

Contents lists available at ScienceDirect

Animal

The international journal of animal biosciences



Effects of spent coffee grounds on production traits, haematological parameters, and antioxidant activity of blood and milk in dairy goats



S. Carta a, E. Tsiplakou b, P. Nicolussi c, G. Pulina A, A. Nudda a,*

- ^a Dipartimento di Agraria, Sezione di Scienze Zootecniche, Università degli studi di Sassari, Viale Italia 39, 07100 Sassari, Italy
- ^b Department of Animal Science, Laboratory of Nutritional Physiology and Feeding, Agricultural University of Athens, lera Odos 75, GR-11855 Athens, Greece
- ^c Istituto Zooprofilattico Sperimentale della Sardegna, Sassari 07100, Italy

ARTICLE INFO

Article history: Received 11 October 2021 Revised 25 February 2022 Accepted 25 February 2022 Available online 1 April 2022

Keywords: Antioxidant status Byproduct Fatty acid methyl ester Milk goats Spent coffee ground

ABSTRACT

Spent coffee ground (SCG) is a byproduct of coffee beverage preparation and a potential source of carbohydrate, protein, and phenolic compounds for livestock feeding. In this study, the effects of SCG supplementation in the diet of lactating goats on milk production traits and health status were studied. The antioxidant status of blood and milk was also evaluated. Twenty-four Saanen goats were fed a total mixed ration containing commercial concentrate, soybean, and haylage; they were divided into three groups: control diet (CON), SCG50 (50 g/d SCG), and SCG100 (100 g/d SCG). The experiment lasted 6 weeks. Linear and quadratic contrasts were used to evaluate the effects of the byproduct doses. SCG supplementation did not affect milk production, but influenced some milk fatty acids. SCG supplementation increased the contents of C18:1, cis-9, trans-11 C18:2, odd and branched-chain fatty acids, and total conjugated linoleic acid. Most of the haematological and biochemical parameters were within the physiological range for goats. The basophil, eosinophil, and glucose contents were quadratically affected by SCG, whereas platelet count increased linearly with the SCG dose. The SCG supplementation had a positive effect on the blood antioxidant status, as evidenced by an increase in ferric reducing antioxidant power and a decrease in malondialdehyde. The SCG supplementation had no effect on the milk antioxidant status. The results show that SCG (up to 100 g/d) did not negatively affect milk production and health status in goats. However, quadratic effects on some antioxidant and biochemical parameters suggest that further investigations are necessary, especially with regard to the optimisation of the supplement dose. © 2022 The Authors. Published by Elsevier B.V. on behalf of The Animal Consortium. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Implications

Spent coffee ground is a byproduct of the global coffee industry, which can be conveniently recycled as a supplement for animal feeding. Spent coffee contains bioactive components that can positively affect animal productive performances and health status. This work showed that this byproduct did not have negative effects on animal performance and milk quality. Also, spent coffee ground decreased the oxidative stress in goat blood, improving the animal welfare. Spent coffee ground can replace some conventional feed ingredients, thus improving profitability of farms and reducing wastes from industry.

E-mail address: anudda@uniss.it (A. Nudda).

Introduction

Coffee is one of the most consumed beverages worldwide. Several byproducts are generated during its various processing stages. Among these byproducts, spent coffee ground (SCG) is the main byproduct of coffee beverage preparation. Approximately, 1 kg of green coffee generates 0.65 kg of SCG, and 1 kg of instant coffee produces approximately 2 kg of wet SCG. Most of this byproduct is incinerated or disposed of in landfills (San Martin et al., 2020). However, SCG contains bioactive compounds that could be recovered and reused, increasing the coffee industry's sustainability. SCG is a source of NDF (64-68%, Choi et al., 2018), protein (15.4%), and lipids (9.3-16.2%, Cruz et al., 2012), with a considerable amount of polyunsaturated fatty acids. In addition, SCG and other coffee byproducts are a good source of polyphenols (1–1.5% total polyphenols, Campos-Vega et al., 2015) that could exert a potential antioxidant effect. The literature has investigated the use of byproducts in livestock feeding (Nudda et al., 2017; Buffa et al., 2020a; Salami et al., 2021); however, further research on this

^{*} Corresponding author.

byproduct is necessary. The effects of SCG on milk composition have been investigated in cows (San Martin et al., 2020) and ewes (de Otálora et al., 2020). According to our literature review, no information is available regarding the effects of SCG on animal production performances, and health status of dairy goats. Thus, the aim of this study was to investigate the effects of SCG supplementation in the diet of lactating goats on milk yield, composition, fatty acid profile, and antioxidant stability. Blood antioxidant activity and animal health status were also studied to evaluate the healthiness of this byproduct. The doses used for this study were chosen, while considering the results of a recent study on sheep by de Otálora et al. (2020) along with the polyphenols and methylxanthines contents of SCG collected and used in this trial.

Preliminary results have been presented at the 24th Congress of the Animal Science and Production Association, and the abstract has been published in the conference proceedings (Carta et al., 2021).

Material and methods

Experimental design

Twenty-four Saanen dairy goats were randomly allocated to three experimental groups (eight animals per group) homogeneous for milk yield $(4.31\pm0.06~kg/d~per~head,~mean\pm SD)$, BW $(61.6\pm0.87~kg,~mean\pm SD)$, body condition score (BCS, $2.84\pm0.05,~mean\pm SD)$, and days in milk $(70\pm15~days,~mean\pm SD)$. The goats were machine milked twice daily at 08:00~and~20:00, and the individual milk production for the morning and evening milking was recorded weekly. The BCS (1-5~scale)~and~BW were recorded at the beginning and at the end of the experiment. The trial lasted 6~weeks, with 1~week of adaptation to the byproduct and 5~weeks of experimental sampling.

Each goat was fed a total mixed ration (**TMR**) as a control diet, containing 2.3 kg/d haylage, 160 g/d soybean meal, 200 g/d beet pulp, and 1.2 kg/d commercial concentrate. Animals of each group were housed together in a barn divided into three adjacent pens to ensure normal social behaviour. However, they were individually fed with the TMR offered four times per day: two times during milking and two times at 14:00 and 24:00 in individual self-capturing feeders. Fresh, clean water was available for ad libitum consumption. The experimental groups received the control diet supplemented with 0 (Control group, **CON**), 50 g (**SCG50**), and 100 g (**SCG100**) of SCG, respectively. The SCG dose was adminis-

Table 2Polyphenols and methylxanthines of spent coffee ground offered to dairy goats.

Items	mg/kg of SCG
Ferulic acid	12 393
Caffeic acid	885
5-Caffeoylquinic acid	158
Catechol	141
Trigonellin	12.5
1,2,4 benzenetriol	<0.1
Caffeine	894
Theobromine/Theophylline	283

Abbreviation: SCG = spent coffee ground.

tered in two aliquots offered to animals during morning and evening milking. The used byproduct was collected from different coffee shops in northern Sardinia and dried at 65 °C to prevent microbial deterioration. The chemical composition and the polyphenolic profile of SCG are reported in Tables 1 and 2, respectively.

Sampling and analysis

All dietary ingredients were collected at the beginning and at the end of the trial and were analysed for DM content by drying at 105 °C for 24 h. Feed samples were also analysed for NDF according to the method developed by Mertens (2002), using heat-stable amylase and expressed exclusive of residual ash (aNDForm). ADL were analysed according to the Association of official Analytical Chemists (AOAC) method 973.18 (AOAC, Official Method of Analysis, 2000; method 973.18); the ADL using the Robertson and Van Soest (1981) method and using sulfuric acid.

The CP was determined by using the Kjeldahl method (AOAC, 2000; method 988.05), and ash was determined using a muffle at 550 °C (AOAC, 2000; method 942.05). All these parameters were expressed as % of DM.

Individual milk samples were collected weekly, at the morning and afternoon milking for a total of five samplings. The samples were analysed separately for fat, protein, casein, lactose, and urea content (Milkoscan 6000, Foss Electric, Hillerød, Denmark) and for somatic cell count (**SCC**) (Fossomatic 360, Foss Electric). The weighted average of the morning and afternoon data was calculated for each considered parameter. Individual milk samples were

Table 1Ingredients and chemical composition of diets offered to dairy goats.

Items	Commercial concentrate	SCG	Diet	Diet			
			CON	SCG50	SCG100		
Ingredients mixed meal (kg/d per head, as fed)							
Spent coffee ground			-	0.05	0.10		
Beet pulp			0.20	0.20	0.20		
Soybean			0.16	0.16	0.16		
Commercial concentrate			1.2	1.2	1.2		
Haylage			2.3	2.3	2.3		
Total offered (kg/d, as fed)			3.86	3.91	3.96		
Total offered (kg/d, on DM)			2.30	2.35	2.39		
Total measured intake (kg/d, as fed)			3.80	3.83	3.90		
Chemical composition (% of DM)							
NDF	27.79	55.48	40.05	40.36	40.64		
ADF	12.31	33.15	23.55	23.74	23.92		
CP	16.93	15.03	15.95	15.93	15.91		
Ash	5.64	91.35	7.56	7.45	7.34		

Abbreviations: SCG = spent coffee ground; CON = diet without supplementation; SCG50 = diet supplemented with 50 g/d per head of spent coffee ground; SCG100 = diet supplemented with 100 g/d per head of spent coffee ground.

also collected every week and stored at $-20\,^{\circ}\text{C}$ for fatty acid methyl ester (**FAME**) analysis (Supplementary Material S1). Finally, milk samples for antioxidant analysis were collected at weeks 1, 2, and 5 of the trial and stored at $-20\,^{\circ}\text{C}$ and then analysed for ferric reducing antioxidant power (**FRAP**), 2,2'-azino-di(3-ethylbenzthia zoline-6-sulforic acid; **ABTS**), malondialdehyde (**MDA**), and protein carbonyls (Supplementary Material S2).

Blood samples were collected by jugular venipuncture after the morning milking at the beginning and at the end of the experiment. Vacutainers with clotting accelerator were used to collect blood for serum, whereas vacutainers with dipotassiumethylenediamine tetraacetic acid, as an anticoagulant, were used to have the whole blood, respectively. For the antioxidant analysis, blood samples were collected with a heparinised vacuum tube and then centrifuged at 3 000g, at 4 °C, for 10 min to separate plasma from the corpusculate fraction and were stored at -80 °C. Serum samples were also analysed by an automatic spectrophotometer to determine the quantity of albumin, alkaline phosphatase, bilirubin, calcium, creatinine, phosphorus, protein, gamma-glutamyl transferase, serum glutamic oxaloacetic transaminase, serum glutamic-pyruvic transaminase, urea, cholesterol, creatine kinase, glucose, and triglycerides. Using an electronic particle counter, the following measurements were conducted: red blood cells, white blood cells, platelets, haemoglobin, mean corpuscular haemoglobin concentration and proportion, corpuscular haemoglobin haematocrit, mean corpuscular volume, lymphocytes, monocytes, granulocytes neutrophils, eosinophils, and basophils.

The antioxidant capacity in milk and blood was evaluated using different assays. In both matrices, the total antioxidant capacity was evaluated by FRAP and ABTS, whereas the lipid and protein oxidative biomarkers were determined by MDA and protein carbonyls assays (Supplementary Material S2).

Statistical analysis

All data (i.e. animal performance, milk composition, and haematological parameters) were analysed as a completely randomised design with repeated measures by using the PROC MIXED procedure of SAS software (version 9.2; SAS Institute, 2008). The model included the fixed effects of diet, sampling date, and diet \times sampling date interaction, and the random effect of the animal (Supplementary Material S3). The differences among the levels of the considered effects were tested using orthogonal polynomial contrasts (linear and quadratic). Effects were considered significant at P < 0.05 and to have a tendency to difference at P < 0.10.

Results

Dry matter intake, body weight, and milk yield and composition

The SCG was completely consumed by the goats of both supplemented groups, and it did not influence the feed intake (P = 0.49). The latter was 3.80, 3.83, and 3.90 kg/d per head for CON, SCG50, and SCG100, respectively. BW was not affected by the SCG supplementation (P = 0.64), but it decreased significantly from the beginning to the end of the experiment in all animals (from 61.6 ± 8.6 kg to 58.5 ± 8.4 kg, overall mean \pm SD). The effect of dietary supplementation of SCG on milk yield and composition is reported in Table 3. Milk yield was not affected by SCG supplementation (P = 0.73), but it decreased throughout the experimental period (P < 0.001; Supplementary Fig. S1). SCG supplementation did not influence the milk protein, fat, and lactose contents. Urea yield (g/d) tended to decrease linearly (P = 0.08) with SCG supplementation. The SCG affected the SCC quadratically, because the low dose decreased the SCC (P < 0.001), whereas the highest dose did not change the cellular count compared with that of the CON group. The content of NaCl linearly increased with the SCG supplementation (P = 0.05).

Milk fatty acid profile

The effect of SCG on milk fatty acid profile is reported in Table 4. The short-chain fatty acids C6:0 (P = 0.05) and C8:0 (P = 0.09) tended to decrease linearly. Branched-chain fatty acids, as isoC14:0 (P = 0.001) and anteisoC15:0 (P = 0.04), increased linearly, and isoC15:0, isoC16:0, and anteisoC17:0 tended to increase linearly with SCG supplementation. The SCG supplementation resulted in a linear increase in C15:0 (P = 0.01). The SCG supplementation affected the concentrations of C18:1 trans-11 that increased quadratically (P = 0.005), and of CLA cis-9 trans-11 that increased linearly.

Blood and milk oxidation

Blood oxidation capacity improved after using SCG in the diet (Table 5). Notably, the coffee byproduct supplementation increased the FRAP (P = 0.02) and decreased the MDA (P = 0.01). The antioxidant capacity of SCG on blood plasma is confirmed by the MDA assays. However, ABTS and protein carbonyls were not influenced by the byproduct. The SCG supplementation did not affect any antioxidant parameters tested in goat milk (Table 6).

Table 3Effect of dietary supplementation of two doses of spent coffee ground on goat milk yield and composition.

Items	Diet			SEM	P-value		
	CON	SCG50	SCG100		L	Q	Time
Yield (g/d)							
Milk	3 680.9	3 592.0	3815.5	58.12	0.669	0.508	< 0.001
Fat	115.65	107.70	114.21	1.88	0.900	0.343	0.009
Protein	111.08	103.98	110.10	1.63	0.910	0.333	< 0.001
Lactose	162.05	156.45	164.25	2.66	0.866	0.549	< 0.001
Urea	1.37	1.30	1.29	0.02	0.082	0.615	< 0.001
Milk composition							
Fat (%)	3.16	3.01	3.02	0.04	0.521	0.740	0.029
Protein (%)	3.03	2.92	2.89	0.02	0.273	0.688	0.488
Lactose (%)	4.41	4.36	4.29	0.02	0.205	0.848	0.020
Urea (mg/dl)	37.53	36.61	33.73	0.53	0.411	0.671	< 0.001
Log SCC (1 000 cell/ml)	2.66	2.46	2.89	0.04	0.074	< 0.001	< 0.001
NaCl	302.50	325.57	334.16	3.08	0.051	0.666	0.015

Abbreviations: CON = diet without supplementation; SCG50 = diet supplemented with 50 g/d per head of spent coffee ground; SCG100 = diet supplemented with 100 g/d per head of spent coffee ground; SCC = somatic cell count; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

 Table 4

 Fatty acid profile and nutritional indices of fatty acids of milk of goat fed with control diet (CON), 50 g spent coffee ground (SCG50) and 100 g of spent coffee ground (SCG100).

Fatty acid	Diet			SEM	P-value			
(g/100 g of FAME)	CON	SCG50	SCG100		L	Q	Tim	
C4:0	1.70	1.68	1.75	0.03	0.778	0.267	0.06	
C6:0	1.82	1.79	1.73	0.02	0.051	0.093	<0.0	
C8:0	2.08	1.97	1.95	0.02	0.089	0.750	<0.0	
C9:0	0.06	0.07	0.08	0.002	0.059	0.801	<0.0	
C10:0	7.59	7.22	7.33	0.10	0.155	0.999	<0.0	
C10:1	0.04	0.04	0.04	0.001	0.363	0.473	<0.0	
C11:0	0.22	0.22	0.20	0.004	0.620	0.157	<0.0	
C12:0	3.38	3.27	3.32	0.05	0.985	0.729	<0.0	
so C13:0	0.02	0.02	0.02	0.0004	0.458	0.990	0.63	
inteiso C13:0	0.03	0.03	0.03	0.001	0.548	0.107	<0.0	
so C14:0	0.09	0.10	0.11	0.002	0.001	0.826	<0.0	
214:0	10.06	9.86	10.18	0.08	0.719	0.334	<0.0	
C14:1cis-9	0.11	0.11	0.10	0.003	0.380	0.422	<0.0	
so C15:0	0.22	0.22	0.24	0.003	0.063	0.565	<0.0	
nteiso C15:0	0.34	0.35	0.36	0.005	0.038	0.688	<0.0	
15:0	0.93	0.94	1.00	0.01	0.013	0.525	<0.0	
15:1	0.05	0.05	0.05	0.001	0.516	0.912	<0.0	
so C16:0	0.29	0.31	0.34	0.005	0.058	0.928	<0.0	
16:0	25.84	26.77	27.03	0.22	0.586	0.386	<0.0	
16:1trans-4	0.04	0.05	0.04	0.001	0.996	0.130	<0.0	
16:1trans-5	0.04	0.04	0.03	0.001	0.776	0.351	<0.0	
16:1 trans-6 + trans-7	0.05	0.05	0.05	0.001	0.214	0.632	0.0	
SoC17_0	0.40	0.40	0.40	0.01	0.793	0.919	0.7	
16:1 trans-9	0.10	0.08	0.09	0.01	0.906	0.243	0.1	
16:1 trans-10	0.02	0.02	0.02	0.0004	0.336	0.829	<0.	
16:1trans-11 + trans-12	0.04	0.04	0.04	0.001	0.339	0.505	<0.	
16:1cis-7	0.30	0.29	0.29	0.0003	0.711	0.217	<0.	
nteiso C17:0	0.43	0.42	0.45	0.005	0.088	0.343	0.0	
16:1cis-9	0.53	0.52	0.48	0.01	0.771	0.500	<0.	
16:1cis-10	0.02	0.02	0.02	0.0004	0.584	0.955	<0.	
16:1cis-11	0.01	0.01	0.01	0.0003	0.671	0.910	<0.	
17:0	0.82	0.81	0.83	0.01	0.215	0.713	<0.	
17:1cis-9	0.28	0.28	0.25	0.01	0.651	0.893	<0.	
18:0 (SA)	11.44	11.30	10.82	0.17	0.508	0.343	<0.	
18:1trans-4	0.01	0.01	0.01	0.0003	0.456	0.741	<0.	
18:1trans-5	0.01	0.01	0.01	0.0004	0.823	0.169	<0.	
18:1trans-6 + trans-8	0.15	0.15	0.15	0.002	0.376	0.594	<0.	
18:1trans-9	0.18	0.19	0.19	0.0002	0.093	0.558	0.0	
18:1trans-10	0.13	0.22	0.13	0.004	0.606	0.887	0.0	
18:1 trans-11(VA)	0.73	0.68	0.74	0.004	0.870	0.005	0.0	
18:1trans-12	0.20		0.74	0.003	0.280	0.657	<0.	
		0.21						
18:1trans-13 + trans-14	0.32	0.33	0.35	0.01	0.328	0.996	0.3	
18:1cis-9	22.02	22.16	21.58	0.22	0.731	0.456	<0.	
18:1cis-11	0.54	0.54	0.52	0.01	0.914	0.370	<0.	
18:1 cis-12	0.21	0.21	0.21	0.003	0.405	0.830	<0.	
18:1cis-13	0.06	0.05	0.05	0.001	0.826	0.679	<0.	
18:1 trans-16 + cis-14	0.23	0.24	0.24	0.004	0.102	0.014	<0.	
18:2trans-9, trans-12	0.01	0.01	0.01	0.0004	0.955	0.533	0.1	
18:2 cis-9, trans-12	0.071	0.072	0.073	0.001	0.033	0.509	<0.	
18:2 trans-9, cis-12	0.02	0.02	0.02	0.0003	0.591	0.583	0.0	
18:2 trans-11, cis-15	0.049	0.044	0.05	0.001	0.037	0.319	0.0	
18:2n-6 (LA)	2.71	2.69	2.70	0.03	0.945	0.386	0.0	
18:3n-6	0.03	0.03	0.03	0.0006	0.612	0.683	<0.	
18:3n-3 (LNA)	0.39	0.35	0.40	0.01	0.813	0.686	<0.	
LA cis-9, trans-11 (RA)	0.411	0.413	0.444	0.01	0.049	0.018	0.0	
18:4n-3	0.01	0.01	0.01	0.0002	0.756	0.586	<0.	
20:0	0.26	0.29	0.28	0.004	0.139	0.249	0.0	
LAtrans-9, cis-11 + C21:0	0.06	0.06	0.06	0.001	0.064	0.126	<0.	
LA trans-10, cis-12	0.01	0.01	0.01	0.0004	0.191	0.480	0.0	
LA trans-10, cis-12 LA trans-11, trans-13	0.02	0.02	0.02	0.0004	0.131	0.664	<0.	
LAtrans-9, trans-11	0.02	0.02	0.01	0.0003	0.922	0.445	<0.	
20:1cis-9	0.03	0.03	0.03	0.0002	0.376	0.358	0.0	
	0.0088	0.03	0.03		0.376	0.393	0.0	
20:2n-9				0.0002				
20:2n-6	0.02	0.02	0.02	0.0005	0.970	0.423	<0.	
20:3n-9	0.05	0.04	0.05	0.001	0.401	0.068	<0.	
20:3n-6	0.03	0.02	0.03	0.0005	0.889	0.300	0.4	
20:4n-6	0.19	0.20	0.20	0.002	0.341	0.900	<0.	
20:3n-3	0.01	0.01	0.01	0.0002	0.733	0.969	0.2	
22:0	0.08	0.08	0.08	0.001	0.200	0.337	<0.0	
20:4n-3	0.01	0.01	0.01	0.0003	0.368	0.373	0.4	
C22:1n-9	0.0111	0.0121	0.0124	0.0002	0.009	0.421	<0.0	
C22:5n-3 (EPA)	0.04	0.03	0.04	0.001	0.887	0.579	0.0	

Table 4 (continued)

Fatty acid	Diet			SEM	P-value		
(g/100 g of FAME)	CON	SCG50	SCG100		L	Q	Time
C22:4n-6	0.19	0.20	0.20	0.001	0.444	0.986	0.033
C24:0	0.03	0.03	0.03	0.001	0.973	0.937	0.336
C22:5n-3 (DPA)	0.08	0.07	0.09	0.002	0.149	0.326	< 0.001
C22:6n-3 (DHA)	0.02	0.02	0.02	0.0005	0.356	0.341	0.103
Groups							
SCFA	13.29	12.76	12.88	0.14	0.154	0.895	< 0.001
MCFA	44.84	45.44	46.18	0.24	0.621	0.541	< 0.001
LCFA	41.86	41.79	40.93	0.32	0.593	0.553	< 0.001
SFA	68.40	68.40	68.83	0.27	0.656	0.399	< 0.001
MUFA	26.89	26.96	26.39	0.25	0.687	0.404	< 0.001
PUFA	4.70	4.62	4.76	0.04	0.452	0.365	< 0.05
UFA	31.59	31.58	31.15	0.27	0.664	0.404	< 0.001
OCFA	2.09	2.10	2.16	0.02	0.026	0.534	< 0.001
BCFA	1.89	1.91	2.00	0.02	0.037	0.882	0.050
OBCFA	3.98	4.01	4.17	0.03	0.017	0.897	< 0.001
PUFA n-6	3.05	3.03	3.05	0.03	0.979	0.425	< 0.001
PUFA n-3	0.55	0.50	0.56	0.01	0.998	0.863	< 0.001
n6:n3	5.69	6.21	5.53	0.09	0.738	0.387	< 0.001
Total CLA	0.52	0.52	0.55	0.01	0.044	0.041	0.002
TFA	2.81	2.78	2.88	0.03	0.188	0.295	< 0.001
Δ9- desaturase indices							
C10:1	0.49	0.51	0.54	0.02	0.373	0.629	0.003
C14:1	1.09	1.11	1.00	0.02	0.176	0.165	< 0.001
C16:1	2.04	1.94	1.77	0.04	0.652	0.482	< 0.001
C18:1	65.85	66.23	66.63	0.39	0.449	0.293	0.064
CLA cis-9, trans-11	36.02	37.70	37.30	0.38	0.022	0.418	< 0.001
AI	2.22	2.24	2.30	0.03	0.886	0.559	< 0.001
TI	2.16	2.24	2.26	0.03	0.788	0.407	< 0.001
h:H	0.73	0.72	0.70	0.01	0.862	0.492	< 0.001

Abbreviations: FAME = fatty acid methyl ester; SA = stearic acid; VA = vaccenic acid; LA = linoleic acid; LNA = linolenic acid; RA = rumenic acid; EPA = eicosapentaenoic acid; DPA = docosapentaenoic acid; SFA = sum of the individual saturated fatty acids; UFA = sum of the individual unsaturated fatty acids; MUFA = sum of the individual monounsaturated fatty acids; PUFA = sum of the individual polyunsaturated fatty acids; TFA = trans fatty acids, sum of the individual trans fatty acids, except CLA isomers; OCFA = odd-chain fatty acids; BCFA = branched-chain fatty acids, sum of iso- and anteiso-FA; OBCFA = odd- and branched-chain fatty acids, sum of odd-, iso-, and anteiso-FA; SCFA = short-chain fatty acids, sum of the individual fatty acids from C1:0 to C10:0; MCFA = medium-chain fatty acids, sum of the individual fatty acids from C18:0 to DHA; PUFA n-3 and PUFA n-6 = sum of individual n-3 and n-6 fatty acids, respectively; CLA = sum of individual conjugated linoleic acids; TI = thrombogenic index; AI = atherogenic index; h:H = hypocholesterolemic to hypercholesterolemic ratio; CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; L = linear effect of the diet; Time = sampling time effect.

Table 5Total antioxidant capacity, malondialdehyde, and protein carbonyls content from goat blood plasma for each treatment.

Items	Diet			SEM	P-value	P-value			
	CON SCG50 SCG100			L	Q	Time			
FRAP (µmol ascorbic acid)	0.72	0.71	0.82	0.02	0.016	0.063	<0.001		
ABTS (% inhibition)	34.21	31.99	31.86	0.51	0.096	0.387	0.006		
MDA (µM)	0.78	0.68	0.66	0.02	0.010	0.396	0.023		
PC (nmol/ml)	8.89	8.81	8.72	0.21	0.607	0.981	< 0.001		

Abbreviations: CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; FRAP = Ferric Reducing Ability of Plasma; ABTS = 2,2'-azino-di(3-ethylbenzthiazoline-6-sulforic acid); MDA = Malondialdehyde; PC = Protein carbonyls; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

Blood analysis

The results of haematological and biochemical parameters are reported in Tables 7 and 8. The eosinophils concentration and percentage (P = 0.03 and 0.02, respectively) and basophils percentage (P = 0.01) quadratically decreased with the SCG supplementation. In addition, platelets count was markedly increased by both doses of SCG (P = 0.02), but its values remained within the physiological range.

Regarding biochemical parameters, the SCG did not cause the alteration of enzymatic activities in the liver and the kidney. The glucose concentration was decreased quadratically by SCG. The SCG supplementation decreased linearly with the creatine kinase (Table 8). Table 9 shows the effects of SCG on the electrophoresis profile of serum protein fractions. The SCG supplementation did

not affect the content of serum blood protein and albumin. However, this byproduct tended to decrease α_1 -globulins (P = 0.08) and to increase γ -globulins (P = 0.09) but always within the physiological range. However, the total globulins and the ratio of globulins to albumin did not change with the SCG inclusion in the diet.

Discussion

Dry matter intake, body weight, and milk yield and composition

In this study, the effect of SCG supplementation on a goat diet was investigated. This byproduct is not commonly used in animal feeding, probably because some its compounds (e.g. melanoidines, alkaloids, or polyphenols) could reduce diet palatability and

Table 6Total antioxidant capacity, malondialdehyde, and protein carbonylis content from goat milk for each treatment.

Items	Diet			SEM	P-value	P-value			
	CON	SCG50	SCG100		L	Q	Time		
FRAP (µmol ascorbic acid)	2.98	2.78	2.76	0.07	0.354	0.655	0.002		
ABTS (% inhibition)	32.90	31.16	30.77	0.65	0.326	0.728	0.121		
MDA (μM)	0.23	0.20	0.21	0.01	0.170	0.233	0.009		
PC (nmol/ml)	4.53	4.07	4.22	0.12	0.147	0.117	< 0.001		

Abbreviations: CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; FRAP = Ferric Reducing Ability of Plasma; ABTS = 2,2'-azino-di(3-ethylbenzthiazoline-6-sulforic acid); MDA = Malondialdehyde; PC = Protein carbonyls; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

 Table 7

 Effect of dietary supplementation of two doses of spent coffee ground on haematological parameters in dairy goats.

Items	Reference values	Diet			SEM	P-value		
		CON	SCG50	SCG100		L	Q	Time
WBC (10 ³ /μl)	6.43-10.93	11.72	12.72	13.40	0.37	0.283	0.922	0.730
Neu $(10^3/\mu l)$	2.09-4.8	5.38	5.63	6.57	0.24	0.191	0.606	0.718
Lym $(10^3/\mu l)$	3.26-5.42	5.54	6.51	6.04	0.18	0.487	0.262	0.352
Mon $(10^3/\mu l)$	0.06-0.38	0.37	0.31	0.39	0.03	0.796	0.414	0.462
Eos $(10^3/\mu l)$	0.16-0.34	0.25	0.11	0.20	0.02	0.417	0.029	0.013
Bas $(10^3/\mu l)$	0.03-0.1	0.06	0.06	0.08	0.004	0.324	0.178	< 0.001
LUC $(10^3/\mu l)$	-	0.10	0.11	0.12	0.01	0.445	0.963	0.081
Neu (%)	27-53	45.82	43.97	48.00	0.92	0.534	0.269	0.888
Lym (%)	39.2-63	47.50	51.48	46.02	0.93	0.680	0.095	0.405
Mon (%)	0.7-4.1	2.91	2.42	3.05	0.20	0.77	0.23	0.43
Eos (%)	1.8-4.5	2.36	0.85	1.44	0.16	0.089	0.021	0.034
Bas (%)	0.3-1.1	0.58	0.44	0.60	0.02	0.768	0.014	0.002
LUC (%)	-	0.84	0.86	0.90	0.05	0.724	0.833	0.094
RBC $(10^6/\mu l)$	12.48-16.03	14.16	14.04	13.93	0.13	0.615	0.973	0.003
HGB (g/dl)	8.3-10.8	9.01	8.96	9.06	0.07	0.841	0.764	0.001
HCT (%)	24.7-32.2	27.20	26.32	27.30	0.28	0.902	0.287	0.057
MCH (pg)	5.6-7.6	6.40	6.40	6.53	0.05	0.533	0.775	< 0.0001
MCHC (g/dl)	31.5-35.9	33.21	33.50	33.26	0.08	0.911	0.359	< 0.0001
MCV (fl)	15.5-24.3	19.26	19.10	19.65	0.19	0.575	0.649	< 0.0001
PLT $(10^{3}/\mu l)$	130-624	151.48	307.13	345.67	21.92	0.023	0.413	< 0.0001

Abbreviations: WBC = white blood cell count; Neu = neutrophil granulocytes; Lym = lymphocytes; Mon = monocytes; Eos = eosinophils granulocytes; Bas = basophiles granulocytes; LUC = Large unstained cells; RBC = red blood cell; HGB = haemoglobin; HCT = haematocrit; MCH = mean corpuscular haemoglobin; MCHC = mean corpuscular haemoglobin concentration; MCV = mean corpuscular volume; PLT = platelet; CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

 Table 8

 Effect of dietary supplementation of two doses of spent coffee ground on biochemical parameters in dairy goats.

Items	Reference values	Diet			SEM	P-value		
		CON	SCG50	SCG100		L	Q	Time
Albumin (g/dl)	2.5-3.5	2.87	2.90	2.73	0.05	0.104	0.146	<0.0001
ALP (u/l)	35-300	196.17	176.54	264.75	44.43	0.730	0.754	< 0.05
Total bilirubin (mg/dl)	0.3-0.6	0.22	0.20	0.22	0.01	0.894	0.448	< 0.05
Calcium (mg/dl)	6-9.5	8.26	8.03	8.03	0.18	0.459	0.680	< 0.0001
Creatinine (mg/dl)	0.3-0.8	0.59	0.62	0.57	0.02	0.605	0.341	< 0.0001
Phosphorus (mg/dl)	3.5-12	6.88	7.15	6.59	0.22	0.683	0.497	< 0.0001
Total protein (g/dl)	6.5-8.5	8.32	8.22	7.99	0.18	0.312	0.818	< 0.0001
GGT (u/l)	40-135	53.75	50.67	64.58	2.45	0.252	0.298	< 0.0001
GOT/AST (u/l)	170-180	135.67	119.21	124.21	4.47	0.492	0.458	< 0.0001
GPT/ALT (u/l)	25-50	32.25	32.75	26.00	1.41	0.805	0.994	< 0.0001
Urea (mg/dl)	50-80	38.71	37.54	36.29	0.87	0.243	0.981	< 0.0001
Cholesterol (mg/dl)	50-95	86.96	88.54	91.58	2.66	0.594	0.923	< 0.0001
CK (u/l)	35-150	268.00	215.29	222.96	12.97	0.074	0.164	< 0.0001
Glucose (mg/dl)	25-60	30.00	34.71	27.25	1.29	0.356	0.022	< 0.0001
Triglyceride (mg/dl)	10-50	16.3	21.3	25.4	0.37	0.326	0.959	< 0.05

Abbreviations: CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; ALP = alkaline phosphatase; GGT = gamma-glutamyl transferase; GOT/AST = serum glutamic oxaloacetic transaminase; GPT/ALT = serum glutamic-pyruvic transaminase; CK = creatine kinase; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

acceptability. A depressive effect on diet palatability and daily intake in sheep supplemented with a 10% level of SCG was reported by Choi et al. (2018). However, in this study, no adverse effects on goat DM intake were observed using 50 or 100 g/d of SCG.

Milk yield decreased during the experimental period (Supplementary Fig. S1) partly due to the high temperature recorded during the trial (T_{mean} 23.3 ± 2.4 °C, T_{max} 31.3 ± 3.6 °C; RH_{mean} 57.5 ± 8.7, RH_{max} 89.9 ± 8.1; Supplementary Fig. S2). However,

Table 9Effect of dietary supplementation of two doses of spent coffee ground on serum protein fractions in dairy goats.

Items	Reference values	Diet	Diet			P-value		
		CON	SCG50	SCG100		L	Q	Time
Total protein (g/dl)	6.5-8.5	7.68	7.81	7.51	0.23	0.679	0.542	<0.001
Albumin (%)	32.8-42.3	43.52	41.93	40.23	1.11	0.229	0.983	0.027
α_1 -globulins (%)	4.5-7.6	6.06	5.63	5.17	0.18	0.085	0.983	0.024
α ₂ -globulins (%)	11.6-16.1	12.81	13.39	12.64	0.26	0.815	0.289	0.379
β-globulins (%)	5.2-8.9	6.68	6.52	6.88	0.34	0.780	0.685	< 0.001
γ-globulins (%)	21-36.7	30.96	32.54	35.08	0.94	0.090	0.811	< 0.001
Total globulins (%)	_	56.51	58.08	59.77	1.11	0.232	0.978	0.028
Globulins/albumin	_	1.37	1.43	1.59	0.06	0.157	0.737	0.041

Abbreviations: CON = control diet; SCG50 = diet containing 50 g/d per head of spent coffee ground; SCG100 = diet containing 100 g/d per head of spent coffee ground; L = linear effect of the diet; Q = quadratic effect of the diet; Time = sampling time effect.

the SCG supplementation did not influence milk protein, fat, and lactose contents, in accordance with results in cows (San Martin et al., 2020), but contrasted with observations in dairy sheep, in which an increase in milk fat and protein contents was found with the ingestion of a similar dose of spent coffee (de Otálora et al., 2020). The high temperatures recorded during the experimental period may have covered the potential impacts of this supplement on milk components.

The observed depressive effect of SCG on milk urea is in accordance with an observation in dairy cows supplemented with coffee hulls in combination with soybean oil (Santos et al., 2014). This result could be partly due to the presence of caffeine, which can affect urea metabolism (Jorda et al., 1989), and ferulic acid, which is the most abundant polyphenolic compound present in SCG. Notably, polyphenols can reduce dietary protein degradation in the rumen and, consequently, ammonia production (Correddu et al., 2020). A study in sheep reported that the inclusion of a 10% level of SCG could reduce protein digestibility and nitrogen absorption (Choi et al., 2018). In spite of the high content of ferulic acid in SCG (12 g/kg of SCG), animals ingested a low amount per kilogram of BW because of the reduced amount of byproduct offered. The ingested amounts were 10.5 and 20.1 mg/kg BW in groups SCG50 and SCG100, respectively. A study conducted on laboratory animals, treated with 25 mg/kg BW of ferulic acid, showed a significant decrease in the levels of urea, uric acid, and creatinine (Manikandan et al., 2014). This result could support the role of ferulic acid in nitrogen metabolism in goats. The SCC was quadratically affected by SCG, which was reduced only by the low dose of the byproduct. A study on dairy cows affected by subclinical mastitis reported that feeding coffee ground silage containing large amounts of polyphenols decreased the milk SCC (Kawai et al., 2018), suggesting a positive role on the immune activity.

Milk fatty acid profile

The fatty acid profile of ruminants' products is gaining attention from consumers, as demonstrated by the studies that have attempted to decipher the phenotypic and genetic background of this trait (Mele et al., 2016; Chikwanha et al., 2018; Cesarani et al., 2019; Correddu et al., 2021). Thus, in this study, the effect of SCG on the milk fatty acid profile was investigated. The SCG supplementation resulted in small variations in the milk fatty acid profile. In particular, a linear increase in some branched chain fatty acids, as isoC14:0 and anteisoC15:0, and C15:0 has been observed. These fatty acid are considered biomarkers of rumen microbial fermentation and microbial de novo lipogenesis. Their increase with SCG supplementation suggests that polyphenols may have affected the rumen microbial profile (Salami et al., 2021). The increase in C15:0 is also notable from a nutritional view because of the bene-

ficial effects on human health: the potential reduction of inflammation and incidence of cardiovascular and liver diseases (Venn-Watson et al., 2020). The increase in C18:1 trans-11 and CLA cis-9 trans-11 with SCG supplementation suggests a slowing of rumen biohydrogenation of dietary polyunsaturated fatty acids, probably due to the presence of polyphenols in the byproduct. However, there was no substantial change in the overall contents of unsaturated and saturated fatty acid in milk, despite the results in the literature that the inclusion of polyphenols in ruminant diets modifies the fatty acid profile of milk (Correddu et al., 2020).

Blood and milk oxidation

The antioxidant capacity of blood was improved by the SCG supplementation, as demonstrated by the increase in FRAP and decrease in MDA. The effect on FRAP, together with MDA values, suggested that the higher the dose the better the antioxiandat capacity. An improvement in blood antioxidant capacity due to coffee consumption measured with FRAP assay was reported in humans (Martínez-López et al., 2019). The same improvement, measured by the MDA assays, was also reported in sheep fed coffee pulp (Salinas-Rios et al., 2015), in cows fed ground coffee silage (Kawai et al., 2018), and in laboratory animals fed coffee (Vitaglione et al., 2010). The antioxidant compounds, determined by the ABTS assay, might be responsible for the reduction of MDA, one of the main products of lipid oxidation. The antioxidant potential of SCG could be related to the ability of caffeine, ferulic acid (4-hydroxy-3-methoxycinnamic acid), and secondary metabolites (e.g. melanoidins) to trap free radicals (Vitaglione et al., 2010; Salinas-Rios et al., 2015). The antioxidant capacity of milk was not modified by the SCG supplementation. Similar results were found in dairy ewes (Buffa et al., 2020b) by using other byproducts rich in polyphenols, such as grape marc and exhausted myrtle berries. These byproducts did not modify milk ABTS and FRAP but reduced milk protein carbonyls. In our trial, the protein carbonyls were not affected by the SCG supplementation; thus, its polyphenols did not show a protective role against milk protein oxidation.

Blood analysis

The results of haematological and biochemical parameters showed an effect of SCG on goat immune response, demonstrated by the quadratic effect on eosinophils concentration and for basophils percentage, that achieved the lowest values in the group fed with the low SCG dose. The eosinophils plays a crucial role in the inflammatory response and in the modulations of acute and chronic hyperimmune reactions, via the release of inflammatory mediators into tissue sites such as specific granule proteins. Eosinophils are also involved in the regulation of the immune response

against helminth infections (pulmonary, intestinal, or hepatic) in sheep, causing their increase above the upper limit of the physiological range. Even if all animals appeared healthy and no parasite search in faeces was assessed during the experiment, the reduction of eosinophils suggests a positive effect of SCG. The increase in platelets with both doses of SCG could be related to the function of caffeine and other compounds in blood homeostasis. Commonly, phenolic compounds of coffee reduce blood platelet aggregation in humans (Olas and Bryś, 2019), but the increase in platelets in goat blood with SCG consumption is difficult to explain, even if it remains within the physiological range. Regarding biochemical parameters, the SCG did not alter enzymatic activities in the liver and kidney. Only the glucose concentration was decreased quadratically by SCG. An opposite pattern was observed in humans: several studies have demonstrated an increase in glucose after coffee consummation, probably due to the implication of caffeine in glucose metabolism (Johnston et al., 2003; Moisev et al., 2008). Notably, ferulic acid has been found to exert antidiabetic properties by stimulating glucose uptake (Sompong et al., 2015) and improving insulin sensitivity (Narasimhan et al., 2015). Unfortunately, little is known of the effects of bioactive compounds specifically present in coffee on blood parameters and on glucose metabolism in ruminants. However, different studies conducted using polyphenols in the diet showed contrasting results: Chedea et al. (2017) did not find an effect on plasma glucose in dairy cows after ingestion of grape pomace; by contrast, Zhong et al. (2011) reported increased glucose levels in goats fed tea catechin. The different results of these studies could be attributed to the different phenolic composition and to the different amounts of consumed phenols. In our study, creatine kinase decreased linearly in the plasma of goats supplemented with SCG. High creatine kinase activity is associated with cardiac and skeletal muscle damages or with muscle intense activity after exercise (Rider and Miller, 1995; Cechella et al., 2014). The reduction of creatine kinase in blood during exercise has been also observed in rats fed caffeine, which can reduce the damaged muscle fibres, probably due to its strong activity against oxidative stress (Cechella et al., 2014).

No effect was found for serum protein and albumin, even if there was a numerical decrease in α_1 -globulins and an increase in γ -globulins. Albumin, α - and β -globulins are synthesised in the liver; γ -globulins are produced by B lymphocytes or plasma cells and are responsible for the humoral response of the immune system. Usually, α - and γ -globulins are involved in the immune response of the mammary gland; notably, α -globulins production tends to decrease, whereas γ -globulins tend to increase after an inflammatory stimulus. In our study, the total globulins and the ratio of globulins to albumin did not change with SCG inclusion in the diet, suggesting a natural variation in globulin fractions in response to a specific treatment, the inclusion of a supplement.

Conclusion

The inclusion of up to 100 g/d of SCG into the diet of dairy goats did not affect feed intake, milk yield, and almost all milk components. The analyses of blood oxidation markers suggest that SCG can be used as a putative dietary ingredient to modulate oxidative stress in goats. The responses of animals to this byproduct were associated with the given doses; thus, an assessment of the most optimum level of SCG in animal diet is necessary. Moreover, it is relevant to explore the effects of longer periods of treatments with bioactive substances on animal production traits and health and oxidative statuses. Finally, the hormetic response of some parameters suggests that further studies on dose–response effects of the bioactive compounds present in several agro-industrial byproducts are necessary for dairy animals.

Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.animal.2022.100501.

Ethics approval

The experiment was approved by the Ethics Committee of the University of Sassari (no. 87140/2020).

Data and model availability statement

None of the data were deposited in an official repository. The data that support the study findings are available upon request.

Author ORCIDs

Carta, Silvia - https://orcid.org/0000-0002-5778-2627, Tsiplakou, Eleni - https://orcid.org/0000-0002-2544-8966, Pulina, Giuseppe - https://orcid.org/0000-0001-5579-0677, Nudda, Anna - https://orcid.org/0000-0002-9807-0626.

Author contributions

S. Carta: investigation, formal analysis, writing - original draft; E. Tsiplakou: formal analysis, writing- review and editing; P. Nicolussi: blood analysis and interpretation; G. Pulina: interpreted data and revised the manuscript; A. Nudda: conceptualised and designed the study, funding acquisition, supervision, and writing. All the authors have read and approved the final version of this manuscript.

Declaration of interest

None.

Acknowledgements

The authors wish to thank Dr. Antonio Piras and Giovanni Pinna for their valuable technical assistance and Roberto Rubattu for his precious assistance in laboratory analysis. The authors gratefully acknowledge the financial support of Fondazione di Sardegna (annualità 2017).

Financial support statement

The research has been funded by Fondazione di Sardegna, annualità 2017 (CUP J85F20000400007).

References

Association of official Analytical Chemists (AOAC), 2000. Official Methods of Analysis. AOAC, Arlington, VA, USA.

Buffa, G., Mangia, N.P., Cesarani, A., Licastro, D., Sorbolini, S., Pulina, G., Nudda, A., 2020a. Agroindustrial by-products from tomato, grape and myrtle given at low dosage to lactating dairy ewes: effects on rumen parameters and microbiota. Italian Journal of Animal Science 19, 1462.

Buffa, G., Tsiplakou, E., Mitsiopoulou, C., Pulina, G., Nudda, A., 2020b. Supplementation of byproducts from grape, tomato and myrtle affects antioxidant status of dairy ewes and milk fatty acid profile. Journal of Animal Physiology and Animal Nutrition 104, 493–506.

Campos-Vega, R., Loarca-Pina, G., Vergara-Castaneda, H.A., Oomah, B.D., 2015. Spent coffee grounds: A review on current research and future prospects. Trends in Food Science & Technology 45, 24–36.

Carta, S., Tsiplakou, E., Nudda, A., 2021. Novel by-products in goat nutrition: effect on milk yield and composition. Book of abstract of the 24th congress of the Animal Science and Production Association, 21-24 September 2021, Padova, Italy, p. 73.

- Cechella, J.L., Leite, M.R., Dobrachinski, F., da Rocha, J.T., Carvalho, N.R., Duarte, M.M. M.F., Soares, F.A.A., Bresciani, G., Royes, L.F.F., Zeni, G., 2014. Moderate swimming exercise and caffeine supplementation reduce the levels of inflammatory cytokines without causing oxidative stress in tissues of middleaged rats. Amino Acids 46, 1187–1195.
- Cesarani, A., Gaspa, G., Correddu, F., Cellesi, M., Dimauro, C., Macciotta, N.P.P., 2019. Genomic selection of milk fatty acid composition in Sarda dairy sheep: Effect of different phenotypes and relationship matrices on heritability and breeding value accuracy. Journal of Dairy Science 102, 3189–3203.
- Chedea, V.S., Pelmus, R.S., Lazar, C., Pistol, G.C., Calin, L.G., Toma, S.M., Dragomir, C., Taranu, I., 2017. Effects of a diet containing dried grape pomace on blood metabolites and milk composition of dairy cows. Journal of the Science of Food and Agriculture 97, 2516–2523.
- Choi, Y., Rim, J.S., Na, Y., Lee, S.R., 2018. Effects of dietary fermented spent coffee ground on nutrient digestibility and nitrogen utilization in sheep. Asian-Australasian Journal of Animal Sciences 31, 363.
- Chikwanha, O.C., Vahmani, P., Muchenje, V., Dugan, M.E., Mapiye, C., 2018. Nutritional enhancement of sheep meat fatty acid profile for human health and wellbeing. Food Research International 104, 25–38.
- Correddu, F., Lunesu, M.F., Buffa, G., Atzori, A.S., Nudda, A., Battacone, G., Pulina, G., 2020. Can agro-industrial by-products rich in polyphenols be advantageously used in the feeding and nutrition of dairy small ruminants? Animals 10, 131.
- Correddu, F., Cesarani, A., Dimauro, C., Gaspa, G., Macciotta, N.P.P., 2021. Principal component and multivariate factor analysis of detailed sheep milk fatty acid profile. Journal of Dairy Science 104, 5079–5094.
- Cruz, R., Cardoso, M.M., Fernandes, L., Oliveira, M., Mendes, E., Baptista, P., Morais, S., Casal, S., 2012. Espresso coffee residues: a valuable source of unextracted compounds. Journal of Agricultural and Food Chemistry 60, 7777–7784.
- de Otálora, X.D., Ruiz, R., Goiri, I., Rey, J., Atxaerandio, R., San Martin, D., Orive, M., Iñarra, B., Zufia, J., Urkiza, J., García-Rodríguez, A., 2020. Valorisation of spent coffee grounds as functional feed ingredient improves productive performance of Latxa dairy ewes. Animal Feed Science and Technology 264, 114461.
- Johnston, K.L., Clifford, M.N., Morgan, L.M., 2003. Coffee acutely modifies gastrointestinal hormone secretion and glucose tolerance in humans: glycemic effects of chlorogenic acid and caffeine. The American Journal of Clinical Nutrition 78, 728–733.
- Jorda, A., Portoles, M., Guasch, R., Bernal, D., Saez, G.T., 1989. Effect of caffeine on urea biosynthesis and some related processes, ketone bodies, ATP and liver amino acids. Biochemical Pharmacology 38, 2727–2732.
- Kawai, K., Kuruhara, K., Matano, Y., Akiyama, K., Hashimura, S., Tanaka, S., Kiku, Y., Watanabe, A., Shinozuka, Y., 2018. Effects of Coffee Ground Silage Feeding in Reducing Somatic Cell Count in Bovine Subclinical Mastitis Milk. Asian Journal of Animal and Veterinary Advances 13, 387–392.
- Manikandan, R., Beulaja, M., Thiagarajan, R., Pandi, M., Arulvasu, C., Prabhu, N.M., Saravanan, R., Esakkirajan, M., Palanisamy, S., Dhanasekaran, G., Nisha, R.G., Devi, K., Latha, M., 2014. Ameliorative effect of ferulic acid against renal injuries mediated by nuclear factor-kappaB during glycerol-induced nephrotoxicity in Wistar rats. Renal Failure 36, 154–165.
- Martínez-López, S., Sarriá, B., Mateos, R., Bravo-Clemente, L., 2019. Moderate consumption of a soluble green/roasted coffee rich in caffeoylquinic acids reduces cardiovascular risk markers: results from a randomized, cross-over, controlled trial in healthy and hypercholesterolemic subjects. European Journal of Nutrition 58, 865–878.
- Mele, M., Macciotta, N.P.P., Cecchinato, A., Conte, G., Schiavon, S., Bittante, G., 2016. Multivariate factor analysis of detailed milk fatty acid profile: Effects of dairy

- system, feeding, herd, parity, and stage of lactation. Journal of Dairy Science 99, 9820–9833.
- Mertens, D.R., 2002. Gravimetric determination of amylase-treated neutral detergent fiber in feeds with refluxing beakers or crucibles: collaborative study. Journal of AOAC International 85, 1217–1240.
- Moisey, L.L., Kacker, S., Bickerton, A.C., Robinson, L.E., Graham, T.E., 2008. Caffeinated coffee consumption impairs blood glucose homeostasis in response to high and low glycemic index meals in healthy men. The American Journal of Clinical Nutrition 87, 1254–1261.
- Narasimhan, A., Chinnaiyan, M., Karundevi, B., 2015. Ferulic acid exerts its antidiabetic effect by modulating insulin-signalling molecules in the liver of high-fat diet and fructose-induced type-2 diabetic adult male rat. Applied Physiology, Nutrition and Metabolism 40, 769–781.
- Nudda, A., Correddu, F., Atzori, A.S., Marzano, A., Battacone, G., Nicolussi, P., Bonelli, P., Pulina, G., 2017. Whole exhausted berries of Myrtus communis L. supplied to dairy ewes: Effects on milk production traits and blood metabolites. Small Ruminant Research 155, 33–38.
- Olas, B., Bryś, M., 2019. Effects of coffee, energy drinks and their components on hemostasis: The hypothetical mechanisms of their action. Food and Chemical Toxicology 127, 31–41.
- Rider, L.G., Miller, F.W., 1995. Laboratory evaluation of the inflammatory myopathies. Clinical and Diagnostic Laboratory Immunology 2, 1–9.
- Robertson, J.B., Van Soest, P.J., 1981. The detergent system of analysis and its application to human foods. In: James, W.P.T., Thean-der, O. (Eds.), The analysis of dietary fiber in food. Marcel Dekker, New York, NY, USA, p. 123.
- Salami, S.A., Valenti, B., Luciano, G., Lanza, M., Umezurike-Amahah, N.M., Kerry, J.P., O'Grady, M.N., Newbold, C.J., Priolo, A., 2021. Dietary cardoon meal modulates rumen biohydrogenation and bacterial community in lambs. Scientific Report 11. 1–15.
- Salinas-Rios, T., Ortega-Cerrilla, M.E., Sánchez-Torres-Esqueda, M.T., Hernández-Bautista, J., Díaz-Cruz, A., Figueroa-Velasco, J.L., Guinzberg-Perrusquia, R., Cordero-Mora, J.L., 2015. Productive performance and oxidative status of sheep fed diets supplemented with coffee pulp. Small Ruminant Research 123, 17–21.
- San Martin, D., Orive, M., Iñarra, B., García, A., Goiri, I., Atxaerandio, R., Urkiza, J., Zufía, J., 2020. Spent coffee ground as second-generation feedstuff for dairy cattle. Biomass Conversion and Biorefinery 11, 589–599.
- Santos, G.T.D., Schogor, A.L.B., Romero, J.V., Lima, L.S.D., Matumoto-Pintro, P.T., Grande, P.A., Kazama, D.C.S., Santos, F.S., 2014. Production, composition, fatty acids profile and stability of milk and blood composition of dairy cows fed high polyunsaturated fatty acids diets and sticky coffee Hull. Brazilian Archives of Biology and Technology 57, 493–503.
- Sompong, W., Cheng, H., Adisakwattana, S., 2015. Protective effects of ferulic acid on high glucose-induced protein glycation, lipid peroxidation, and membrane ion pump activity in human erythrocytes. PLoS One 8, e0129495.
- Venn-Watson, S., Lumpkin, R., Dennis, E.A., 2020. Efficacy of dietary odd-chain saturated fatty acid pentadecanoic acid parallels broad associated health benefits in humans: could it be essential? Scientific Reports 10, 1–14.
- Vitaglione, P., Morisco, F., Mazzone, G., Amoruso, D.C., Ribecco, M.T., Romano, A., Fogliano, V., Caporaso, N., D'Argenio, G., 2010. Coffee reduces liver damage in a rat model of steatohepatitis: the underlying mechanisms and the role of polyphenols and melanoidins. Hepatology 52, 1652–1661.
- Zhong, R., Xiao, W., Ren, G., Zhou, D., Tan, C., Tan, Z., Han, X., Tang, S., Zhou, C., Wang, M., 2011. Dietary tea catechin inclusion changes plasma biochemical parameters, hormone concentrations and glutathione redox status in goats. Asian-Australasian lournal of Animal Sciences 24, 1681–1689.