



## Short communication: In vitro rumen gas production and starch degradation of starch-based feeds depend on mean particle size

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### ABSTRACT

Our objective was to model the effect of mean particle size (mPS) on in vitro rumen starch degradation (IVSD) and the kinetics of gas production for different starch-based feeds. For each feed, 2 batches of the same grains were separately processed through 2 different mills (cutter or rotor speed mills), with or without different screens to achieve a wide range of mPS (0.32 to 3.31 mm for corn meals; 0.19 to 2.81 mm for barley meals; 0.16 to 2.13 mm for wheat meals; 0.28 to 2.32 mm for oat meals; 0.21 to 2.36 mm for rye meals; 0.40 to 1.79 for sorghum meals; 0.26 to 4.71 mm for pea meals; and 0.25 to 4.53 mm for faba meals). The IVSD data and gas production kinetics, obtained by fitting to a single-pool exponential model, were analyzed using a completely randomized design, in which the main tested effect was mPS ( $n = 6$  for all tested meals, except  $n = 7$  for corn meals and  $n = 5$  for sorghum meals). Rumen inocula were collected from 2 fistulated Holstein dairy cows that were fed a total mixed ration consisting of 16.2% crude protein, 28.5% starch, and 35.0% neutral detergent fiber on a dry matter basis. The IVSD, evaluated after 7 h of rumen incubation, decreased linearly with increasing mPS for corn, barley, wheat, rye, pea, and faba meals, and decreased quadratically with increasing mPS for the other meals. The y-axis intercept for 7-h IVSD was below 90% starch for corn, barley, and rye feeds and greater than 90% for the other tested feeds. The mPS adjustment factors for the rate of rumen starch degradation varied widely among the different tested feeds. We found a linear decrease in starch degradation with increasing mPS for barley, wheat, rye, and pea meals, whereas we noted a quadratic decrease in starch degradation for the other tested meals. Further, we observed a linear decrease in the rate of gas production with increasing mPS in each

tested feed, except for pea meal, which had a quadratic relationship. For each 1 mm increase in mPS, the gas production was adjusted by  $-0.009 \text{ h}^{-1}$  for corn,  $-0.011 \text{ h}^{-1}$  for barley,  $-0.008 \text{ h}^{-1}$  for wheat, and  $-0.006 \text{ h}^{-1}$  for faba, whereas numerically greater adjustments were needed for oat ( $-0.022 \text{ h}^{-1}$ ), rye ( $-0.017 \text{ h}^{-1}$ ), and sorghum ( $-0.014 \text{ h}^{-1}$ ). These mPS adjustment factors could be used to modify the starch-based feed energy values as a function of mean particle size, although in vivo validation is required.

**Key words:** in vitro method, processing, fermentation kinetics, nutritional model

### Short Communication

Farmers typically give high-energy diets to lactating dairy cows. However, diets that are rapidly fermented in the rumen can lead to the rapid production of VFA. If the production of these acids exceeds the ability of the rumen to neutralize and absorb them, SARA can occur, thus worsening microbial fermentation, rumen epithelial function, animal health, and milk production (Silveira et al., 2007; Penner et al., 2009). Therefore, many developers of ruminant nutrition models have recently focused on prediction of starch digestion dynamics in the digestive tracts of dairy cows (Higgs et al., 2015; Bannink et al., 2016; Ghimire et al., 2017). Two recent meta-analyses (Patton et al., 2012; Moharrery et al., 2014) aimed to determine the amount of starch digested in the different compartments of the gastrointestinal tract and to identify the main factors affecting starch digestion dynamics. In both cases, the proposed starch digestion submodels were mainly based on starch sources and starch intake levels. Nonetheless, it was difficult to include certain other factors in these models. Thus, Patton et al. (2012) declared "...inaccuracies in prediction of starch degradability in the rumen may be mainly due to processing effects and particle sizes, but these were not well reported in literature and were difficult to estimate," and Moharrery et al. (2014) stated "...effects of physical structure and heat treatment were initially tested, however data balance

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did not allow for conclusive statements for the present dataset and approach.”

Previous studies suggested that the mean particle size (**mPS**) of feeds can affect digestion rate (Hoffman et al., 2012; Zhao et al., 2015; Tagawa et al., 2017), and that differences in the passage rates of large, medium, and small particles exist within the different gastrointestinal compartments of ruminant animals (Nocek and Tamminga, 1991; Offner and Sauvant, 2004; Ferraretto et al., 2013). We previously modeled the effects of mPS on in vitro rumen starch degradation (**IVSD**) and OM fermentability (Gallo et al., 2016) by examination of a wide range of mPS, with meals consisting of dry whole-kernel corn ( $n = 11$  mPS, 0.46 to 3.50 mm) and dry hulless whole-kernel barley ( $n = 10$  mPS, 0.11 to 2.98 mm), by use of well-established rumen-based in vitro methods. We proposed the use of mPS adjustment factors (i.e., the slopes of regression terms in which mPS represented the independent variable) to characterize the rate of rumen starch degradation (**kd starch**) and the amount of starch degradation after 7 h of in vitro rumen incubation (**7hIVSD**). In particular, the previous study indicated that for each 1-mm increase in mPS, corn feed had a linear decrease of  $0.049 \text{ h}^{-1}$  in the kd starch and a 6.3 percentage units decrease in the 7hIVSD, and barley feed had a linear decrease of  $0.092 \text{ h}^{-1}$  in the kd starch and a 6.5 percentage units decrease in the 7hIVSD.

The purpose of the current study was to extend the previous approach to other starch-based feeds, with the aim to model the effect of mPS on IVSD data and the kinetics of gas production. In particular, we selected the same raw starch-based feeds examined in the previous meta-analysis of Moharrery et al. (2014).

Two 5-kg batches of dry whole corn, dry hulled barley, dry wheat, dry oat, dry rye, and dry sorghum kernels were collected over 2 wk (1 batch each week) from the same feedstock grains stored in grain storage bins at local industrial feed mills. Two batches of dry whole pea (*Pisum sativum*) and faba (*Vicia faba* var. minor) beans were generously donated by the Centro Ricerche Produzioni Animali S.p.A. (CRPA, Reggio Emilia, Italy).

To obtain different mPS, subsamples of about 1.3 to 1.5 kg for each batch were processed as described by Hoffman et al. (2012), in which a cutter mill (Pulviresette 19, Fritsch, Idar-Oberstein, Germany) that was fitted with 4.0-, 3.0-, 2.0-, or 1.0-mm screens (1 passage) or without screens (1 to 5 passages) was used. Samples were also passed through a rotor speed mill Pulverisette 14 (Fritsch) that was equipped with 1.0- and 0.5-mm screens (1 passage; Table 1). Afterward, a representative amount (100 g) of the various grinds was run for 10 min through a sieve shaker (Multidimensional

Sieveshaker IG/1/S, Giuliani Tecnologie s.r.l., Torino, Italy) that had 9 different screen sieves with nominal aperture sizes of 4.00, 3.50, 2.50, 1.50, 1.00, 0.75, 0.50, 0.25, and 0.125 mm, followed by a pan. The mPS was measured using equation 1 of ASAE S319.3 method, as reported in ASABE (2006). In particular, the mPS of each material retained on a sieve was calculated on a weight basis as the geometric mean of the diameter of the openings in the 2 adjacent sieves in the stack (Pfost and Headley, 1976). A portion of each subsample that was ground through a screen of over 0.50 mm was re-ground by the rotor speed mill equipped with 0.50-mm screen and analyzed for total starch (Megazyme assay kit K-TSTA 07/11), ash (AOAC International, 2000; method 942.05), and CP (AOAC International, 2000; method 984.13).

The IVSD was evaluated by an in vitro rumen-based method, which was slightly modified from the method of Sveinbjörnsson et al. (2007). Rumen fluid was collected from 2 fistulated dairy cows that received a TMR (16.2% CP, 28.5% starch, and 35.0% NDF on a DM basis), formulated according to the NRC (2001) for an average BW of 600 kg, 140 DIM, and 35 kg of milk yield (3.75% fat and 3.35% protein). The diet consisted of corn silage, energy-protein supplement, alfalfa, and grass hays (31.2, 48.0, 16.7, and 4.1% DM, respectively). Rumen liquor was maintained in a warm insulated flask, filtered through 2 layers of cheesecloth, and used within 20 min of collection. Samples containing 250 mg of starch were weighed in 125-mL glass bottles (Wheaton borosilicate glass serum bottle; 54 mm diameter  $\times$  107 mm height; Z114014; Sigma-Aldrich Co., Milan, Italy), which were filled with 30 mL of the diluted rumen fluid (buffer-to-rumen ratio 2:1, vol/vol), gassed with  $\text{CO}_2$ , closed with rubber stoppers (gray butyl stoppers; 20-mm diameter; 27232; Sigma-Aldrich Co.), and then incubated at  $39^\circ\text{C}$  in a shaking water bath (50 rpm). Blank samples (diluted rumen fluid only) and an internal standard (Gelose 80 maize starch; Penford Food Ingredients Co., Englewood, CO) were also included. After different incubation times, bottles were plunged into a bath containing ice to stop starch degradation. Residual starch was quantified using a 2-step enzymatic approach detailed previously (Gallo et al., 2016).

The IVSD after 7 or 120 h of rumen incubation was calculated as

$$\text{IVSD, \% starch} = [1 - (\text{resStarch} - \text{blnStarch}) / \text{incStarch}] \times 100\%, \quad [1]$$

where resStarch is the amount of residual starch after 7 or 120 h of rumen incubation; blnStarch is the blank

**Table 1.** Mean values<sup>1</sup> of starch content and mean particle size (mPS<sup>2</sup>) of starch-based feed processed through 2 different grinding mills equipped without or with different screens

Item	Screen (mm)	Transits (No.)	Starch-based feed							
			Corn meal	Barley meal	Wheat meal	Oat meal	Rye meal	Sorghum meal	Pea meal	Faba meal
Starch (% DM)			71.2	52.9	66.5	59.1	55.8	72.5	52.9	45.1
Ash (% DM)			1.5	3.2	2.2	3.8	2.1	1.9	3.3	4.0
CP (% DM)			8.6	11.1	12.1	13.4	9.3	11.1	23.0	31.4
mPS after milling <sup>3</sup> (mm)										
Rotor speed	0.5	1	0.32	0.19	0.16	0.28	0.21	0.40	0.26	0.25
Rotor speed	1.0	1	—	—	—	—	0.31	—	—	—
Cutter mill	1.0	1	0.62	0.36	0.54	—	0.67	0.79	—	—
Cutter mill	2.0	1	0.92	0.94	0.79	0.77	0.99	1.04	0.84	0.79
Cutter mill	3.0	1	1.45	—	1.39	—	—	—	—	—
Cutter mill	4.0	1	—	1.55	1.52	1.26	1.78	1.42	1.58	1.33
Cutter mill	—	5	2.12	2.08	—	1.52	—	—	—	2.35
Cutter mill	—	4	—	—	—	2.12	—	1.79	2.65	—
Cutter mill	—	3	—	—	—	—	—	—	2.91	3.40
Cutter mill	—	2	2.57	—	—	—	—	—	—	—
Cutter mill	—	1	3.31	2.81	2.13	2.32	2.36	—	4.17	4.53

<sup>1</sup>Values are means of 2 replicates, 1 for each batch.

<sup>2</sup>Calculated using a sieve shaker (Multidimensional Sieveshaker IG/1/S, Giuliani Tecnologie s.r.l., Torino, Italy) equipped with screen sieves with nominal openings of 4.00, 3.50, 2.50, 1.50, 0.75, 0.50, 0.25, and 0.125 mm, followed by a pan. The mPS was measured applying equation number 1, as reported in the ASABE (2006), ASAE S319.3 method.

<sup>3</sup>Cutter mill Pulviresette 19 and rotor speed mill Pulverisette 14 (Fritsch, Idar-Oberstein, Germany).

correction at the corresponding incubation time; and incStarch is the amount of starch in the sample before incubation. The resStarch and blnStarch values were calculated from the glucose concentration and dilutions, and by converting free glucose to starch using a fixed factor of 0.9. All terms are expressed in grams and 3 bottles were used for each incubation time. The kd starch (per hour) was calculated from the IVSD data after 7 h of rumen incubation, using a first-order model with a fixed discrete lag time of 0.5 h. A fixed indigestible starch fraction (iStarch) of 0.5% starch was used because the 120-h IVSD values were greater than 98 to 99% starch in all analyzed samples. Thus, the kinetic model (Bender et al., 2016) was

$$R(t) = (100 - \text{iStarch}) \times [1 - e^{\text{kd starch} \times (t - \text{Lag})}], [2]$$

where  $R(t)$  is the percentage of rumen starch degraded at time  $t$ ; kd starch is the kinetic constant of rumen starch degradation (per hour);  $t$  is time (h); iStarch is the percentage of rumen indigestible starch; and Lag is the lag time (0.5 h).

To measure rumen fermentability (gas production; Menke and Steingass, 1988), samples were incubated in the same diluted rumen fluid (buffer-to-rumen ratio 2:1, vol/vol) used in the IVSD experiments. Briefly, about 220 mg of each feed sample was weighed in graduated 100-mL glass gas-tight syringes equipped with a piston (Sigma-Aldrich Co.) and then 30 mL of diluted rumen fluid was added. Before injection into the syringes, the

medium was saturated with CO<sub>2</sub> and the pH corrected to 6.5 to 6.6. All gas was then expelled from the syringe, after which the lower end was closed. Syringes with samples, blanks, and an internal standard were placed vertically in a water bath at 39°C, and gas production was measured at 1, 2, 4, 6, 8, 12, 24, 48, 72, 96, and 120 h of incubation. Syringes were manually agitated at every measurement.

The kinetics of gas production (**kd**) was computed by a single-pool exponential model (Wang et al., 2013):

$$V(t) = V_f \times [1 - e^{-\text{kd} \times (t - \text{Lag})}], [3]$$

where  $V(t)$  is the volume of gas accumulated (mL/g of OM) at time  $t$ ;  $V_f$  is the final gas volume (mL/g OM); kd is the kinetic constant (per hour);  $t$  is time (h), and Lag is the lag time (h). For both in vitro tests, samples were incubated in triplicate in 2 separate runs. Samples within the run were considered analytical repetitions, and samples between runs as experimental replicates.

The IVSD data and kinetics of gas production were evaluated using a completely randomized design with the MIXED procedure of SAS Institute (2003). Each tested starch-based feed was separately analyzed using the model

$$Y_{ij} = \mu + \text{mPS}_i + e_{ij},$$

where  $Y_{ij}$  is the response variable;  $\mu$  is the overall mean;  $\text{mPS}_i$  is the fixed effect of mPS for a specific feed ( $i =$

7 for corn meal,  $i = 6$  for barley, wheat, oat, rye, pea, and faba meals, and  $i = 5$  for sorghum meal); and  $e_{ij}$  is the random residual error. The REG proc of SAS Institute (2003) was used for each specific feed to verify relationships between mPS (independent variable) and the parameters of interest (7hIVSD, kd starch, kd, and lag). Before regression analysis, all experimental replicates were averaged. A regression term (y-axis intercept, linear and quadratic terms) was considered significant at  $P < 0.05$ . When the quadratic effect was not significant, it was removed by regression model.

Several feed evaluation systems (NRC, 2001; NorFor, 2011; Higgs et al., 2015) take into account the effect of mPS of meals entering into dairy cow diets to evaluate their energy content. However, the proposed adjustment factors are categorical, being related to feed categories or distinguishing meals into broad mPS classes (e.g., coarse-, medium-, or fine-ground feed grains). For instance, NRC (2001) used an empirical approach based on a processing adjustment factor to adapt the truly digestibility of the NFC fraction to the physical form of meals. The processing adjustment factor was 1.00 for ground corn, 0.95 for cracked dry corn, and 1.04 for all other cereal meals or rolled grains.

The adopted procedure of processing the different starch-based feeds led to a wide range in mPS (Table 1). In particular, the mPS of samples ranged from very small mPS values (i.e.,  $<0.40$  mm) to values greater than 2.00 mm, except for sorghum meals (i.e., 1.79 mm). The starch, ash, and CP of the starch-based feeds were in accordance with their expected chemical compositions and did not numerically differ among meals varying in mPS within the same starch-based feed.

Tables 2 and 3 shows the results of the IVSD and gas production experiments. Overall, the 7hIVSD and kd starch decreased ( $P < 0.05$ ) as the mPS increased, in agreement with previous studies performed in vitro (Hoffman et al., 2012; Tagawa et al., 2017) and in vivo (San Emeterio et al., 2000; Callison et al., 2001). In particular, the 7hIVSD values decreased linearly with mPS ( $P < 0.05$ ) in corn, barley, wheat, rye, pea, and faba feeds, and quadratically ( $P < 0.05$ ) for the other feeds (Figure 1). The 7hIVSD y-axis intercepts were below 90% starch for corn, barley, and rye feeds, but greater than 90% starch for the other feeds. Analysis of the kd starch indicated linear decreases ( $P < 0.05$ ) with mPS for barley, wheat, rye, and pea feeds, and quadratic decreases ( $P < 0.05$ ) for the other feeds. Furthermore, the y-axis intercepts were low for corn ( $0.335 \text{ h}^{-1}$ ), barley ( $0.289 \text{ h}^{-1}$ ), and rye ( $0.296 \text{ h}^{-1}$ ); moderate for wheat ( $0.394 \text{ h}^{-1}$ ) and faba ( $0.399 \text{ h}^{-1}$ ); and high for oats ( $0.458 \text{ h}^{-1}$ ), sorghum ( $0.443 \text{ h}^{-1}$ ), and pea ( $0.470 \text{ h}^{-1}$ ). Consequently, the mPS adjustment factors differed for the different feeds. The mPS can influence the

surface area of the starch granules, and thereby affect bacterial attachment and starch degradation (Hoffman et al., 2012; Giuberti et al., 2014). Thus, the present findings can be explained as resulting from interactions of several physicochemical properties of the selected starch feeds. These include the chemical composition of the feed, starch granule morphology, amylose-to-amylopectin ratio, presence of a protein matrix encapsulating the starch granules, and adaptation of rumen bacteria to different starch sources (Svihus et al., 2005; Lanzas et al., 2007; Giuberti et al., 2014).

Some criticisms could be raised concerning the methods we used to calculate kd starch. In particular, the kd starch calculation was computed using 2 fixed points common to all tested starch meals (0% starch or 99.5% starch for rumen incubations of 0 or 120 h) and by adopting a fixed lag time of 0.5 h. Consequently, the only factor determining rate was the 7hIVSD, thus representing the simplest approach to get a rate. However, the development of a starch degradability profile by adopting IVSD methods with a greater number of incubation times is difficult, as previously reported (Cone, 1991; Giuberti et al., 2014; Gallo et al., 2016). In particular, the IVSD assay is more labor-intensive and has lower reproducibility than other in vitro approaches. Thus, optimization of IVSD methods is a topic that needs further investigation.

The gas production technique has been widely used to evaluate the nutritive value of feeds (Getachew et al., 1998). In particular, gas production data, in combination with chemical analysis, can predict in vivo OM digestibility, ME, and rumen protein degradability of several feedstuffs. However, the use of gas production parameters often fails to properly explain the extent of degradation of specific nutrients in the rumen or animal performance, thus limiting its application in nutrition models (France et al., 2000; Chai et al., 2004; Lanzas et al., 2007). The kinetic parameters of current gas production data are in line with previous results (Gallo et al., 2016). In particular, we found a linear decrease in kd ( $P < 0.05$ ) with increasing mPS for each tested feed except for pea meals, which was better described by a quadratic fit ( $P < 0.05$ ; Figure 2). The y-intercepts for kd ranged between 0.047 (faba) and  $0.078 \text{ h}^{-1}$  (oat). For each 1-mm increase in mPS, the kd declined by  $0.009 \text{ h}^{-1}$  for corn,  $0.011 \text{ h}^{-1}$  for barley,  $0.008 \text{ h}^{-1}$  for wheat, and  $0.006 \text{ h}^{-1}$  for faba. Thus, the mPS adjustment factors for kd as a function of mPS were numerically different for oat ( $-0.022 \text{ h}^{-1}$ ), rye ( $-0.017 \text{ h}^{-1}$ ), and sorghum ( $-0.014 \text{ h}^{-1}$ ). Furthermore, the lag increased linearly ( $P < 0.05$ ) with increasing mPS for barley, wheat, oat, sorghum, and faba meals ( $P < 0.05$ ) and quadratically for corn and pea ( $P < 0.05$ ).



**Table 2.** In vitro rumen starch degradation data (7hIVSD = in vitro rumen starch degradation evaluated after 7 h of rumen incubation, % starch; kd starch = rate of rumen starch degradation per hour) and gas production kinetic parameters (kd = rate of gas production per hour; lag = lag time in hours; Vf = final gas volume, mL/g of OM) of processed corn, barley, wheat, and oat feed meals differing in mean particle size (mPS)

Rank of mPS <sup>1</sup>	Corn meals				Barley meals				Wheat meals				Oat meals					
	Starch degradation		Gas production		Starch degradation		Gas production		Starch degradation		Gas production		Starch degradation		Gas production			
	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf		
Meal 1	86.3	0.309	0.054	1.5	328.6	86.8	0.313	0.050	0.0	328.1	92.9	0.421	0.057	0.3	316.4	0.067	0.7	300.0
Meal 2	84.9	0.290	0.047	1.8	323.4	81.2	0.256	0.055	0.1	291.9	90.7	0.370	0.047	0.2	347.5	0.064	0.6	305.9
Meal 3	78.6	0.236	0.044	2.2	333.3	80.4	0.250	0.049	0.8	307.5	78.2	0.234	0.046	0.9	308.3	0.051	0.3	338.2
Meal 4	75.9	0.218	0.038	2.8	337.7	74.9	0.213	0.043	1.4	318.1	79.6	0.243	0.043	1.4	313.7	0.054	0.7	309.7
Meal 5	75.7	0.217	0.035	2.9	323.5	73.8	0.206	0.035	1.3	323.8	79.9	0.249	0.046	1.3	309.9	0.039	0.8	290.5
Meal 6	72.6	0.200	0.024	3.3	345.6	72.5	0.198	0.021	1.6	274.9	76.7	0.223	0.037	1.8	341.7	0.017	1.9	297.7
Meal 7	72.0	0.195	0.029	3.0	325.3	—	—	—	—	—	—	—	—	—	—	—	—	—
SEM	1.45	0.0144	0.0015	0.19	5.17	1.47	0.0129	0.0006	0.17	6.81	2.29	0.0359	0.0007	0.15	8.23	0.0015	0.11	5.12
P-value	0.001	0.004	<0.001	0.002	0.113	0.003	0.005	<0.001	<0.001	0.001	0.009	0.031	<0.001	0.002	0.051	<0.001	0.001	0.006

<sup>1</sup>Within each tested starch-based feed meal, the rank referred to the increasing values of mPS, as detailed in Table 1. The range in mPS (mm) was 0.32 (meal 1) to 3.31 (meal 7) for corn, 0.19 (meal 1) to 2.81 (meal 6) for barley, 0.16 (meal 1) to 2.13 (meal 6) for wheat, and 0.28 (meal 1) to 2.32 (meal 6) for oat.

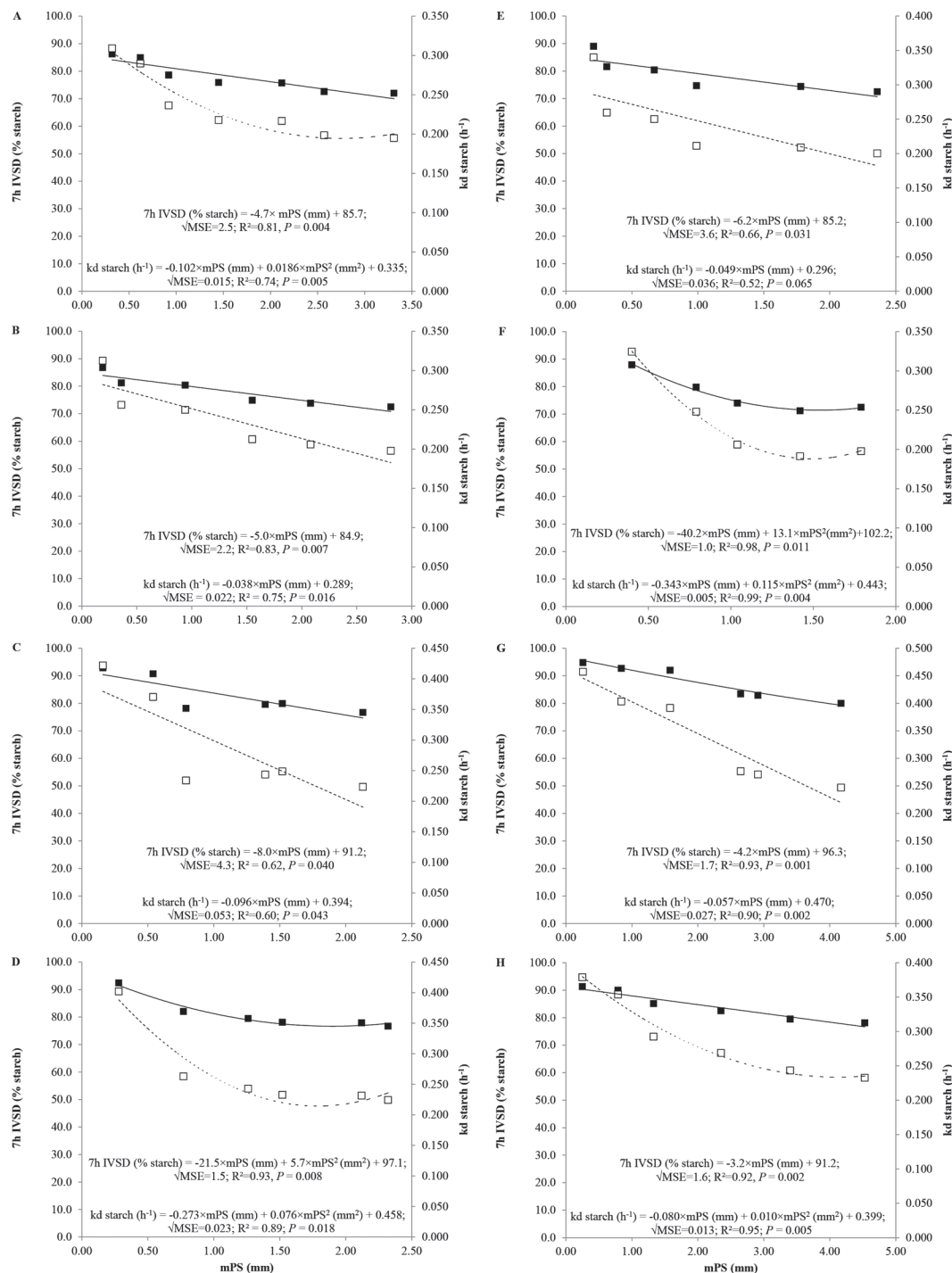
**Table 3.** In vitro rumen starch degradation data (7hIVSD = in vitro rumen starch degradation evaluated after 7 h of rumen incubation, % starch; kd starch = rate of rumen starch degradation per hour) and gas production kinetic parameters (kd = rate of gas production per hour; lag = lag time in hours; Vf = final gas volume, mL/g of OM) of processed rye, sorghum, pea, and faba feed meals differing in mean particle size (mPS)

Rank of mPS <sup>1</sup>	Rye meals				Sorghum meals				Pea meals				Faba meals								
	Starch degradation		Gas production		Starch degradation		Gas production		Starch degradation		Gas production		Starch degradation		Gas production						
	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf	7hIVSD	kd	lag	Vf					
Meal 1	89.0	0.340	0.064	0.0	347.3	87.9	0.324	0.050	1.9	333.3	94.8	0.457	0.062	0.2	302.0	91.3	0.379	0.049	0.0	333.8	
Meal 2	81.6	0.260	0.065	0.0	328.3	79.8	0.248	0.035	2.3	345.6	92.7	0.403	0.051	1.0	306.2	90.0	0.354	0.039	0.0	316.4	
Meal 3	80.4	0.250	0.046	0.0	410.1	74.0	0.206	0.040	2.1	331.9	92.0	0.391	0.043	1.5	336.2	85.1	0.292	0.041	0.5	276.8	
Meal 4	74.7	0.211	0.051	0.0	357.4	71.2	0.191	0.034	3.0	333.3	83.4	0.276	0.034	2.7	325.6	82.5	0.270	0.033	1.7	313.2	
Meal 5	74.4	0.209	0.046	1.3	331.0	72.5	0.198	0.026	3.7	330.0	82.9	0.271	0.033	2.6	332.1	79.5	0.243	0.026	2.9	307.9	
Meal 6	72.5	0.200	0.020	0.7	309.1	—	—	—	—	—	80.0	0.247	0.028	2.5	338.5	78.1	0.233	0.023	3.3	311.9	
SEM	2.32	0.0163	0.0020	0.37	14.92	1.92	0.0151	0.0016	0.15	7.19	1.126	0.0205	0.0007	0.03	6.48	1.33	0.0163	0.0016	0.33	9.98	
P-value	0.017	0.007	<0.001	0.043	0.030	0.008	0.007	0.001	0.002	0.604	<0.001	<0.001	<0.001	<0.001	<0.001	0.026	0.002	0.003	0.001	0.001	0.080

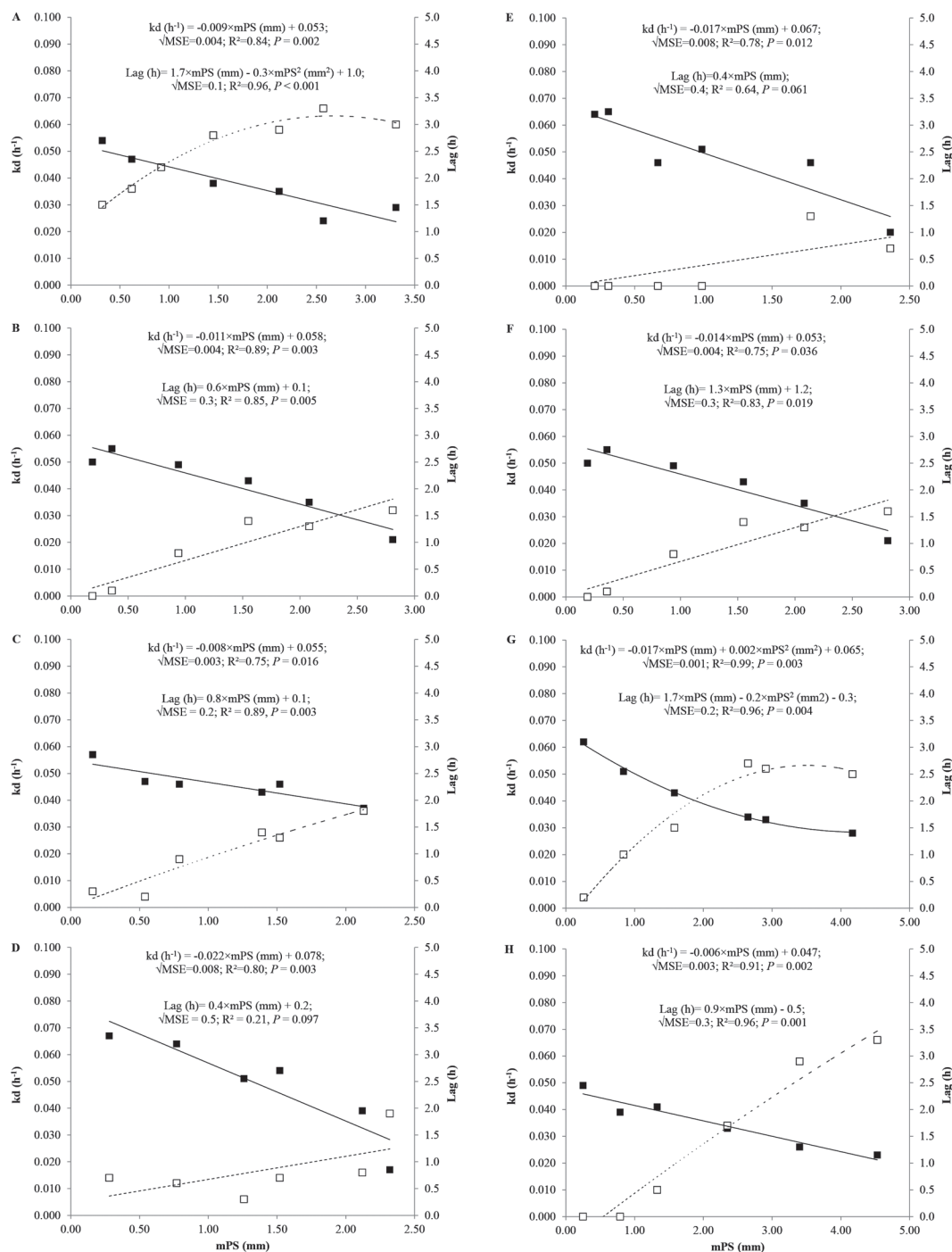
<sup>1</sup>Within each tested starch-based feed meal, the rank referred to the increasing values of mPS, as detailed in Table 1. The range in mPS (mm) was 0.21 (meal 1) to 2.36 (meal 6) for rye, 0.40 (meal 1) to 1.79 (meal 5) for sorghum, 0.26 (meal 1) to 4.17 (meal 6) for pea, and 0.25 (meal 1) to 4.53 (meal 6) for faba.

In the current study, we described gas production kinetics by fitting the data to a single pool exponential model. In a previous study (Gallo et al., 2016), we evaluated the opportunity to identify fast and slow

pools by adopting a 2-pool exponential model and a Gompertz model. We did so in an attempt to improve the dynamic feed evaluation models by incorporating specific kd for fast and slow digestible pools of carbohy-



**Figure 1.** In vitro starch degradation data of corn (A), barley (B), wheat (C), oat (D), rye (E), sorghum (F), pea (G), and faba (H) meals differing in mean particle size (mPS). 7h IVSD = in vitro starch degradation evaluated after 7 h of rumen incubation; kd starch = rate of starch degradation. Black or white squares refer to variables of interest displayed on left (primary) or right (secondary) sizes of Y-axes, respectively. MSE = mean squared error.



**Figure 2.** Fitted kinetic parameters of gas production of corn (A), barley (B), wheat (C), oat (D), rye (E), sorghum (F), pea (G), and faba (H) meals differing in mean particle size (mPS). kd = rate of gas production; Lag = lag time. Black or white squares refer to variables of interest displayed on left (primary) or right (secondary) sizes of Y-axes, respectively. MSE = mean squared error.

drate fractions (Zontini et al., 2015). The 2-pool exponential model provided inadequate characterization of the feeds, whereas the 2-pool sigmoidal model permitted identification of fast and slow digestible pools, but

only in corn, barley, wheat, oat, and rye meals (data not shown).

In conclusion, the data from current in vitro experiments permitted us to model the effect of mPS on IVSD

and the kinetics of gas production of diverse starch-based feeds. However, these results must be validated by controlled in vivo digestibility studies that evaluate the effect of different starch sources and particle sizes on starch digestion in the rumen.

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