

Effects of lactation stage, parity, β-lactoglobulin genotype and milk SCC on whey protein composition in Sarda dairy ewes

Anna Nudda¹, Maria Feligini², Gianni Battacone¹, Nicolò Pietro Paolo Macciotta¹, Giuseppe Pulina¹

¹Dipartimento di Scienze Zootecniche. Università di Sassari, Italy ²Istituto Sperimentale Italiano Lazzaro Spallanzani. Milano, Italy

Corresponding author: Prof. Giuseppe Pulina. Dipartimento di Scienze Zootecniche. Università di Sassari. Via E. De Nicola 9, 07100 Sassari, Italy - Tel. +39-079-229307 - Fax: +39-079-229302 - Email: gpulina@uniss.it

Paper received October 31, 2002; accepted February 3, 2003

ABSTRACT

In 90 Sarda dairy ewes the effects of lactation stage, parity, β -lactoglobulin genotypes, and somatic cell count (SCC) on the milk content of total protein (TP), casein (CN), whey protein (WP) and its fractions α -lactalbumin (ALA), β -lactoglobulin (BLG), serum albumin (SA), immunoglobulin (IG) and lactoferrin (LF) were analysed using a linear mixed model. Mean values of variables (g/l) were: TP (54.0), CN (43.0), WP (11.0), BLG (4.78), ALA (1.37), SA (0.61), IG (3.83) and LF (0.28). The lactation stage significantly affected all the variables analysed. TP, CN and WP concentrations tended to increase throughout lactation, with the increase of WP being more pronounced than the corresponding variation in CN. There was no definite trend in BLG content, whereas ALA concentration decreased as lactation progressed. The parity affected almost all variables studied. WP concentration differed significantly only between the second and fourth parity (10.45 vs 11.44 g/l). BLG and SA concentrations were significantly lower in the youngest ewes. The BLG genotype affected milk yield, but no effects were observed on the components of the milk. The SCC influenced almost all variables studied. The TP concentration was significantly higher in milk with SCC >1,000,000 (55.0 g/l) than in milk with lower SCC (53.4 g/l). This was mainly due to the increase of WP (12.52 and 10.24 g/l in milk with SCC above and below 1,000,000/ml respectively), especially in those WP fractions originating from blood.

Key words: Dairy ewes, Milk whey protein, Lactation stage, Parity, SCC

RIASSUNTO

EFFETTI DELLO STADIO DI LATTAZIONE, DELL'ORDINE DI PARTO, DEL GENOTIPO DELLA β -LATTOGLOBULINA E DEL CCS SULLA COMPOSIZIONE DELLE SIEROPROTEINE DEL LATTE IN PECORE DI RAZZA SARDA

Su un gregge di 90 pecore di razza Sarda è stato studiato l'effetto dello stadio di lattazione, dell'ordine di parto, del genotipo al locus β -lattoglobulina e del contenuto in cellule somatiche (CCS) sui contenuti in proteina totale (TP), caseina (CN) e sieroproteina (WP) e sulle frazioni sieroproteiche β -lattoglobulina (BLG), α -lattoalbumina (ALA), sieroalbumina (SA), immunoglobuline (IG) e lattofferina (LF), con l'impiego di un modello lineare misto. I valori medi delle variabili esaminate (g/l), sono stati: TP (54,0), CN (43,0), WP (11,0), BLG (4,8), ALA (1,4), SA (0,61), IG (3,83) e LF (0,28). Lo stadio di lattazione ha influenzato significativamente tutte le variabili esaminate. I contenuti in TP, CN e WP sono tenden-

zialmente aumentati nel corso della lattazione con una aumento delle WP più pronunciato della corrispondente variazione in CN. Il contenuto in BLG non ha evidenziato un trend ben definito, mentre l'ALA è diminuita con il progredire della lattazione. Il contenuto in WP è risultato significativamente differente tra gli animali di secondo e quarto ordine di parto (10.5 vs 11.4 g/l). I contenuti in BLG e SA sono risultati più bassi negli animali più giovani. Nessuna influenza dell'ordine di parto è stato riscontrato sulle altre frazioni sieroproteiche. Il genotipo al locus BLG ha influenzato significativamente solo la produzione di latte, mentre nessun effetto è stato riscontrato su tutti i componenti del latte esaminati. Il CCS ha influenzato significativamente quasi tutte le variabili analizzate. Il contenuto in TP è risultato significativamente

Il CCS ha influenzato significativamente quasi tutte le variabili analizzate. Il contenuto in TP è risultato significativamente più elevato nel latte con CCS >1,000,000 (55.0 g/l) rispetto al latte con un CCS inferiore (53.4 g/l). Ciò è dovuto principalmente ad un aumento delle WP (12.52 e 10.24 g/l nel latte con CCS maggiori e minori di 1.000.000/ml rispettivamente) ed in particolare di quelle di derivazione ematica.

Parole chiave: Pecore da latte, Sieroproteine, Stadio di lattazione, Ordine di parto, CCS

Introduction

Whey is the principal by-product of the dairy sheep industry, as almost all sheep milk is used to make cheese. Some 674,000 tons of sheep milk were produced in 1999 and 613,000 tons of this was processed into cheese (Pulina and Furesi, 2001). As a result, given an average cheese yield of 0.20, one can estimate that some 490,000 tons of whey were produced. Only a part of the whey is used to produce "Ricotta", a soft cheese cream obtained from heating the whey to 80°C. Most of it is thrown away and this creates serious waste disposal problems. Due to its chemical composition (c. 90% water, 1.25% fat, 1.63% protein and 4.75% lactose), sheep milk whey is a potential source of recyclable proteins (Comendador et al., 1996). In recent years there has been increasing interest in the principal whey proteins (WP) such as β-lactoglobulin (BLG), α-lactalbumin (ALA), immunoglobulin (IG), serumalbumin (SA) and lactoferrin (LF). This is because they have biomedical, technological and functional properties, such as water holding, gelling, surface activity (Euston and Hirst, 2000). WP may have anti-carcinogenic (Hakkak et al., 2000) and immuno-modulating effects (Cross and Gill, 1999), and is also used to produce bioactive peptides (Philanto-Leppala *et al.*, 2000). There is also a notable relationship between WP and the technical properties of milk; denaturation of BLG at high temperatures markedly reduces the rennetability of the milk (Dalgleish, 1993). Moreover, if the concentration of WP in milk is high this may reduce the drainage of curd and thus cause undesirable proteolysis or delay the maturing of the cheese (Mistry and Maubois, 1993).

The protein fraction contents in sheep milk are affected by environmental and genetic factors. Some research has been carried out on relationship between the genetic variants (A and B) of BLG and milk production, composition and coagulation properties (Giaccone *et al.*, 2000; Herget *et al.*, 1995; Rampilli *et al.*, 1997), but little information is available on the principal factors which influence the concentration of WP fractions.

Rampilli *et al.* (1997) in their study on Massese ewes reported higher BLG concentrations in BLG-AA genotype, while Duranti and Casoli (1991) found that WP content tends to increase as the SCC increases. More information on the main factors affecting the concentration and composition of WP in sheep milk is required for it to be correctly exploited.

The aim of the present work is to evaluate the effects of lactation stage, parity, BLG genotype and SCC on milk WP fractions in dairy ewes.

Material and methods

A flock of 211 Sarda ewes was screened for BLG genotypes in order to create balanced BLG groups for parity order. Three groups of 30 ewes each were selected for each BLG genotype (AA, AB, and BB). Each group was composed of 10, 5, 10 and 5 ewes for 2nd, 3rd, 4th and 5th parity. Lambings took place over period of 10 days. Ewes were fed on natural (clover) and artificial (common oats, vetch) pasture, plus a supplement of 300-400 g/head of concentrate at each milking. All 90 selected ewes tested negative for the California Mastitis Test at the

proteir	ii ii accioii ana s	300		
		n.	Mean	SE
Milk	g/d	495	1077	13.70
Fat	g/l	495	66.39	0.54
Lactose	"	495	43.90	0.29
Total protein (TP)	"	492	54.00	0.25
- Casein (CN)	"	492	42.98	0.22
CN/TP		492	0.80	0.00
Whey protein	"	491	11.03	0.14
- β-lactoglobulin	"	491	4.78	0.08
- α -lactalbumin	"	491	1.37	0.02
- Serum albumin	"	491	0.61	0.02
- Immunoglobulin	"	491	3.83	0.09
- Lactoferrin	"	476	0.28	0.01
SCC 10 ³	Log ₁₀	495	2.88	0.03

Table 1. Arithmetic means and standard errors of milk yield, milk composition, whey protein fraction and SCC

beginning of the trial. A mastitis control program was not applied during the experiment, but mastitis was detected visually in two ewes during the period of the experiment.

Individual milk yield was measured every two weeks from March to June at both daily milkings (morning and evening), beginning from 90 days in milking (DIM). Individual samples from morning and evening milking, weighted according to their respective milk yields, were analysed.

Laboratory analyses

Total nitrogen (TN), non-protein nitrogen (NPN) and non-casein nitrogen (NCN) were determined by the Kjeldahl method, following the FIL-IDF methodology (1993; 1996). Casein content (CN) was calculated as (TN - NCN) x 6.38 and total whey protein (WP) as (NCN - NPN) x 6.38. Fat and lactose concentration was measured on a Milkoscan 605 (Foss Electric, Hillerød, Denmark) and SCC on a Fossomatic 360 (Foss Electric, Hillerød, Denmark).

BLG genotypes were determined by isoelectric focusing. Electrophoretic fractioning of the whey proteins was made by SDS-PAGE, as described by Feligini *et al.* (1997). Gels were automatically scanned with a GS-670 Densitometer (Bio-rad) to determine densitometric peak areas. The data are reported as an area proportion of each WP fraction

and were converted to concentrations (g/l) of BLG, ALA, LF, SA, and IG, based on the WP concentration.

Statistical analysis

Data were analysed using the mixed linear model:

$$Y_{ijklpo} = \mu + DIM_i + P_j + G_k + SCC_l + (DIM \times SCC)_{il} + E_p + \varepsilon_{ijklpo}$$

where

Y = the dependent variable considered,

μ = overall mean,

DIM_i = fixed effect of DIM i (i = 6 DIM of 2 week intervals each, starting from 90 DIM),

P_j = fixed effect of the parity j (j = 2,3,4,5 parity),

 G_k = fixed effect of the BLG genotype k (k: AA; AB; BB),

SCC₁ = fixed effect of somatic cells (1 = 3 levels; SCC<500,000; SCC between 500,000 and 1,000,000; and SCC >1,000,000 cells/ml),

 E_p = random effect of ewe p (p = 1...90),

 ε_{ijkpo} = random residual.

The SCC classes were established following those suggested by Bergonier and Berthelot (2003), to distinguish between healthy and infected udders.

Results and discussion

Means of milk yield, fat, lactose, TP, CN, WP and WP fractions and SCC are reported in table 1. Daily milk yield and SCC values were close to the mean for the Sarda breed (ARA, 2002). Fat, lactose, TP, CN and WP contents are within the range of values reported in other studies for Sarda (Pulina et al., 1993) and other Mediterranean dairy sheep breeds (Cavani et al., 1991; Fuertes et al., 1998). BLG and ALA concentrations agree with previous results in dairy sheep (Duranti and Casoli, 1991; Law, 1995) but are higher than those reported for goats (Law, 1995) and cows (Law, 1995; Mackle et al., 1999). As cow milk is the major industrial source of serum proteins, this result is particularly interesting because sheep milk may be a new and more profitable source of whey protein. The LF was higher than the values observed in sheep (Qian et al., 1995), cow (Sanchez et al., 1988) and goat milk (Greppi and Roncada, 2001). The IG and SA concentrations were also higher than those previous reported for sheep (Duranti and Casoli, 1991; Law, 1995), cows and goats (Law, 1995).

The SCC classes significantly influenced almost all variables considered, except fat and CN content (Table 2). The highest milk yield has found in milk with SCC<1,000,000. The CN content tended to

decrease as the SCC rose, while the WP and its fractions were significantly lower in the milk with SCC<1,000,000. An increase in WP fractions is usually evidence of mammary gland inflammation (Sordillo et al., 1997; Urech et al., 1999). The high content of IG and SA in mammary secretion derives mostly by paracellular passage through disrupted tight junctions, or by enhanced specific IG receptors during involution or intramammary inflammation (Schanbacher et al., 1997). Milk LF comes from the degranulation of blood neutrophils (Neville and Zhang, 2000; Sordillo et al., 1997), which were the only type of somatic cells that increased in sheep milk as the SCC rose (Cuccuru et al., 1997), and from mammary cell synthesis (Sanchez et al., 1992; Schanbacher et al., 1997). Although mastitis was detected by visual inspection in only two ewes during the experiment, about 50% of the ewes could be considered as potentially infected according to Bergonier and Berthelot's rule (2003). Thus a possible explanation for the increase of SA and IG in the highest SCC class could be the higher supply of these proteins from blood; it cold also be partly caused by increased concentration, due to the lower milk yield with SCC over 1,000,000. The increase of BLG and ALA concentration in milk with SCC>1,000,000, should be mainly attributed to concentration effect rather than to an increase of syn-

Table 2. Least square means of milk yield, milk composition and whey protein fraction in milk of animals classified according to the SCC

		SCC classes		
		<500,000	500,000-1,000,000	>1,000,000
Milk	g/d	1114ª	1104ª	1015⁵
Fat	g/l	66.15	67.07	68.04
Lactose	**	45.33 ^A	44.42 ^A	42.35 ⁸
Total protein (TP)	w	53.57°	53.17ª	55.05⁵
- Casein (CN)	"	43.59	42.70	42.54
CN/TP		0.81 ^A	0.80 ^A	0.77⁵
Whey protein	w	10.07 ^A	10.41 ^A	12.52 ^B
- β-lactoglobulin	"	4.60 ^A	4.62 ^A	5.29 ⁸
- α-lactalbumin	w	1.28°	1.34°	1.47⁵
- Serum albumin	w	0.55°	0.60°	0.72⁵
- Immunoglobulin	w	3.30 ^A	3.44 ^A	4.44 ^B
- Lactoferrin	W	0.20 ^A	0.23 ^A	0.41 ^B

Figure 1. Time evolution of milk yield, fat, total protein, and lactose in milk with different SCC classes (<500,000; 500,000-1,000,000; >1,000,000 cells/ml)

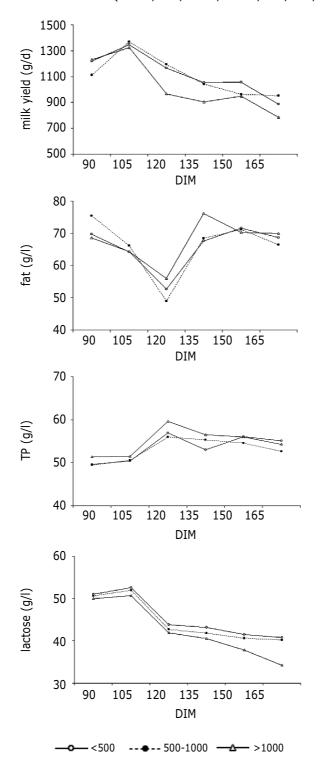
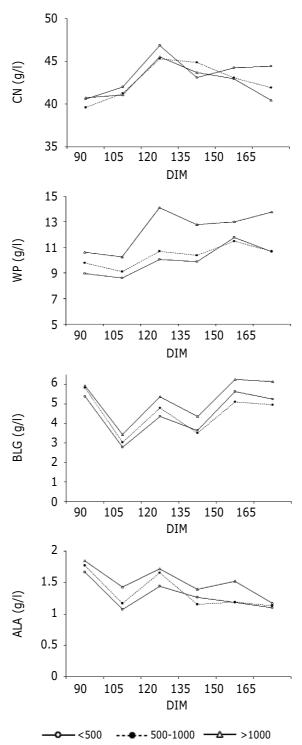


Figure 2. Time evolution of casein, total whey protein and whey protein fractions synthesised by the mammary gland in milk with different SCC classes (<500,000; 500,000-1,000,000; >1,000,000 cells/ml)



thesis of these proteins by the mammary cells: there is an increase of BLG and ALA of about 13% in the SCC>1,000,000 class, which corresponds with a reduction of milk yield of the same extent (Table 1).

DIM affected all variables studied significantly, as was expected. The patterns of the variables under consideration during lactation, separated for different SCC classes, are reported in Figure 1, 2 and 3. Milk yield (Figure 1) decreased from 105 DIM towards the end of the experiment, especially in the higher SCC class. The decrease of fat content at 120 DIM can be explained by phenomenon of subacidosis due to the high doses of concentrate in the diet and the lower NDF content of the grass in spring. The TP content increased to 120 DIM due to effects of concentration, and remained fairly stable until the end of the experiment. Lactose concentration decrease during lactation, following the pattern of milk yield, and was significantly lower in class with higher SCC only at 150 and 165 DIM. The CN content (Figure 2) showed a pattern similar to TP content in all SCC classes except at 165 DIM, where a significant difference was found between the highest and the lowest SCC class. The WP content tended to increase during lactation, but this was more marked in the highest SCC class after 120 DIM.

There was no definite trend in BLG content,

although some authors have reported an increase during lactation both in sheep (Bolla *et al.*, 1989) and goats (Quiles *et al.*, 1994). The ALA concentration tended to decrease during lactation, as did milk yield and lactose content. Similar behaviour for milk yield, lactose and ALA concentration have been already observed in dairy cattle, where the reduction in milk yield was attributed to the diminished synthesis of lactose caused by the reduction in ALA concentration (Prosser and McLaren, 1997).

The SA and IG content increased, peaking at 120 and 135 DIM, respectively (Figure 3). LF content remained fairly stable for the whole experimental period in milk with SCC<1,000,000. It was, however, markedly higher in the highest SCC class, and decreased from 120 DIM to the end of experiment.

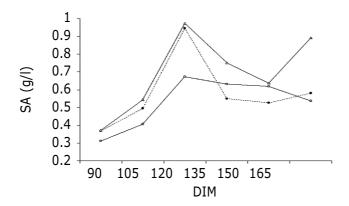
The parity influenced almost all the variables considered significantly, except CN/TP, ALA, IG and SCC (Table 3). TP, CN and WP tended to increase with parity, reaching the highest value at fourth parity. An increase of these protein with age has been reported for the Churra breed (Fuertes et al., 1998) whereas Casoli et al. (1989) reported a decrease of total WP concentration in the Massese breed with parity. BLG and SA concentrations were lowest in the youngest ewes, while no difference was observed

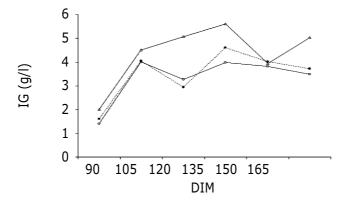
Table 3. Least square means of milk yield, milk composition, whey protein fraction and SCC in milk with different parities

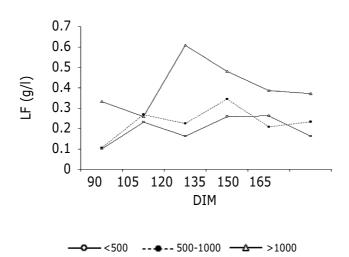
	Parity				
		2	3	4	5
Milk	g/d	1164ª	1129°	1042 ^b	975⁵
Fat	g/l	62.68 ^A	68.50 ⁸	69.75⁵	67.40 ^B
Lactose	"	44.70	43.14 ^B	44.21 ^{AB}	44.08 ^{AB}
Total protein (TP)	"	52.20°	54.22⁵	55.31⁵	54.01 ^b
- Casein (CN)	"	41.75°	42.92ab	43.88⁵	43.21 ^b
CN/TP		0.80	0.79	0.79	0.80
Whey protein	"	10.45°	11.38ab	11.44 ^b	10.72ab
- β-lactoglobulin	"	4.32°	5.20⁵	5.04⁵	4.77⁵
- α-lactalbumin	"	1.41	1.41	1.36	1.28
- Serum albumin	"	0.51 ^A	0.67 ⁸	0.66 ⁸	0.66 ⁸
- Immunoglobulin	"	3.78	3.63	3.90	3.59
- Lactoferrin	"	0.25°	0.29ab	0.33⁵	0.24
SCC 10 ³	Log ₁₀	2.90	2.90	2.90	2.84

a,bP<0.05; A,BP<0.01

Figure 3. Time evolution of whey protein deriving from blood in milk with different SCC classes (<500,000; 500,000-1,000,000; >1,000,000 cells/ml)







among other parities. This agrees with previous results obtained for the Sarda breed (Bolla *et al.*, 1989). The lower concentration in SA in second parity ewes can be explained by the better physiological condition of the udder in young animals, as is confirmed by the higher lactose concentration and milk yield. The SCC did not differ significantly among parities. This is different from previous results, where an increase of the SCC with number of lactations was reported in sheep (Bergonier *et al.*, 1996; Bencini and Pulina, 1997).

BLG genotype (Table 4) had a significant effect only on milk yield. BLG BB ewes had the lowest milk yield. No effects were observed on milk composition.

However studies on the relationships between BLG genotype and composition have produced conflicting results. Some authors found that the BLG genotype affects TP, CN and WP (Di Stasio *et al.*, 1997; Giaccone *et al.*, 1997; Herget *et al.*, 1995) whereas others did not (Barillet *et al.*, 1993; Taibi *et al.*, 1999). These differences, which also occur in studies on dairy cattle, may be due to the structure of the data analysed and the model used for statistical analysis (Giaccone *et al.*, 2000; Ng-Kway-Hang,

1997). The balanced experimental design and the mixed linear model used in this study allowed the BLG genotype classes to be compared more accurately, thus avoiding the lack of balance among genotypes, which is one of the most conspicuous features of the studies cited above.

Conclusions

Our results confirm that sheep milk has higher WP content than cow or goat milk. WP content and its fractions varied mainly according to lactation stage and milk SCC. The increase of TP in milk during lactation and in milk with SCC above 1,000,000 was principally due to the increase of WP, particularly for those fractions which originated from blood. Thus milk produced in late lactation can be considered as an important source of useful whey proteins.

The authors wish to thank Mr. Roberto Rubattu for his technical assistance.

This work was supported by the National Project DIETOLAT (MiPAF).

Table 4. Least square means of milk yield, milk composition, whey protein fraction and SCC in milk with different β -lactoglobulin genotypes

		βLG Genotype		
		AA	AB	ВВ
Milk	g/d	1116 ^A	1129 ^A	988 ^B
Fat	g/l	67.37	67.64	66.24
Lactose	"	44.38	43.76	43.97
Total protein (TP)	"	53.47	53.85	54.48
- Casein (CN)	"	42.68	42.57	43.57
CN/TP		0.80	0.79	0.80
Whey protein	"	10.79	11.31	10.89
- β-lactoglobulin	"	4.82	4.91	4.77
- α-lactalbumin	"	1.34	1.40	1.35
- Serum albumin	"	0.62	0.66	0.60
- Immunoglobulin	"	3.56	3.89	3.74
- Lactoferrin	"	0.28	0.28	0.26
SCC 10 ³	Log10	2.90	2.85	2.90

A,BP<0.01

REFERENCES

- ARA, Associazione Regionale Allevatori della Sardegna, 2002. Home page address: www.ara.sardegna.it/
- BARILLET, F., MAHÈ, M., PELLEGRINI, O., GROSCLAUDE, F., BERNARD S., 1993. Polymorphisme genetique des proteines du lait en race ovine de Lacaune. Proc. 5th Inter. Symp. Machine Milking of Small Ruminants, Budapest, Hungary: 199-207.
- Bencini, R., Pulina, G., 1997. The quality of sheep milk: a review. Austr. J. Exper. Agric. 37:485-504
- BERGONIER, D., BERTHELOT, X., 2003. New advances in epizootiology and control of ewe mastitis. Livest. Prod. Sci., 79:1-16.
- Bergonier, D., Lagriffoul, G., Berthelot, X., 1996. Facteurs de la variation des comptages de cellules somatiques chez les ovins et les caprins laitieres. In: R. Rubino (ed.), Proc. Int. Symp. on Somatic Cells and Milk of Small Ruminants, EAAP Publ., Wageningen Pers, Netherlands, pp 113-135.
- BOLLA, P., CAROLI, A., FRAGHÌ, A., PIREDDA, G., RAMPILLI, M., 1989. Quantitative and qualitative variability of β-lactoglobulin in sheep milk. Zoot. Nutr. Anim. 15:586 (Abstr.).
- Casoli, C., Duranti, E., Morbidini, L., Panella, F., Vizioli, V., 1989. Quantitative and compositional variations of Massese sheep milk by parity and stage of lactation. Small Rum. Res. 2:47-62.
- Cavani, C., Bianconi, L., Manfredini, M., Rizzi, L., Zarri, M.C., 1991. Effects of a complete diet on the qualitative characteristics of ewe milk and cheese. Small Rum. Res. 5:273-284.
- Comendador, F.J., Nardo N., Bertone, A., Maurizi, A., Quaglia, G., 1996. Effect of ultrafiltration on functional properties of sheep milk "scotta". Sci. Tec. Latt.-cas. 47:103-112.
- Cross, M.L., Gill, H.S., 1999. Modulation of immune function by a modified bovine whey concentrate. Immunol. Cell Biol. 77:345-350.
- Cuccuru, C., Moroni, P., Zecconi, A., Casu, S., Caria, A., Contini, A., 1997. Milk differential cell counts in relation to total counts in Sardinian ewes. Small Rum. Res. 25:169-173.
- Dalgleish, D.G., 1993. The enzymatic coagulation of milk. In: P.F. Fox (ed.) Cheese: chemistry, physics and microbiology. Chapman & Hall London, UK, vol.1, pp 69-100.
- DI STASIO, L., PORTOLANO, B., TODARO, M., FIANDRA, P., GIACCONE, P., FINOCCHIARO, R., ALICATA, M.L., 1997. Effect of ovine β-Lactoglobulin phenotype on cheese yield and composition. In: Proc. IDF Seminar Milk Protein Polymorphism. Publ. IDF, Brussels, Belgium, pp 324-327.
- Duranti, E., Casoli, C., 1991. Variations in the nitrogen composition and in the lactodinamographic parameters of ewe's milk in relation to somatic cell content. Zoot. Nutr. Anim. 17:99-105.

- EUSTON, S.R., HIRST, R.L., 2000. The emulsifying properties of commercial milk protein products in simple oil-in-water emulsions and in a model food system. J. Food Sci. 65:934-940.
- Feligini, M., Nudda, A., Pulina, G., Enne, G., 1997. Relationship between protein polymorphism and coagulation properties of milk in dairy ewes. pp 309-315 in Proc. 32nd Int. Symp. Anim. Prod., Elsevier, Paris, France.
- FIL-IDF. 1993. IDF standard 20b.
- FIL-IDF. 1996. IDF standard 29.
- FUERTES, J.A., GONZALO, G., CARRIEDO, J.A., SAN PRIMITIVO, F., 1998. Parameters of test day milk yield and milk components for dairy ewes. J. Dairy Sci. 81:1300-1307.
- GIACCONE, P., DI STASIO, L., PANNO, M., TODARO, M., 1997. Variazione della composizione chimica del latte nel corso della lattazione per i differenti genotipi della β-Lattoglobulina. pp 429-430 in Proc. 51st Congr. S.I.S.Vet., Bologna, Italy.
- GIACCONE, P., DI STASIO, L., MACCIOTTA, N.P.P., PORTOLANO, B., TODARO, M., CAPPIO-BORLINO, A., 2000. Effect of β-lactoglobulin polymorphism on related traits of dairy ewes' milk analysed by a repeated measures design. J. Dairy Res. 67:443-448.
- Greppi, G.F., Roncada, P., 2001. Tipicità del latte e formaggi in relazione al territorio: la filiera del latte di capra. I. La componente proteica. Proc. 36th Simp. Int. Zootec., ©MG Editori, Milano, Italy, pp 59-84.
- HAKKAK, R., KOROURIAN, S., SHELNUTT, S.R., LENSING, S., RONIS, M.J., BADGER, T.M., 2000. Diets containing whey proteins or soy protein isolate protect against 7,12-dimethylbenz(a)anthracene-induced mammary tumors in female rats. Cancer Epidemiol. Biomarkers Prev. 9:113-117.
- HERGET, I., SCHMOLL, F., BREM, G., SCHELLANDER, K., 1995. β-Lactoglobulin polymorphism in East Friesian milk sheep. Page 21 in Proc. 46th EAAP Meet., Wagenigen Pers, Wagenigen, The Netherlands.
- LAW, A.J.R., 1995. Heat denaturation of bovine, caprine and ovine whey proteins. Milchwissenschaft 50:384-388.
- MACKLE, T.R., BRYANT, A.M., PETCH, S.F., HILL, J.P., AULDIST, M.J., 1999. Nutritional influences on the composition of milk from cows of different protein phenotypes in New Zealand. J. Dairy Sci. 82:172-180.
- MISTRY, V.V., MAUBOIS, J.L., 1993. Application of membrane separation technology to cheese production.
 In: P.F. Fox (ed.) Cheese: chemistry, physics and microbiology. Chapman & Hall London, UK, vol.1, pp 493-522.
- Neville, M.C., Zhang, P., 2000. Lactoferrin secretion into milk: comparison between ruminant, murine, and human milk. J. Anim. Sci. 78:26-35.

- NG-KWAI-HANG, K.F., 1997. A review of the relationship between milk protein polymorphism and milk composition/milk production. Proc. IDF Seminar Milk Protein Polymorphism. Publ. IDF, Brussels, Belgium, pp 22-37.
- PHILANTO-LEPPALA, A., KOSKINEN, P., PIILOLA, K., TUPASELA, T., KORHONEN, H., 2000. Angiotensin Iconverting enzyme inhibitory properties of whey protein digests: concentration and characterisation of active peptides. J. Dairy Res. 67:53-64.
- PROSSER, C.G., McLAREN, R.D., 1997. Effect of atropine on milk protein yield by dairy cows with different β-lactoglobulin phenotypes. J. Dairy Sci. 80:1281-1287
- Pulina, G., Furesi, R., 2001. L'allevamento e la produzione di latte ovino in Italia. Sci. Tec. Latt.-cas., 52:209-216
- PULINA, G., SERRA, A., MACCIOTTA, N.P., NUDDA, A., 1993 La produzione continua di latte nella specie ovina in ambiente mediterraneo. pp 353-356 in Proc. 10th Congr. A.S.P.A., Bologna, Italy.
- QIAN, Z.Y., JOLLÈS, P., MIGLIORE-SAMOUR, D., FIAT, A.M., 1995. Isolation and characterization of sheep lactoferrin, an inhibitor of platelet aggregation and composition with human lactoferrin. Biochim. Biophys. Acta 1243: 25-32.
- QUILES, A., GONZALO, C., BARCINA, Y., FUENTES, F., HEVIA, M., 1994. Protein quality of Spanish Murciano-Granadina goat milk during lactation. Small Rum. Res. 14:67-72.

- RAMPILLI, M., CECCHI, F., GIULIOTTI, L., CATTANEO, T.M.P., 1997. The influence of β-lactoglobulin genetic polymorphism on protein distribution and coagulation properties in milk of Massese breed ewes. Proc. IDF Seminar Milk Protein Polymorphism. Publ. IDF, Brussels, Belgium, pp 311-315.
- SANCHEZ, L., LUIJAN, L., ORIA, R., CASTILLO, H., PEREZ, D., ENA, J.M., CALVO, M., 1992. Synthesis of lactoferrin and transport of transferrin in the lactating mammary gland of sheep. J. Dairy Sci. 75:1257-1262.
- SANCHEZ, L., ARANDA, P., PÉREZ, M.D., CALVO, M., 1988. Concentration of lactoferrin and transferrin throughout lactation in cow's colostrum and milk. Biol. Chem. Hoppe Seyler 369:1005-1008.
- SCHANBACHER, F.L., TALHOUK, R.S., MURRAY, F.A., 1997. Biology and origin of bioactive peptides in milk. Livest. Prod. Sci. 50:105-123.
- SORDILLO, L.M., SHAFER-WEAVER, K., DEROSA, D., 1997. Immunobiology of the mammary gland. J. Dairy Sci. 80:1851-1865.
- TAIBI, L., PILLA, F., TRIPALDI, C., ANNICCHIARICO, G., BEVILACQUA, C., NAPOLITANO, F., DELL'AQUILA, S., 1999. Influenza del tipo genetico e del genotipo al locus β-Lattoglobulina sulla produzione e sulle qualità chimico-fisiche e tecnologiche del latte di pecora. Zoot. Nutr. Anim. 25:187-193.
- Urech, E., Puhan, Z., Schallbaum, M., 1999. Changes in milk protein fraction as affected by subclinical mastitis. J. Dairy Sci. 82:2402-2411.