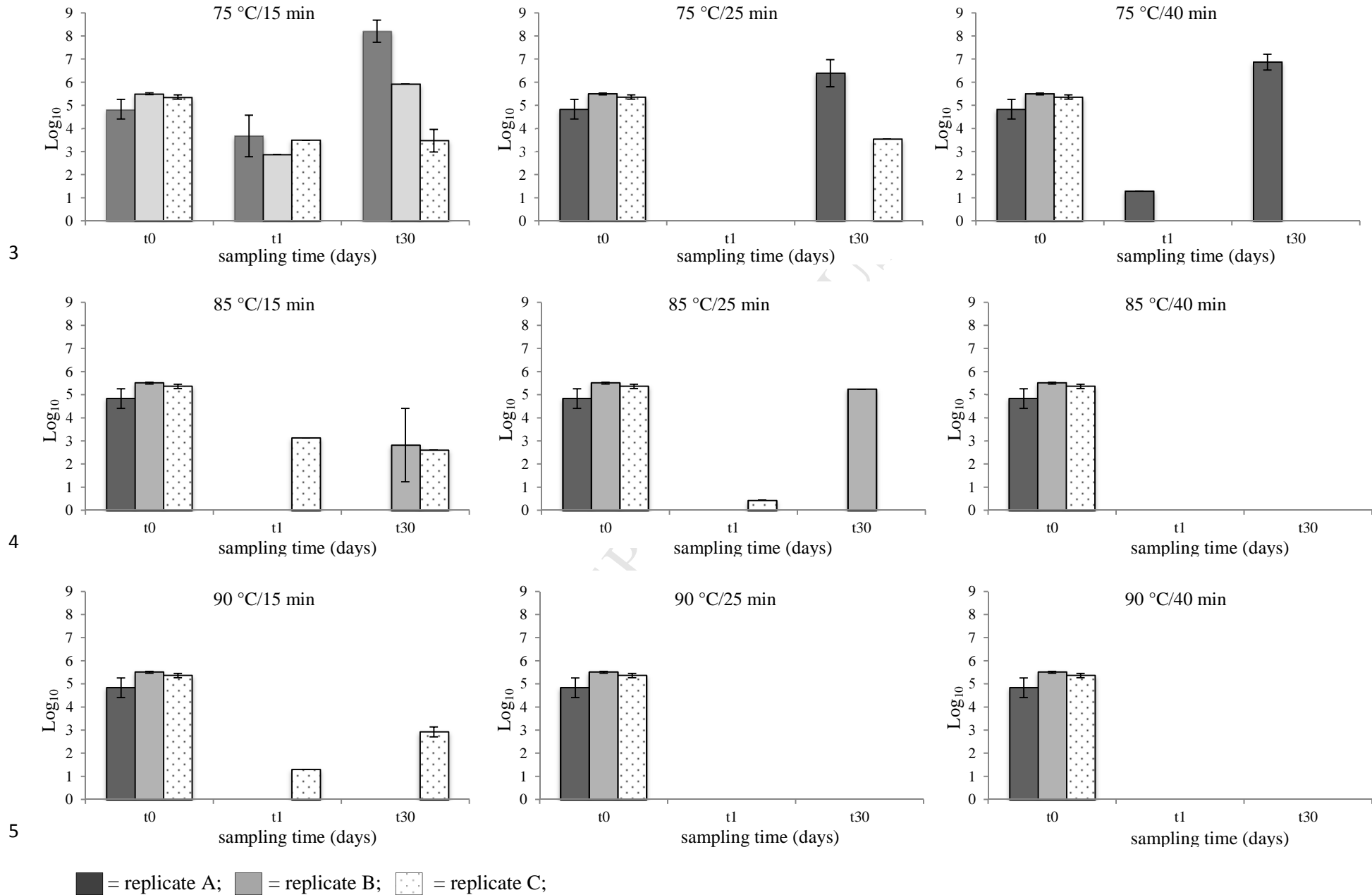
Table 7. Mean values and standard deviations of sensory differences among the control (SEc) and the samples heat treated (SEt).

Temperature-time condition	Differences
Blind samples	2.3 <sup>a*</sup> ± 1.5
75 °C x 15 min	3.4 <sup>ab</sup> ± 2.3
75 °C x 40 min	3.8 <sup>b</sup> ± 2.0
85 °C x 25 min	4.1 <sup>b</sup> ± 2.4
90 °C x 15 min	3.6 <sup>b</sup> ± 2.3
90 °C x 40 min	3.9 <sup>b</sup> ± 2.5

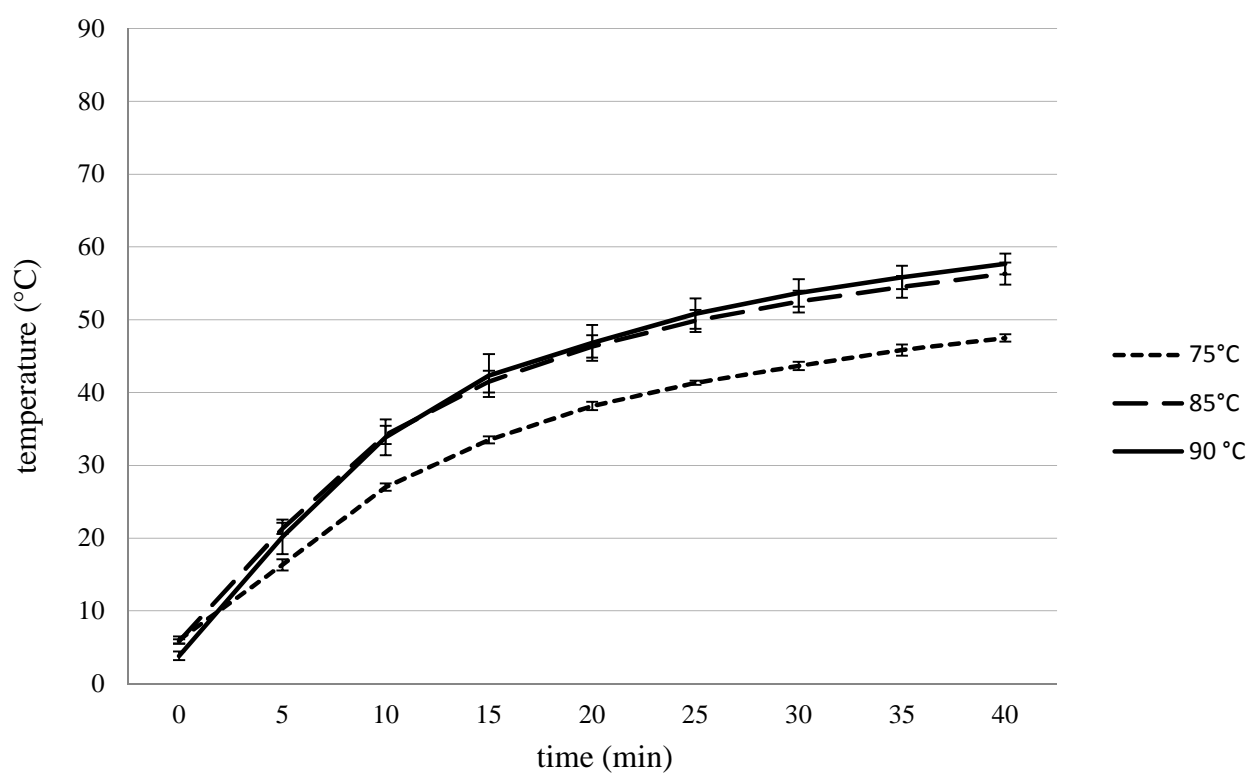
Mean values with different superscript letters are significantly different among samples.\* ( $P \leq 0.05$ ).

1 **Figure 1.** Reduction in *L. monocytogenes* counts ( $\log_{10}$  cfu  $\text{g}^{-1}$ ) of artificially contaminated Ricotta salata wheels ( $T_0$ ) analyzed 24 h ( $T_1$ ) and 30

2 days ( $T_{30}$ ) after waterbath heat treatment with 9 temperature-time combinations.



6 **Figure 2.** Temperatures recorded 1.5 cm below ricotta salata surface during water bath heat treatment



7

8 Each data point is the mean of temperatures recorder in the three replicates (batch A, B and C).

**Highlights**

1. Post-lethality treatment on *L. monocytogenes* was assessed in ricotta salata.
2. A reduction of 5 log<sub>10</sub> cfu g<sup>-1</sup> in *L. monocytogenes* count was validated.
3. No effect of treatments on sensory properties was observed.