



Relationship between different livestock managements and stress response in dairy ewes

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Abstract. The gradual diffusion of intensive and semi-intensive production systems, especially in dairy sheep breeds, has led to the growing concern of consumers about the life conditions of farmed animals. Space allowance and structures of sheep houses are described as the main potential sources of discomfort for housed flocks, together with inappropriate milking procedures and human–animal interactions. The aim of the present study was to evaluate whether the structure relative to milking room could represent a stressor in Sarda dairy ewes. Animals were divided into two groups according to their farm of origin. Group A ($n = 40$) was from a farm whose milking room was an old warehouse with a waiting area limited and located outdoors and at a different level with respect to the milking room. The passageway of the entrance in the milking room was narrow and perpendicular to the milking positioning so the animal must bend 90° to enter in the room. Group B ($n = 40$) was from a farm whose milking room was wide and modern with a large waiting area located at the same level. From all animals blood samples were collected at T_0 (2 h before milking procedure at 06:00), at T_1 (immediately after the animals entered the milking room, about 08:00) and at T_2 (after milking procedure). In addition plasma cortisol and glucose values were evaluated. Statistical analysis showed significant effect of milking room ($P < 0.001$) and of sampling time ($P < 0.05$) on cortisol and glucose levels. The results obtained in the present study suggest that, in addition to milking, the characteristics of the room where this procedure occurs represent stressful stimuli that could influence negatively the productivity and welfare of dairy ewes.

1 Introduction

Animal welfare has contributed to a better understanding of how animals perceive their social and physical environment, their motivations, and the physiological and behavioural needs, enabling the design of environments to better satisfy these needs (Webster, 2005). Several studies have addressed the definition of animal welfare and the most suitable indicators to its evaluation, particularly, in the production under intensive conditions, where animals frequently face situations implying behavioural, physiological and physical stress including restraint, handling or novelty, hunger, thirst, fatigue, injury or thermal extremes (Mason and Mendl, 1993; Blokhuis et al., 2003; Mason and Latham, 2004). Stress can be defined as a physiological response elicited when threat to

homeostasis is perceived. In response to stressors, the central nervous system of livestock (and other mammalian species) evokes physiological responses that ultimately result in activation of the hypothalamo–pituitary–adrenocortical (HPA) axis and the sympathoadrenal axis inducing endocrine and metabolic changes (Carcangiu et al., 2008). The responses of these major systems are presumed to have adaptive and homeostatic value during periods of stress. Reducing stress during handling will provide advantages of increasing productivity (Grandin, 1998). The severity of the reaction is governed by a number of factors including individual sensitivity, unfamiliarity to the stimulus and the relative intensity (Grandin, 1997). The stress of handling can be reduced by using well-maintained systems and the development of ap-

appropriate systems should be guided by the requirement to ensure high standards of animal welfare (Goddard et al., 2006). Dairy ewe production systems have traditionally been characterized by the utilization of extensive grazing areas (Boya-zoglu and Morand-Fehr, 2001), where spatial restriction may be considered small, or virtually non-existent. Nonetheless, the progressive transition from traditional to modern production systems has led to the use of more intensive husbandry procedures and higher productive efficiency in sheep (Shrestha, 2011). The transition to modern farming systems can impose severe restrictions on the space available for dairy sheep. The use of the milking machine belongs to the modern farming systems and it determines a reduction in the animals handling and, consequently, a decrease in stress (Yardimci et al., 2013). However, the milking rooms show some critical points that can be a source of stress. It has been demonstrated that stress during milking can substantially reduce milk yield through a central inhibition of oxytocin secretion and peripheral action of catecholamines (Rushen et al., 2001). Moreover, the behavioural response of the animal to stress can increase the risk of injury and reduce the efficiency of milking (Rushen et al., 2001). Therefore, it is important to identify and reduce emotional or physical stress of dairy animals during the milking process in order to increase their productivity and to maintain their health status.

In view of such considerations, the aim of the present study was to evaluate whether the structure relative to milking room could represent a stressor in dairy ewe by measuring plasma cortisol and glucose levels before and after milking procedure.

2 Material and methods

2.1 Farm conditions and animals

A total of 80 Sarda dairy ewes were selected from 2 farms (Farm A and B) of sheep with a total of 200 animals each located in Sardinia (Italy) at the same altitude (529 m above sea level) and environmental conditions. The farms were different for milking room characteristics. In Farm A the milking room was an old warehouse with a waiting area limited and located outdoors and at a different level with respect to the milking room. The passageway of the entrance in the milking room was narrow and perpendicular to the milking positioning so the animal must bend 90° to enter in the room. In Farm B the milking room was wide and modern with a large waiting area located at the same level. From both Farm A and Farm B, 40 ewes with a mean body weight of 41.5 ± 0.5 kg and age between 2 and 5 years old were selected constituting Group A and Group B, respectively.

All the animals in the study were clinically healthy with no evidence of disease and free from internal and external parasites. Their health status was evaluated based on rectal temperature, heart rate, respiratory rate, appetite, faecal consistency and hematologic profile. Fresh faecal samples were

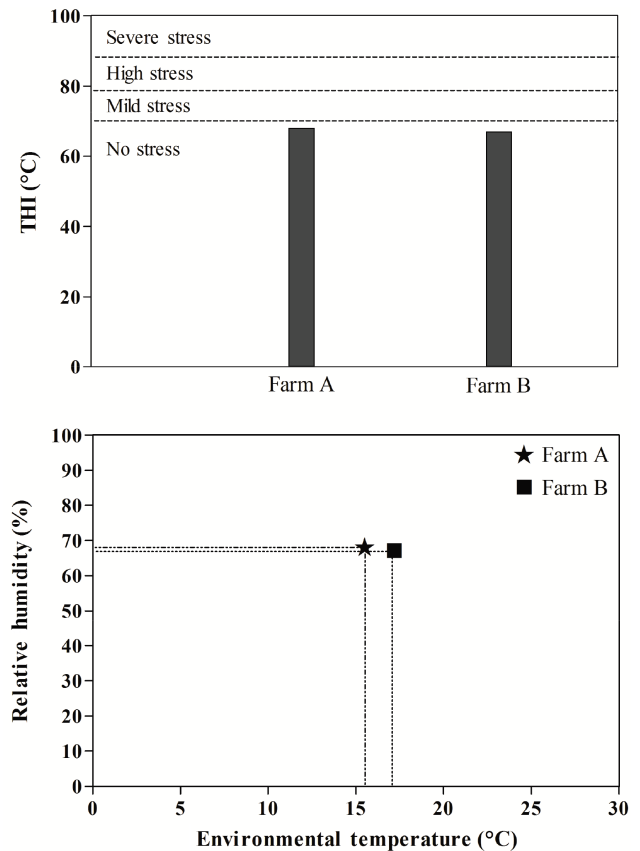


Figure 1. Temperature–humidity index (THI) values calculated for Farm A and Farm B with the respective environmental conditions.

examined according to the McMaster method based on protocols previously described by Maffa (1989). All the animals were kept in two indoor pens under natural photoperiod (sunrise at 05:54, sunset at 19:10 over the study period) and natural environmental temperature. Thermal and hygrometric records were carried out inside the box for the whole study by means of a data logger (Gemini, UK), and they followed the normal spring seasonal pattern for the location. The temperature–humidity index (THI) value, an indicator of thermal comfort, was calculated using the National Weather Service temperature humidity index formula for ruminant species (Potter and Jacobsen, 2000):

$$\text{THI (}^{\circ}\text{C)} = T^{\circ}\text{ ambient} + (0.36 \cdot \text{point of steam condensation}) + 41.5.$$

The temperature–humidity index (THI) values calculated for Farm A and Farm B with the respective climatic conditions are reported in Fig. 1.

All ewes were fed with a diet composed of a concentrate mixture which consisted of the following ingredients: oat 12 %, faba bean 15 %, barley 25 %, pea 10 %, sugar beet pulp 20 %, molasses 5 %, and mineral and vitamins supplements 3 %. Forage-based diets were alfalfa (*Medicago sativa* L.)

hay. About 250 g per animal of concentrate was distributed twice a day. Water was available ad libitum.

Lambs were weaned 35 days after birth. After weaning all ewes were milked twice daily with the milking machine. The animals were handled every day by the same operator.

All the animals were identified by the number plate of the local sanitary company and by a numbered collar applied to make the recognition of the subjects easier.

2.2 Blood sampling and laboratory analysis

From each animal of both farms blood sampling was performed by the same operator after approximately 20 days from the start of milking to ensure the adaptation of ewes to these procedures. Blood was sampled by jugular venipuncture into heparinized tubes (Becton Dickinson, Plymouth, UK) at T_0 (2 h before milking procedure at 06:00), at T_1 (immediately after the animals entered the milking room, about 08:00) and at T_2 (after the milking procedure).

The milking procedure lasted about 10 min. The blood was immediately centrifuged at 3000 rpm for 20 min at 4 °C, and the obtained plasma samples were stored at -20 °C until analysis. Plasma concentration of cortisol was measured with the Immulite 2000 (Siemens Healthcare Diagnostic, Deer field, IL, USA), which uses a solid-phase competitive enzyme-amplified chemiluminescent immunoassay. Plasma glucose concentration was evaluated using an enzymatic colorimetric method GOD-POD-PAP (Sentinel Chemical, Milan, Italy).

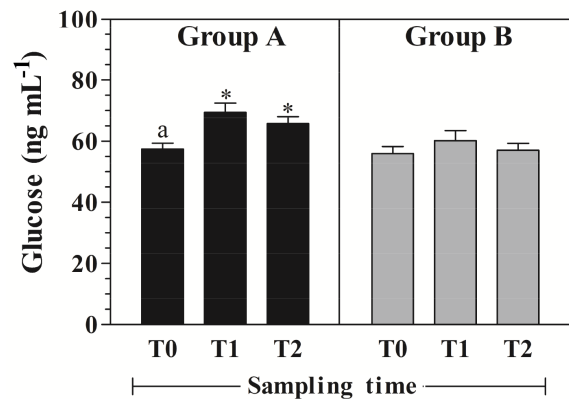
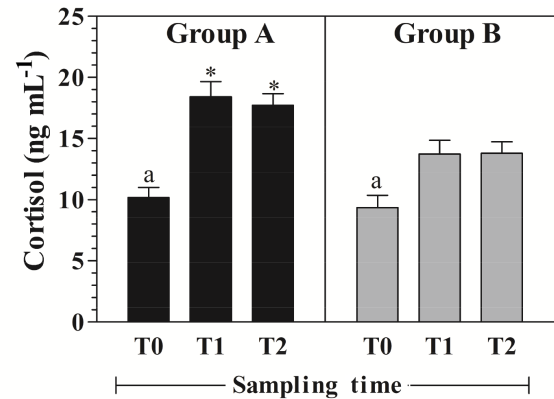
Protocols of animal husbandry and experimentation were reviewed and approved in accordance with the standards recommended by the Guide for the Care and Use of Laboratory Animals and Directive 2010/63/EU for animal experiments.

2.3 Statistical analysis

Data, expressed as mean values \pm standard deviation (SD), were tested for normality using the Shapiro–Wilk normality test. All data were normally distributed ($P > 0.05$) and the statistical analysis was performed. Two-way analysis of variance (ANOVA) for repeated measure was applied to assess significant effects of the experimental conditions (milking room and time) on plasma cortisol and glucose levels. When significant differences were found, Bonferroni's post hoc comparison was applied. P values < 0.05 were considered statistically significant. Statistical analysis was performed using the STATISTICA software package (STATISTICA 7 Stat Software Inc., Tulsa, Oklahoma).

3 Results

Statistical analysis showed a significant effect of milking room ($P < 0.001$) on plasma cortisol and glucose levels. As shown in Fig. 2, Group A showed higher cortisol and glucose concentration with respect to Group B at T_1 and T_2 .



Significant effect of milking room: * vs. Group B at T1 and T2 ($P < 0.001$)
Significant effect of sampling time: ^a vs. T1 and T2 ($P < 0.05$)

Figure 2. Mean values \pm standard deviation (\pm SD) of plasma cortisol and glucose obtained from dairy ewes (Group A and Group B) at T_0 (2 h before milking procedure at 06:00), at T_1 (immediately after the animals entered the milking room, about 08:00) and at T_2 (after milking procedure).

A significant effect of sampling time was found on studied parameters. In particular, higher plasma cortisol values were found at T_1 and T_2 with respect to T_0 in Groups A and B ($P < 0.05$). Plasma glucose showed increased values ($P < 0.05$) at T_1 and T_2 with respect to T_0 in Group A (Fig. 2).

4 Discussion

Domestic animals are routinely exposed to a variety of anthropogenic stressors. Milking management represents a critical point in sheep farm. The time animals need to adapt to machine milking, pre-parturition training to milking parlour, and type of milking (i.e. hand or machine milking), can markedly affect the welfare, health and production performance of dairy sheep. Although the use of the milking machine belonging to the modern farming systems determines

a reduction in the animal handling and, consequently, a decrease in stress (Yardimci et al., 2013), the milking rooms show some critical points that can be a source of stress. Under stressful stimuli the body must find a new dynamic equilibrium, and this requires several adaptive body responses.

It is well established that season, ambient temperature, as well as THI, and the other climatic conditions affect physiological, biochemical and hematological parameters in sheep (Piccione et al., 2011, 2012). THI values of 70 or less are considered comfortable, 75–78 stressful, and values greater than 78 cause extreme distress (Di Grigoli et al., 2009). The two farms considered in the present study showed very similar values of ambient temperature, relative humidity and THI. THI calculated for Farm A and Farm B was 61.78 and 63.72 °C, respectively, and were within the thermoneutral zone reported for the sheep (Nikitchenco et al., 1998; Di Grigoli et al., 2009). This excludes the influence of climatic conditions on studied parameters.

The results obtained in this study seem to suggest that milking room characteristics influence welfare of dairy ewes. Animals belonging to Farm A showed statistically significant higher plasma cortisol and glucose levels in comparison to animals of Farm B immediately after the animals entered the milking room (T_1) and after the milking procedure (T_2). Contrary to Farm B, whose milking room was wide and modern with a large waiting area located at the same level, milking room of Farm A was an old warehouse with a limited waiting area. Moreover, the animals of Farm A are forced in small bunches into tight spaces before entering the milking position, and they had to curve a rigid 90° to move to the milking location. All these factors could represent stress stimuli for dairy animals. Effectively, change in the basal level of cortisol in response to short-term stress can be considered an important indicator of animal welfare. Several studies carried out on sheep showed that some management activities, including isolation, artificial milk feeding and weaning, represent stressful conditions leading to an increase in the blood cortisol levels (Carcangiu et al., 2008; Napolitano et al., 1995, 2003; Orgeur et al., 1998). Animals of both considered groups showed statistically significant higher cortisol values at T_1 and T_2 with respect to T_0 probably due to milking procedures. Effectively, it has been suggested that machine milking is associated with a peak in plasma cortisol in lactating animals (Ndibualonji et al., 1995). Such phenomena may be assimilated to suckling stimuli, which have been reported to induce the release of corticosteroids in rats (Voogt et al., 1969). It has been suggested that both suckling (Voogt et al., 1969) and milking (Koprowski and Tucker, 1973) stimulate nerve endings in the nipples, which send impulses to the brain via the spinal cord, resulting in the sequential release of ACTH-releasing factor, pituitary ACTH, and finally adrenal corticosteroids. The animal response to stress factors is mainly centred in the activation of the sympathetic system and the hypothalamic–hypophysis–adrenal axis through catecholamine and glucocorticoid production

(Miller and O’Callaghan, 2002). These hormones render the animals alert, thus giving them the ability to react to environmental stimuli, in order to preserve organic homeostasis (Herman and Cullinan, 1997). Glucocorticoids and catecholamines also cause a rise in glycaemia, starting from glucidic and non-glucidic substrates such as proteins and an increase of non-esterified fatty acid (NEFA), derived from lipid mobilization (McMahon et al., 1988). Taken together, these effects produce a greater availability of energy for the brain and muscles, and thereby a more efficient behavioural response to stress. Plasma glucose concentration rose in both groups at T_1 and T_2 with respect to T_0 , although this increase was statistically significant in Group A, only. The rise in glucose levels is directly proportional to cortisol levels and must therefore be attributed to the hyperglycaemic effect of this hormone, to which we may add an increased glucose production by the liver, due to stimulation of sympathetic–adrenergic activity (Ali et al., 2001).

5 Conclusion

The results obtained in the present study suggest that, in addition to milking, the characteristics of the room where this procedure occurs represent stressful stimuli that could influence negatively productivity and welfare of dairy ewes. Therefore, the breeders should pay particular attention to the structures of their farms in order to make the functional spaces less stressful for animals.

Data availability. Data are available from the corresponding author upon request.

Author contributions. All authors have made substantial contributions to each step of experimental procedure and manuscript preparation: VC contributed to the experiment design and prepared the manuscript. FA performed the laboratory analysis. SL and CG performed the sampling. RM and PPB analysed the data. GP supervised all stages of the experimental study.

Competing interests. The authors declare that they have no conflict of interest.

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