

Human cells involved in atherosclerosis have a sex

Questa è la versione Post print del seguente articolo:

Original

Human cells involved in atherosclerosis have a sex / Franconi, Flavia; Rosano, Giuseppe; Basili, Stefania; Montella, Andrea Costantino Mario; Campesi, Ilaria. - In: INTERNATIONAL JOURNAL OF CARDIOLOGY. - ISSN 0167-5273. - 228:(2017), pp. 983-1001. [10.1016/j.ijcard.2016.11.118]

Availability:

This version is available at: 11388/173553 since: 2022-06-01T09:42:40Z

Publisher:

Published

DOI:10.1016/j.ijcard.2016.11.118

Terms of use:

Chiunque può accedere liberamente al full text dei lavori resi disponibili come "Open Access".

Publisher copyright

note finali coverpage

(Article begins on next page)

Human cells involved in atherosclerosis have a sex

Flavia Franconi^{1,2} Giuseppe Rosano³, Stefania Basili⁴, Andrea Montella², Ilaria Campesi^{2,5*}

¹ *Assessorato alle Politiche per la Persona of Basilicata Region, Potenza, Italy;*

² *Department of Biomedical Sciences, University of Sassari, Sassari, Italy*

³ *Cardiovascular and Cell Sciences Research Institute, St George's University of London.*

⁴ *Department of Internal Medicine and Medical Specialties - Research Center on Gender and Evaluation and Promotion of Quality in Medicine (CEQUAM), Sapienza University of Rome, Italy*

⁵ *Laboratory of Sex-Gender Medicine, National Institute of Biostructures and Biosystems, Osilo, Italy*

Corresponding author: Ilaria Campesi, Department of Biomedical Sciences, University of Sassari,
Via Muroni 23, 07100, Sassari, Italy. E-mail: icampesi@uniss.it

Conflict of Interest The authors report no relationships that could be construed as a conflict of interest.

Acknowledgements: We would like to thank the Mayor of Osilo for the granting of the structure devoted to the National Laboratory of Gender Medicine. IC was supported by a grant from Regione Sardegna “Progetti di Farmacovigilanza attiva, finanziabili attraverso i fondi fv 2008/09”.

Abstract

The influence of sex has been largely described in cardiovascular diseases. Atherosclerosis is a complex process that involves many cell types such as vessel cells, immune cells and endothelial progenitor cells; however, many, if not all, studies do not report the sex of the cells. This review focuses on sex differences in human cells involved in the atherosclerotic process, emphasizing the role of sex hormones. Furthermore, we report sex differences and issues related to the processes that determine the fate of the cells such as apoptotic and autophagic mechanisms. The analysis of the data reveals that there are still many gaps in our knowledge regarding sex influences in atherosclerosis, largely for the cell types that have not been well studied, stressing the urgent need for a clear definition of experimental conditions and the inclusion of both sexes in preclinical studies.

Keywords: sex differences; vessel wall cells; immune cells; atherosclerosis; humans

1. Introduction

Atherosclerosis starts very early in life and becomes clinically relevant only after many years [1]. Compelling evidence links inflammation and adaptive immune responses to atherogenesis [2]. The potential major antigens involved in atherosclerosis are neo-epitopes derived by oxidation reactions of low density lipoprotein (LDL) or when cells undergo apoptotic death [3]. Six different morphological types of atherosclerotic lesions [4] and the involvement of numerous cell types have been described. The cell types include: endothelial cells (ECs), vascular smooth muscle cells (VSMCs) and endothelial progenitor cells (EPCs), which participate in the arterial endothelial damage repairing endothelial dysfunction [5]; and haematogenous cells including monocytes/macrophages and cellular fragments [[6, 7] and quoted literature].

Even though the Institute of Medicine report “*Exploring the Biological Contribution of Sex*” affirms that sex affects all aspects of cellular function from “*womb to tomb*” [8] and that sex and gender could affect the outcome, interpretation and applicability of data [9], studies rarely consider the sex of donors [10], even when the effects of sex hormones are analysed [[11] and quoted literature]. Atherosclerosis exhibits numerous sex differences (SD) [12, 13]. Some of these differences have been attributed to oestrogens [14, 15]. Furthermore, SD have been found in the levels of numerous mRNAs in samples obtained from the aortas of women and men with or without atherosclerotic lesions; for example, peroxisome proliferator-activated receptor (PPAR) γ is more highly expressed in samples from healthy men than in samples from healthy women [16].

Although still at an early stage, some findings show that cells freshly prepared from blood or from blood vessels can be used to study sexual dimorphism [11, 17, 18]. This is relevant because there is no ideal animal model as each has its own advantages and limitations with respect to manipulation of the atherogenic process and modelling human atherosclerosis [19].

Therefore, this review focuses on sex differences in human cells in primary culture, as “*sex memory*” may be lost during culturing. For example, it has been shown that differences in the expression levels of oestrogen receptors (ER) disappear after 14 passages post-isolation [20] and GPR30 expression declines in VSMCs maintained in primary culture (vs. expression in freshly isolated tissue) [21]. The review analyses, for each cell type, the main aspects that are important in the atherosclerotic process, starting from immune cell count and function and including the influence of the menstrual cycle, menopause, and pregnancy when the donor is a woman. The effects of sex hormones in immune cells including the influence of the menstrual cycle on immune cell function have recently been reviewed, [22-25] including the effect of ageing on the influence of sex in immune cells [26]. Therefore, these specific aspects are only summarized here in relation to human cells. Moreover, differences in the expression levels and activities of sex hormone receptors have been reported, as less is known about whether they present sexual disparity at least in human cells [27]. Furthermore, our attention is focused on cell fate, namely apoptosis and autophagy. Apoptosis, a highly regulated cell death process, plays an important role in numerous pathologic conditions involving cardiovascular diseases (CVDs) [28]. Studies have documented apoptosis of VSMCs in atherectomy specimens from atherosclerotic and restenotic lesions, and apoptosis of VSMCs appears to contribute to plaque rupture [29]. Apoptosis has also been observed in macrophages and T cells within atherosclerotic lesions [30]. Increased EC apoptosis may initiate atherosclerosis, as the inducers of EC apoptosis are known risk factors for atherosclerosis [31]. Therefore, it is relevant to know whether apoptosis is a sex-divergent process. On the other hand, autophagy is a process of self-cannibalization of organelles, aggregates, proteins and toxic molecules in the lysosomes. In immune cells, autophagy is also a regulator of inflammation promoting the end of the inflammatory response [32]. Autophagy preserves the health of cells and tissues by replacing outdated and damaged cellular components. In the early stage of atherosclerosis, it appears to protect plaques against stressors through the degradation of intracellular components;

however, it appears to become dysfunctional in advanced stages of atherosclerosis [33] and quoted literature]. However, the role of autophagy in atherosclerosis remains poorly understood, and even more poorly understood is the role of sex in the autophagic processes of the cells involved in atherosclerosis. On the contrary, SD in redox state have been reviewed previously [34, 35] and are not discussed here.

Finally, as a complex interplay between lifestyle, such as smoking, and genetic risk factors is present in atherosclerosis, the effect of smoking on cell count has been considered in view of the fact that increased cell counts are considered risk factors for CVDs, and the pro-atherosclerotic activity of smoking has been considered as smoking is more hazardous for women than for men [36].

2. Human ECs

Endothelial dysfunction plays an early and pivotal role in vascular inflammation and in the pathogenesis of atherosclerosis [37]. In the context of ECs, it is important to note that ECs have significant phenotypic heterogeneity depending on the location and vessel type [5], implicating that SD found in one location cannot automatically be extended to another one. For example, ER β is more highly expressed in human umbilical artery endothelial cells (HUAECs) than in human umbilical vein endothelial cells (HUVECs), whereas ER α expression does not differ between ECs obtained from the two vessels [38].

HUVECs express genes and proteins for ERs and ARs without significant differences between the sexes [11] (Table 1). Regarding ER α and ARs, there are not univocal results because Annibalini et al. [39] reported that male and female HUVECs do not express ER α , while ER β and AR expression is similar (Table 1). Toth and collaborators show that HUVECs of unknown sex lack ER α and PRB but express ER β and PRA [40]. Finally, Kim-Schulze and collaborators [41] show expression of ERs in ECs of unknown sex, whereas human cerebral vascular endothelial cell (HCVECs) express ER β and

GPER1 but there are no reports on ER α expression in HCVECs [42]. PRA and PRB are present in freshly isolated female HUVECs and human dermal ECs [43], but only PRA is found in a commercial line of HUVECs [43]. ARs have been identified in HAECs (human aortic endothelial cells) [43], and other authors found higher AR levels in male-derived ECs versus female-derived ECs [44, 45].

2.1 Function of human ECs

ECs have many functions [46], and their impairment is a risk factor for vascular diseases [47-49]. The activity of healthy endothelium depends on the release of numerous molecules (prostacyclin, endothelium-derived hyperpolarizing factor (EDHF), NO, hydrogen sulfite (H₂S), thrombomodulin, tromboxane, antithrombin, endothelin (ET), Von Willebrand's factor (VWF), fibronectin, etc.) that have regulatory functions on VSMCs, circulating white cells, erythrocytes and PLTs and on vascular permeability [46]. Globally, sex affects the function of ECs. Compared to healthy, age-matched men, healthy women tend to have better endothelial function [50]. Endothelium-dependent vasodilation is higher in healthy females than in males. Endothelium-dependent vasodilation is also higher in females than in males after CAD and is positively associated with ageing [51-53]. Furthermore, obesity and insulin resistance reduce endothelial vasodilatation in both sexes, but this effect is more marked in men [54]. Type II diabetes impairs endothelial vasodilatation above and beyond that observed with obesity alone only in women [54]. Additionally, the vasodilation induced by methacholine, an agonist of muscarinic receptors, which is a trigger of NO and EDHF release, is bigger in premenopausal healthy women than in age-matched healthy men, whereas no difference is observed in diabetic individuals [54].

The effects of sex hormones on endothelial function have been described both *in vivo* and *in vitro*. Endothelial function has been shown to be influenced by the menstrual cycle in healthy women. In particular, endothelium-dependent vasodilation is larger during the luteal phase than during the

follicular phase [55, 56]. Nevertheless, some authors state that NO production is higher during the follicular phase than during the ovulatory phase [57]. This has not been unanimously confirmed, as Jilma and collaborators [58] have indicated that NO production is not influenced by the menstrual cycle. An apparent decline in endothelial function during menopause has been reported [59, 60]. Notably, HRT increases the levels of NO in postmenopausal women [61], indicating the importance of oestrogens in regulating the production of this relevant vasodilator. In line with previous results, Forte [62] shows that the whole-body production of NO is greater in healthy premenopausal women than in men. However, Jilma and collaborators [58] indicate that the NO concentration, such as the level of exhaled NO, is higher in men than in women [63]. In primary cultures of HUVECs, Addis and collaborators [11] show that female HUVECs express significantly higher levels of both the NOS3 gene and protein compared to male HUVECs, indicating a significant sex difference in the ability to produce the potent vasodilator NO. *In vitro*, testosterone induces NO production in HAECs that is associated with activation of NOS3, which in turn is inhibited by incubation with nilutamide, an AR antagonist, or an AR siRNA [[64] and quoted literature]. However, others do not report effects of testosterone on NO production [65, 66]. In HAECs, oestrogens also activate NOS3 [[64] and quoted literature]. In fact, it is not known whether H₂S signalling is influenced by sex, but its plasma levels are higher in healthy men than in women. This is not surprising because its synthesis is under the control of androgens [67] and OCs reduce the plasma levels of its precursor, cysteine [68].

In vitro, dihydrotestosterone induces VCAM1 expression, increasing monocyte binding to the endothelium. Furthermore, dihydrotestosterone triggers the proliferation of HAECs, probably through the up-regulation of the expression of vascular endothelial growth factor A and cyclins A and D1 [69]. The stimulation of EC proliferation in the CV system by activation of AR could contribute to the repair of EC injury/damage [[64] and quoted literature].

The potent vasoconstrictor ET1 is produced by vascular ECs as well as by many other tissues, and the endothelin system is up-regulated in a number of pathological conditions associated with endothelial dysfunction including atherosclerosis [70]. The endothelin system appears to act in a sex-specific manner. Plasma ET1 levels are higher in men than in women, and pregnant women have lower levels compared with non-pregnant women [71]. HRT increases the levels of ET1 in postmenopausal women [61]. The previous data suggest a role of sex hormones in the regulation of the endothelin system. This is confirmed by the fact that oestradiol reduces basal and stimulated ET1 secretion from HAECs. Conversely, testosterone induces an increase in the number of cells that secrete ET1, up-regulating ET1 mRNA [72]. Another vasoconstrictor, thrombin, is more active in female HUVECs than in male ones in stimulating prostacyclin and prostaglandin E₂ synthesis [73].

Finally, Lorenz and collaborators [74] report different transcriptional profiles in HUVECs from male and female donors, with 70 genes being differentially expressed between the sexes. For example, genes related to immune responses (humoral, innate and inflammatory responses) and some genes involved in metabolism (for example, leptin, insulin receptors and some apolipoproteins) are higher in female than in male HUVECs, as is the viability after serum starvation and tube formation capacity. These results indicate that some SD reside in autosomal genes.

In HUVECs, 17 β -oestradiol increases the expression and activity of SR-BI/CLA-1, which facilitates the cellular uptake of cholesterol from HDL [154]. Moreover, after shear stress induction, a higher number of genes are up- or down-regulated in female HUVECs than in male ones; for example, VCAM-1 expression is down-regulated almost 22 times in female HUVECs and only 3.5 times in male HUVECs [74]. Finally, unstimulated male HUVECs release more MCP-1 and IL-8 than female HUVECs [75].

2.2 Human ECs and cell fate

As mentioned above, SD in death pathways receive little attention. Recently, the use of freshly isolated cells from male and female individuals provided information on sexual dimorphism in cell fate control. In ECs, SD regard either apoptosis or autophagy. SD in the induction of cell death may, to some extent, explain the disparity between the sexes in many human diseases including atherosclerosis.

2.2.1 Apoptosis

EC apoptosis may be relevant for endothelial injury, including atherosclerosis, and it may lead to endothelial disruption. Notably, oxidized LDL (oxLDL) induces apoptosis and autophagy (see below) in HUVECs of unknown sex [76]. Intriguingly, inhibition of apoptosis occurs from an inhibition of autophagy, whereas inhibition of autophagy occurs with an increase in apoptosis [77]. Sex hormones influence apoptotic processes. For example, testosterone enhances TNF α -induced apoptosis after serum deprivation in HUVECs [78], whereas, in human coronary artery ECs, 17 β -oestradiol increases apoptosis and elevates the pre-apoptotic Fas mRNA and protein expression, the expression of FasL mRNA and the secretion of FasL protein [79]. At least in experimental animals, epoxyeicosatrienoic acids, the activity of which is controlled by soluble epoxide hydrolase, are more highly expressed in male cerebral vessels compared with female cerebral vessels [80], and they seem to protect ECs from ischaemic injury [81]. Genistein, an isoflavone that is structurally similar to oestradiol, has pro-apoptotic activity in human coronary ECs obtained from middle-aged women [82]. Finally, RLIP76, a Ral effector GTPase-activating protein, significantly alters the percentage of apoptosis only in female HUVECs [83].

2.2.2 Autophagy

Autophagy, or type II programmed cell death, might also be involved in the progression of atherosclerosis [84]. In ECs from atherosclerotic plaques obtained during carotid endarterectomy in

individuals with acute ischaemic stroke and severe carotid artery stenosis, an increase in autophagy compared with normal arteries is observed [85]. In HUVECs of unknown sex, autophagy is activated by OxLDL through the LC3/beclin1 pathway causing vascular endothelial dysfunction [76]. The increase in autophagy is blocked by 3-methyladenine, an inhibitor of autophagic response, and is increased by the autophagy inducer rapamycin [76].

A recent study characterizes the phenotype of HUVECs according to the sex of the umbilical cord donor and individuates that male cells have a higher degree of constitutive autophagy than female ones as they express higher levels of Beclin-1 and have a higher LC3-II / LC3-I ratio [11]. These results have been confirmed by ultrastructural analysis showing a higher build-up of autophagic vacuoles at different stages in male HUVECs than in female ones. However, the protein expression levels of mTOR and AKT, critical regulators of autophagy, are not different between the sexes. The tendency of male HUVECs to undergo autophagy could depend, at least in part, on increased oxidative behaviour compared with female HUVECs [11].

Another important function of ECs is the secretion of VWF, which is required for PLT adhesion to the vessel wall. EC autophagosomes contain abundant VWF protein, and its secretion is inhibited *in vitro* by inhibitors of autophagy, suggesting that inhibition of autophagy can prevent thrombosis [86]. In HUVECs, angiotensin II progressively increases autophagy, apoptosis and senescence, which are reduced by valsartan. Additionally, autophagy has an early protective effect on vascular endothelial damage due to Ang II [87], and autophagy induced by hyperglycaemia [88, 89] may have a protective effect versus senescence and apoptosis. However, it is not yet known whether the protective effect of autophagy is influenced by sex.

3. Human VSMCs

The medial layer of the vascular wall is composed of VSMCs and the synthesized extracellular matrix, and it plays a key role in the maintenance of vascular structure and function [90]. Notably, VSMCs may trans-differentiate into other cell phenotypes such as macrophages, which can lead to the formation of foam cells [[91] and quoted literature].

Very little information exists on the effect of sex on human VSMCs as the majority of studies are performed using animals as cell donors. However, it is known that ER α , ER β , and PR mRNAs and proteins (Table 1) are present in cells from different veins and arteries such as the coronary arteries, saphenous veins, aortas, mammary arteries, and iliac arteries obtained from both men and women [92, 93]. When sex is considered, it has been found that ER β mRNA is predominantly expressed in female VSMCs obtained from the coronary artery and saphenous vein [94]. ER β appears to be more highly expressed in women who have not undergone HRT versus users of HRT, whereas ER α levels do not differ among cohorts. Furthermore, in HRT users, only the expression of ER α declines with age [95]. In aortas obtained by autopsy, the expression of ER decreases with the level of atherosclerosis. No change is reported in men [96].

Finally, human umbilical artery smooth muscle cells from male and female neonates display sexual dimorphism in ER β expression, with ER β being more highly expressed in male-derived cells, while ER α is similarly expressed in both sexes (Table 1) [97]. ER α is also localized in human VSMCs starved for five days to allow for ER α up-regulation, and ER α is more highly expressed on average in VSMCs from female donors than in VSMCs from male donors [98]. In human aortic VSMCs, PRB is similar in male- and female-derived cells, but PRA is lower in male-derived cells than in female-derived cells [99]. The GPER has also been identified in cultured VSMCs [43]. PRs have been identified in VSMCs. In particular, PRB is equally expressed in men and women, while PRA is more highly expressed in vessels obtained from postmenopausal women [43]. In VSMCs prepared from human internal mammary arteries, binding and physiological studies confirm the presence of ARs

[100]. Globally, sex hormone receptors are present, but they exhibit dishomogeneous expression in the VSMCs of different arteries.

3.1 *Function of human VSMCs*

Considering the presence of sex hormone receptors, it is not surprising that sex hormones affect the function of human VSMCs. Oestrogens have similarly antiproliferative effects on male and female human aortic smooth muscle cells [101], acting through ER β , which is the prevalent isoform and mediates vasodilation and VSMC relaxation [102]. Moreover, *in vitro*, female sex hormones (oestradiol and progesterone) are able to reduce collagen deposition in female aortic VSMCs much more than testosterone, while testosterone up-regulates gene and protein expression of matrix metalloproteinase 3 [103]. Steroid sulfatase (STS) and oestrogen sulfotransferase (EST) are involved in the metabolism of oestrogens and are both present in aortic VSMCs. In women, STS expression declines with the level of atherosclerosis, while the decline is not evident in men. EST expression is significantly higher in male aortas than in female aortas with mild atherosclerotic changes. In women, EST is more highly expressed when the aorta exhibits severe atherosclerotic damage [104].

3.2 *Human VSMCs and cell fate*

Atherosclerotic lesions result from a dynamic interplay involving proliferation, autophagy and apoptosis in response to injury of the ECs and VSMCs of the artery wall [84, 105]. However, it has been suggested that autophagy can contribute to attenuation of inflammation through the eradication of damaged molecules and cellular organelles, thus preventing apoptosis of VSMCs and stabilizing the lesion [106]. Excessively stimulated autophagy may also play a detrimental role in plaque formation [106]. In this context, it is fundamental to know the influence of sex to optimize the therapeutic approach in women.

3.2.1 Apoptosis

Apoptosis and cell death are frequent events in the initial formation of plaques as well as in more evolved plaques. In the thin fibrous cap of advanced lesions, apoptosis enhances plaque rupture, thereby triggering thrombosis and myocardial infarction and contributing to plaque instability and sudden coronary death [107, 108]. Importantly, cells undergoing apoptosis need to be efficiently removed from atherosclerotic lesions because their permanence is a potent inducer of the coagulant cascade [109, 110]. Recently, it has been that mitophagy has a role on human VSMC apoptosis induced by oxLDL suggesting that mitophagy is a safeguard mechanism versus apoptosis [111]. Nevertheless, the importance of apoptosis in VSMCs and the influence of sex on apoptosis of human VSMCs have still not been studied, although animal studies suggest that there are significant SD [112-114].

3.2.2 Autophagy

VSMCs have the capacity to assume different phenotypes and this phenotype flexibility needs the integration of transcriptional, metabolic, and ultra-structural programs and in doing this autophagy seems to assume the role of main coordinator [115]. For example platelet derived growth factor BB (PDGFBB) induces autophagy reducing the transition to the synthetic VSMC phenotype and elevating cell survival in condition of increased oxidative stress [115], whereas other stimuli such as angiotensin II promotes cell death [115].

It is believed that autophagy plays an important role in the aetiology of atherosclerosis. However, the molecular basis of autophagy in the pathogenesis of atherosclerosis remains poorly understood, and even less is known about how the sex influences the process. In VSMCs obtained from atherosclerotic plaques from carotid endarterectomies in individuals with acute ischaemic stroke and severe carotid

artery stenosis, an increase in autophagy compared with normal arteries was observed [85]. Some authors suggest that an increase in autophagy is relevant to maintaining plaque stability. In the atherosclerotic plaques, autophagy may play a protective role [116], and cell death induced by low doses of statins may be reduced by the autophagy inducer 7-ketocholesterol, at least in rabbit aortic VSMCs [117], suggesting that stimulation of autophagy could protect VSMCs from death. However, excessive autophagy may cause autophagic death of SMCs, which conversely results in plaque destabilization. In this context, it is relevant to consider that human umbilical artery smooth cells from male neonates have higher levels of Beclin-1 than female cells, while the LC3-II / LC3-I ratio is similar between the sexes [97, 118]. Beclin-1 has a role in the crosstalk between autophagy and cell proliferation [89]. Therefore, the sex difference observed may be of interest. When autophagy (measured as the LC3-II / LC3-I ratio) is induced by stimuli such as verapamil, starvation and rapamycin, the autophagy programme is different in male and female human umbilical artery smooth muscle cells [118]. Importantly, the SD are stimulus specific. In brief, serum starvation-induced autophagy is more pronounced in female human umbilical artery smooth muscle cells than in male cells, while 250 nM rapamycin produces autophagy only in female cells [118]. Moreover, verapamil-induced autophagy does not exhibit sex differences, even if Beclin-1 is increased in female cells. Finally, PmTor does not differ under basal conditions, but it is significantly down-regulated by starvation in female human umbilical artery smooth muscle cells and by rapamycin in both male and female cells [118]. The SD in autophagy indicate sex-based differences in the pharmacodynamic effects of verapamil and rapamycin. Autophagy in VSMCs may be considered a housekeeping process [89] that provides protection against cell death in several vascular diseases, whereas excessive autophagy would be deleterious. In particular, the defective autophagy of VSMCs may cause irreversible cellular senescence and subsequently contribute to atherosclerosis progression and other vascular disorders. Therefore, more sex studies of this specific point are urgently needed.

4. Human PLTs

These small circulating cell fragments play a pivotal role in haemostasis and endothelial repair and are also involved in atherogenesis and thrombosis [119, 120]. There is both indirect and direct evidence that sex hormones affect human PLT biology.

ER α and ER β have been found in circulating PLTs [121, 122], with similar expression levels in the two sexes (Table 1) [122]. These receptors can exert non-genomic activity having a pro-aggregating effect through ER β [123]. AR has been detected in human PLTs [121].

4.1 Human PLT count and morphology

Human PLT count is influenced by sex. Women have a higher PLT count than men [124-130]. The average PLT count is also influenced by age and ethnicity [127, 128, 130]. Notably, the SD observed with HRT and OC use in adults are still present in the elderly [68, 128-131]. It is possible that the higher haematocrit values observed in men versus women, with relatively less plasma and greater *in vitro* dilution due to the addition of anticoagulant solutions, lead to the erroneous measurement of lower platelet counts in men compared with women [132, 133]. In pregnant women, there is a light thrombocytopenia due to increased platelet consumption caused by the increase in the utero-placental circulation [134]. Women with acute strokes have elevated PLT counts, although PLT reactivity is lower in women than in men [125].

In addition, SD are also detected in PLT morphology, with the mean PLT volume being higher in women than in men [126, 135].

4.2 Human PLT count and smoking

Studies do not provide univocal results regarding smoking and PLT count [128, 136-138]. It has been reported that female smokers have lower PLT counts than non-smokers, while this difference is not detected in men [128]. However, others report that smoking increases PLT count in fertile women (in the follicular phase) but not in men [136].

4.3 Function of human PLTs

Sex effects on the functions of PLTs are present, and they have recently been reviewed [[132, 139] and quoted literature]. Briefly, women have more receptors for fibrinogen [132], bind more fibrinogen after adenosine diphosphate (ADP) and present more spontaneous aggregation than men [[132, 139] and quoted literature]. Furthermore, PLT degranulation measured as CD62P expression and activation of GPIIb/IIIa measured as PAC-1 binding do not exhibit sexual dimorphism. However, when PLTs are exposed to ADP, female PLTs appear to be more activated and undergo major morphological alterations. The PLTs obtained from women produce a significant increase in phosphorylated protein kinase substrates such as RhoA and phosphorylated myosin light chain [140]. Lawrence and collaborators [133] also observed more spreading and adherence in men than in women.

Furthermore, in women without CAD, PLTs are more reactive to ADP and to thrombin receptor agonist than in men [141, 142]. At rest, PLTs of women with CAD respond better to serotonin and epinephrine compared with those of men [[132, 139] and quoted literature]. Also protease-activated receptor-mediated PLT reactivity is increased in females [143]. Finally, following mental stress, women have higher collagen-stimulated PLT aggregation responses than men [144]. SD are also measured with a test that measures both PLT adhesion and aggregation (primary haemostasis). A Korean study of healthy men and women reported that women have a significantly higher closure time than men [145]; however, this is not an univocal result [146].

SD taking into account drug response have previously described and were recently reviewed [[132, 147] and quoted literature]. The Genetic Study of Aspirin Responsiveness performed in healthy men and women revealed higher PLT reactivity in women than in men for numerous agonists [[132] and quoted literature], and this higher reactivity in women is still present after the administration of low doses of aspirin when collagen or ADP are used as agonists [[132] and quoted literature]. In contrast, SD disappear in subjects treated with clopidogrel or with both aspirin and clopidogrel [[132] and quoted literature].

Recombinant human erythropoietin (rhEPO) increases PLT aggregation induced by ADP with some SD, especially when collagen is the agonist. In fact, rhEPO increases and decreases collagen aggregation in males and females, respectively. In addition, rhEPO increases the PLT membrane glycoprotein complex $\alpha 2\beta 3$ in both sexes suggesting that homotypic (PLTs-PLTs) aggregation occurs without significant SD. Conversely, membrane P-selectin, an index of heterotypic aggregation (PLTs-leukocytes), is increased by rhEPO only in women. These results indicate that the prothrombotic risk observed with rhEPO occurs in a sex-specific manner, with females being more sensitive [148].

Activated PLTs participate in inflammatory responses that further amplify their response and endothelial activation during plaque rupture. Activated PLTs release numerous molecules with inflammatory and mitogenic activities into the local microenvironment [149], enforcing the recruitment of monocytes and their differentiation into macrophages [150]. The formation of leukocyte-PLT aggregates in patients undergoing angioplasty and stenting is more pronounced in females than in males, whereas the expression of P-selectin and GPIIb/IIIa does not differ significantly between the sexes [139]. In this context, a special role is played by TLRs, which appear to be more highly expressed in women than in men and which interact with the immune system engaging the neutrophil population [[132] and quoted literature] and favouring PLT-leukocyte aggregates. Indeed, in women, TLR expression is associated with soluble P selectin [151]. In contrast, TLR expression in men is more

frequently associated with soluble TNF α receptor-1 and ICAM1 [151]. Notably, the expression of TLR1, TLR3, TLR6, and TLR7 is associated with body mass index in women. However, in men, the expression of TLR5, TLR7, and TLR10 is associated with the total cholesterol to high-density lipoprotein ratio [151]. Brain-derived neurotrophic factor (BDNF), a nerve growth factor, is also stored in human PLTs [152] and plays a critical role in the CV system [153]. Interestingly, PLT BDNF levels are lower in healthy women than in men [154]. The opposite is observed in individuals with stable coronary artery diseases [155].

Some SD depend on sex hormones, however the effect of the female menstrual cycle on platelet activity is not clear. Some studies show that PLTs bind more fibrinogen during the luteal phase than during the follicular phase [142, 156], whereas PLTs closure time is higher during the follicular phase than during the luteal phase [157], and others show that PLTs adhesion has a biphasic peak during the menstrual cycle [158]. However, other studies find no evidence of any associations among PLTs activity and menstrual cycle phase [159-161]. OCs are able to activate PLTs [162], and some of the effects of OCs depend on the androgenic property of progestin [157]. Furthermore, OCs elevate factor VII levels, PLTs activity, and levels of fibrinogen and plasminogen activator inhibitor-1 [163]. However others do not find any influences of OCs [159-161]. HRT leads to an increase in circulating and activated PLTs in users than in non-users [164], leaving the mean PLTs volume and aggregation rates unchanged [165]. During the third trimester, pregnancy elevates PLT aggregation and the concentration of coagulation factors and decreases fibrinolytic capacity [166]. Pregnancy also elevates the activity of calcium adenosine triphosphatase, beta-thromboglobulin and platelet factor-4, suggesting increased activation of PLTs [167].

Very recently, a more direct role of PLTs in the atherosclerotic process has been reported. Specifically, PLTs can internalize lipids and promote the formation of foam cells [168, 169]. It is not known whether these processes are sex dependent.

Generally, PLT reactivity is enhanced in women; however, this is not an univocal result. Additionally, SD also depend on the stimulus applied. Unfortunately, the paucity of data does not permit us to individuate the clinical relevance of these SD in view of the fact that the sex effect may also be dependent on hormonal status.

4.4 Human PLsT apoptosis and autophagy

PLTs have the ability to undergo apoptosis, which can be induced by a multitude of stimuli. Increased apoptosis results in thrombocytopenia, bleeding disorders and the induction of microparticles [170]. PLTs also have autophagy machinery. Constitutive autophagy is present in resting PLTs and is elevated in response to PLT activation. Inhibition of autophagy during megakaryocyte differentiation reduces PLT formation, and affect PLT function, whereas inhibition of autophagy in mature megakaryocytes induces abnormal PLT activation [171]. It seems that basal autophagy playing an important role in normal PLTs activation [172]. Indeed in diabetic PLTs the induction of mitophagy plays a beneficial role versus oxidative stress, and its absence increases thrombosis [172]. However, the role of constitutive autophagy and mitophagy is not yet clearly understood [173]. In fact, it is not known whether apoptosis and autophagy are influenced by sex.

5. Human EPCs

Endothelial dysfunction is the first step of atherogenesis, and it has been proposed that the lack of or small number of EPCs is a limitation that can lead to endothelial dysfunction [174]. Circulating male and female EPCs express classical ER α and ER β [17], with similar levels of the receptor protein in male and female EPCs (Table 1). However, the literature is not univocal because some reports indicate that cultured human male EPCs express ER α but not ER β [175], while a significant sex difference in mRNA levels for ER α and ER β is reported by Fadini et al. [18], with men having the highest levels.

The receptor expression is influenced by hormonal fluctuation. In particular, the expression of ER β is higher in premenopausal women versus men (Table 1). Furthermore, ER β expression is higher during the ovulatory phase in comparison with the follicular and luteal phases, as well as in comparison with postmenopausal women. Meanwhile, ER α is expressed similarly in both sexes, and, in women, ER α is not influenced by variation in sex hormones [176]. The presence of ARs has been demonstrated by immunohistochemistry and by the measurement of AR mRNA and protein expression [177], but it is not known if AR is influenced by sex.

5.1 Human EPC counts

EPCs are estimated to comprise 0.02-0.2% of blood cells [178, 179]. It has been reported that the number of EPCs is higher in young fertile and pre-hypertensive women than in healthy and pre-hypertensive age-matched men [18, 180-182]. However, this is not an univocal result because one study [183] does not report SD. Additionally, some authors do not observe a significant difference in the number of EPCs between middle-aged women and men [18, 180, 184, 185], indicating that SD could be age dependent. The number of EPCs depends on numerous factors such as rhEPO, growth hormones, and some medications (statins and angiotensin converting enzyme inhibitors) [186-188]. Notably, Pelliccia et al. (2009) reported that postmenopausal women and men with CAD have the same number of EPCs [189]. The previous results suggest that the number of EPCs is regulated by sex hormones. Considering that a reduced number of EPCs is linked with the incidence of CVD [174], it would be clinically relevant to know the reference value of EPCs with respect to the hormonal phase of women.

5.2 EPC count and smoking

The effects of smoking on EPCs are complex and do not appear to be univocal. Yue et al. [190] found that circulating levels of EPCs are significantly lowered in smokers with CAD compared to controls and non-smokers with CAD. In patients with CAD, Werner et al. report a positive association between smoking and high baseline levels of EPCs [191]. Similarly, Mobarrez et al. [192] show that CD34⁺ cells increase after smoking one cigarette. Other authors have shown that the number of EPCs is directly proportional to the number of cigarettes smoked [193]. Others found that smoking does not affect the EPC count [183]. Unfortunately, sex was not considered in the last two studies mentioned.

5.3 Function of human EPCs

The ability of EPCs to migrate and form colonies *ex vivo* is an independent marker of vascular health [194]. In basal conditions, EPCs obtained from young adult men and women do not exhibit any significant differences in the number of migrated cells [17]. However, Hoetzer et al. [194] show that EPC migration is higher in middle-aged women (58 years old) than in age-matched men, suggesting that age could influence the sex effect. Migration appears to be affected by oestrogens. However, conflicting results have been reported regarding the effect of oestradiol *in vitro*. Fadini and colleagues [18] observe that oestradiol elevates the number of colonies in males with a smaller effect on female EPCs, while others [17] found that oestradiol, at physiological concentrations, inhibits EPC migration by approximately 50% in premenopausal women but not in young men. Another compound with oestrogenic properties such as bisphenol A decreases migration only in female cells [17]. Globally, in male EPCs, migration appears to be less sensitive to oestrogenic compounds. In line with previous data, colony formation and migration are influenced by the phase of the menstrual cycle, menopause and HRT [18, 194, 195], again suggesting the involvement of oestrogens at least in females. Recently, it has been shown that patients with hypogonadism have a low number of EPCs, and testosterone replacement therapy can increase the number of circulating EPCs in men [196] indicating that

androgens could also play a role. In view of the fact that mobilization of EPCs contributes to increased neovascularization, which could play a role in the prevention of CVDs [197], the lower migratory capacity of female EPCs could help to explain the higher prevalence of endothelial dysfunction in women [198, 199]. In pre-hypertensive premenopausal women, the activity of circulating EPCs is better preserved than in pre-hypertensive men, with the number and activity of EPCs partially associated with enhanced NO production, whereas vascular endothelial growth factor or granulocyte macrophage colony-stimulating factor are not varied [182].

5.4 Human EPC and cell fate

To the best of our knowledge, there is no data regarding sex influences on autophagy and apoptosis in circulating human EPCs.

6. Human leukocytes

Leukocytes produce numerous chemokines and cytokines and express receptors for these molecules. Complex chemokine and cytokine interactions promote different pathways involved in all stages of atherosclerosis [200]. There is a paucity of research on the influence of sex on this specific point. However, a small study [201] showed that the expression of the chemokine (C-C motif) ligand 5 (CCL5) is higher in women than in men. This is similar to sCD40L [202], and people with high levels of sCD40L are considered to have high cardiovascular (CV) risk [203]. Women have higher expression levels of platelet-derived growth factor (PDGF)-AA and PDGF-BB than men [201]. Ageing modifies the expression of chemokines and cytokines, and this appears to occur in a sex-specific way: macrophage-derived chemokine is, for example, age-dependent in women but not in men [202]. Indeed, the most common polymorphisms of the fractalkine receptor (CX3CR1) gene, T280M and V249I, increase waist circumference in a sex-specific manner [204]. Women with two copies of the

T280M and V249I alleles have higher waist circumferences than women with one copy of the T280M and V249I alleles, while men with one copy of T280M exhibit a higher waist circumference [204].

The human X chromosome encodes many immune-regulatory genes such as TLRs 7 and 8. Fifteen percent of X-linked genes escape silencing, resulting in the increased expression of certain gene products in females as compared to males [26]. In addition, the X chromosome is rich in micro-RNAs (miRNAs) [26]. This is particularly relevant to SD.

6.1 Human leukocyte count

The total leukocyte count is lower in men than in women [205]. In particular, in women, the count varies with the menstrual cycle; it is higher around the 8th to 10th days of the menstrual period than around the 22nd to 24th days [206]. The global leukocyte count is considered an inflammatory marker and a risk factor for CV events [207]; it is associated with CV mortality in both sexes and with non-CV mortality in women [207]. Longevity is also associated with a lower, yet still normal, count, and this correlation is more pronounced in men than in women [208].

Therefore, it is necessary to be aware of the menstrual phase and to build up reference values for the single menstrual phase before performing leukocyte counts. The availability of these data would improve the comparison between men and women and would help to ameliorate the evaluation of CV risk in women.

6.2 Human leukocyte count and the smoking effect

Smoking affects the leukocytes count. In particular, the leukocyte count is higher in current smokers than in non-smokers [136, 205, 209]. Indeed, it is still not clear if there are significant SD because the leukocyte count has been found to be equal between sexes or higher or lower either in men or in

women [124, 209-215]. Notably, smoking increases the total leukocyte count after 20 weeks of pregnancy [216]. The above results highlight the mandatory and urgent need to conduct sex and gender research, at least for CV risk factors, in order to develop evidence-based CV prevention strategies.

7. Human neutrophils

Neutrophils comprise most of the circulating leukocytes and are pivotal in the defence against microorganisms. Historically, they have received little attention in the context of atherosclerosis; however, recent findings show that, among white blood cells, neutrophils are the strongest predictors of coronary heart disease [217].

In both men and women, neutrophils express ER α and ER β . In women, ER α and ER β are up-regulated in the ovulatory phase of the menstrual cycle (Table 1), and, *in vitro*, both ERs are up-regulated by 17 β -oestradiol in neutrophils obtained from premenopausal women [218]. In neutrophils obtained from men, only ER α is up-regulated by incubation with 17 β -oestradiol [218]. Regarding progesterone receptors (PRs), it has been described that intracellular PRs are not present in neutrophils (Table 1) [27]. Finally, the androgen receptor (AR) is expressed in neutrophil lineages from the proliferative precursors to mature neutrophils with no significant SD (Table 1) [219].

7.1 Human neutrophil count

In humans, increases in leukocyte blood counts, most notably increases in neutrophils, have been directly related to the endothelial dysfunction associated with ageing and a growing susceptibility to CVDs [220]. In line with previous results, positive correlations among neutrophil blood counts and CVDs, CV mortality, and all-cause mortality are described [221-224]. Girls have higher neutrophil counts than boys [225]. However, in adults, there are ambiguous results regarding neutrophil counts. The majority of authors have found that they are higher in women than in men [205, 226, 227], while some authors have found that they are similar between the sexes [203]. The discrepancy could arise

from the fact that this last study included only women in the follicular phase [136]. This discrepancy could be due to the relevant increase in neutrophil count observed in the luteal phase as compared to the follicular phase of the normal ovarian cycle [228]. Pregnancy modifies the granulocyte count [229]. Neutrophilia and a delay in apoptosis are observed in at-term pregnancies [230-232]. The above data suggest that the neutrophil count is dependent on sex hormones. However, the effect of exogenous hormones (oral contraceptives, OCs) is not clear, as OCs may or may not increase granulocyte numbers, depending on the type of OCs used [68, 228].

7.2 Human neutrophil count and smoking

Smoking affects the number of neutrophils [205]. The neutrophil counts of current smokers are higher than those of individuals who never smoked especially in Caucasian and Mexican-American smokers [205]. Moreover, a recent study suggests that this occurs only in men, at least in Caucasian smokers [136].

7.3 Human neutrophil function

The role of neutrophils in atherosclerosis was reported in a recent review [[233] and quoted literature]. Importantly, neutrophils are present at the sites of plaque erosion or rupture [234, 235]. Notably, there are fewer neutrophils in plaques in women than in men [236]; however, women with acute stroke have higher neutrophil activity [125].

Although the influence of sex was not considered in studies involving the activation of neutrophils, some SD emerged. In particular, the production of cytokines is higher in male cells versus female ones [237-241]. Notably, SD can depend on the stimulus and can vary according to the considered cytokine [242].

The cascade of arachidonic acid is pivotal in inflammation, and leukotrienes (LTs) are formed in atherosclerotic lesions. In human neutrophils and monocytes, the key enzyme in the synthesis of LTs, 5-lipoxygenase (5-LO), is regulated by androgens, resulting in the sex-specific formation of LTs [243]. It is now becoming clear that LTs are formed more predominantly in the whole blood or in neutrophils isolated from women than in those from men. This appears to be linked to extracellular signal-regulated kinases (ERKs), which are regulated by androgens [244], indicating that LT-induced recruitment and activation of immune cells, the induced proliferation of VSMCs and endothelial dysfunction [245] may be influenced by sex.

The neutrophils of men and women express nitric oxide synthase 1 (NOS1), which is lower in neutrophils from males than in neutrophils from females [246]. In women, the expression of NOS1 is higher during the ovulatory phase versus that measured during the first 2 days of the follicular phase in the very same donor [246]. Hormone replacement therapy (HRT) increases NOS1 protein levels and greatly induces the production of nitric oxide (NO) [246]. In neutrophils from males, *in vitro* exposure to oestrogens increases NOS1 expression, and this increase is inhibited by tamoxifen (an antiestrogenic drug, widely used for the treatment of ER α -positive breast cancer) and ICI 182780 (an ER antagonist with no agonist effects which downregulates cellular levels of both the ERs and PRs). The increased expression of the NOS1 protein is associated with a reduction in the adhesion activity of these cells, which is blocked by NO inhibitors [246].

Neutrophils from females have more β -adrenergic receptors than neutrophils from males. In line with these results, the non-selective agonist of β -adrenergic receptors, isoprenaline, elevates the chemokinesis and the release of a chemotactic factor from neutrophils from females, while it is inactive in neutrophils from males [247]. On the contrary, isoprenaline inhibits IL8-induced chemotaxis, and, again, the effect of isoprenaline is sex-dependent [247]. These results suggest that stressors could have different sex effects.

Finally, CD11b expression and redox activity are similar at baseline between neutrophils from males and neutrophils from females [248]. CD11b expression is not significantly altered by physiological concentrations of oestradiol and progesterone in neutrophils from women [248]. The differences in redox state have already been discussed by Malorni and collaborators [34].

7.4 Human neutrophils and cell fate

The lifespan of neutrophils is tightly regulated, and programmed cell death participates to efficiently resolve inflammation [249]. In neutrophils, autophagy can also play a role in infectious and inflammatory diseases [250], participating in the elimination of endogenous materials and cytosol-colonizing microbes (xenophagy) [251]. Therefore, there is an urgent and mandatory need to determine the influence of sex on cell fate.

7.4.1 Human neutrophils and apoptosis

In these cells, constitutive apoptosis is reduced in women versus men [248]. Indeed, physiologic concentrations of oestradiol and progesterone delay constitutive apoptosis in both sexes but do not change Fas antibody-induced apoptosis, which is essential for the elimination of activated immune cells from the peripheral circulation. However, serum sFas (inhibitor of apoptosis) is significantly higher in men than in women, while sFasL (stimulator of apoptosis) and cytochrome c levels (released from cells during apoptosis) are lower in men than in women [252]. Notably, androgens have no effect [248]. The paucity of data on the effect of sex indicates that it is imperative to study SD in constitutive and induced apoptosis to clarify the impact of these processes on atherosclerotic diseases.

7.4.2 Human neutrophils and autophagy

Some data suggest that autophagy in white blood cells is associated with coronary artery disease (CAD). The LC3 (marker gene for autophagy) and LC3-II (a membrane marker for autophagosomes and autophagolysosomes) genes are significantly reduced in the white blood cells of patients with CAD versus controls; however, a multivariate analysis indicates that only reduction in gene expression is linked with CAD [253]. LAMP-2 gene (lysosomal membrane marker gene) expression in leukocytes is increased in patients with CAD versus controls. Unfortunately, these studies did not conduct an analysis of sex differences.

8. Human monocytes and macrophages

Monocytes represent 5-10% of the peripheral blood mononuclear cells and are progenitors of macrophages and DCs [25]. The contribution of monocytes and macrophages to atherosclerotic progression and the maintenance of vessel-wall inflammation is well established [254].

The data regarding ER expression on monocytes is controversial. Phiel and colleagues [255] found that monocytes express from low to undetectable levels of ERs; whereas Pelekanou and colleagues [256] showed that expression of α 36-kDa splice variant and G-protein coupled receptor 30/G-protein ER occurs in a sex-divergent manner. ER β appears to be the main receptor in monocytes [43]. Intracellular PRs are not present in resting monocytes (Table 1) [27]. AR expression on human monocytes (Table 1) is higher in young men than in young women [257], but this SD in expression disappears with age [43]. Human macrophages (Table 1) express both ER α and ER β [68, 258, 259]. Notably, the ER profile is not modified during differentiation into macrophages or DCs, and it is shared by macrophages present in the atherosclerotic plaque [256]. Some authors have shown that ER α is the predominant receptor on macrophages; whereas, ER β is the predominant receptor on monocytes, and it is up-regulated by oestrogen only in macrophages [43]. Human macrophages also express GPER and PRs [43, 260]. The levels of ERs are higher in monocyte-derived macrophages (MDMs) from women taking OCs versus

non-users. In both OC users and non-users, ER α is inactive [68], while ER β is more highly expressed and more active in untreated women.

In MDMs obtained from premenopausal women, the AR mRNA levels are lower than in men; MDMs from postmenopausal women have AR mRNA levels lower than both men and premenopausal women [257]. In both cases, receptor expression is lower than that in male-derived cells [43]. Indeed, in MDMs obtained from young individuals, AR protein expression does not exhibit sexual dimorphism, and, in MDMs obtained from women, the use of OCs does not influence AR expression [68].

8.1 Human monocyte count

The increase in monocyte count is associated with an increase in the risk for heart disease [261], and it is also an independent predictor of future plaque formation [262]. In non-smoking men, the presence of carotid atherosclerosis is associated with significant increases in the counts of many leukocytes subtypes. On the contrary, in non-smoking women, this correlation is absent. These results are compatible with recently recognized SD in the mechanism and pathophysiology of atherosclerosis. It is evident that monocyte counts are higher in men than in women [136, 263]. Interestingly, the monocyte count is influenced by sex hormones, at least in women, for whom sex hormones are higher in the luteal phase than in the follicular phase or during pregnancy [228, 264]. HRT—in menopause reduces the monocyte count [228], while the monocyte count is not influenced by OC use [68]. Globally, these findings suggest that female sex hormones decrease monocyte numbers probably because the hormones induce mitotic arrest and apoptosis [228]. It is also important to note that some subsets of circulating monocytes such as CD14⁺⁺16⁺⁺ are higher in men than in women. The percentage of monocytes expressing CD99⁺⁺, an index of adhesion and diapedesis, is lower in women than in men [265]. The adipokine leptin, a satiety signal that stimulates the immune system [266], is associated with monocyte

count, especially in men [267]. Finally, ageing influences the monocyte count, which is primarily increased in women [202].

8.2 Human monocyte count and smoking

Interestingly, smoking elevates the monocyte counts only in women [136]. Transcriptomic studies show that, in humans, smoking produces perturbations of gene expression in blood cells and alterations in blood parameters [136, 268-271], suggesting that many other parameters may be influenced by smoking in a sex-specific manner.

8.3 Human monocyte and macrophage functions

Numerous SD have been described in inflammation and immunity, and they involve different pathways [22-24]. One of the most important functions of monocytes is the production of cytokines and chemokines, and some SD regarding this point have been observed. However, some of these SD have been found to be controversial *in vitro* [25]. In particular, the basal release of TNF α in men is more than twice that in women, whereas LPS-induced TNF α release is typically greater in women than in men [[25] and quoted literature]. Moreover, TNF α release from monocytes obtained from women with CAD is lower in comparison with male patients [272]. Basal IL1 β , IL6 and IL8 production by monocytes and in response to LPS is similar or less in women compared to men [[25] and quoted literature]. Interestingly, the basal and LPS-induced IL1 β release is higher in the luteal phase compared to the follicular phase [25]. LPS-induced IL12 release is greater in men than in women, and it is not affected by the menstrual cycle [25].

Human monocytes also produce LTs, and female-derived monocytes have a higher capacity (1.8-fold) to produce pro-inflammatory LTs than male-derived monocytes [273]. *In vitro*, the exposure to dihydrotestosterone greatly reduces LT synthesis in female-derived monocytes, while oestradiol and

progesterone have practically no effect. Notably, pregnancy elevates 5-LO by-products partly because pregnancy elevates neutrophil and monocyte counts [274]. Progesterone reduces the production of 5-LO metabolites in human primary monocytes and is more active in female cells than in male cells. The progesterone effect is rapid and reversible [243]. Phospholipase D activity and diacylglycerol formation are 1.4- to 1.8-fold lower in male-derived monocytes than in female-derived monocytes, and this is linked to increased phosphorylation of ERKs [273]. Therefore, the arachidonic acid appears to be more active in women than in men.

Male- and female-derived monocytes respond differently to leptin, an adipokine involved in regulating food intake, body weight and immunity. In fact, recombinant leptin induces CD16 expression only in cells isolated from men [267]. *In vitro*, this leptin effect is inhibited by oestrogens [275]. Considering the pro-atherotic role of leptin in human monocytes [276], the SD just described could, at least in part, explain the SD in atherosclerosis.

It is hard to investigate SD in Toll-like receptors (TLRs), especially TLR4 responsiveness (LPS is an agonist of TLR4). TLR4 is present on monocytes, macrophages, DCs and other cells that are involved in atherosclerosis by mediating monocyte/macrophage infiltration and foam cell formation [277]. Therefore, understanding the influence of sex on this system is of special interest. Unfortunately, there is no clear evidence of a sex difference in TLR4 expression on monocytes [[25] and quoted literature], although reduced TLR4 responsiveness to LPS has been observed in monocytes/macrophages derived from women *in vitro* [[25] and quoted literature].

Monocytes obtained from women with CAD have a higher expression of PPAR γ protein compared with male patients [272]. Diurnal expression of PPAR α and brain and muscle aryl hydrocarbon receptor nuclear translocator like-1 (BMAL1) mRNA profiles in monocytes obtained from women in the fed state differ qualitatively and quantitatively from those of men. Diurnal PPAR α and BMAL1 profiles of fasted women resembled those of men in the fed and fasted states [278]. In other words,

women change their diurnal expression profiles of PPAR α and BMAL1 when switching from the fed to the fasted state, whereas men do not. This could explain why PPAR α agonists such as fenofibrate significantly reduce the total bile acid concentration only in men [279] and why the combination therapy of statin and fibrate is active in diabetic men but not in diabetic women [280]. SD have also been observed with statins alone. In particular, *in vitro* exposure to simvastatin (SMV) and atorvastatin (ATV) inhibits spontaneous and LPS-induced chemotaxis especially in female-derived monocytes [281]. However, an *in vivo* study in type 2 diabetic patients did not confirm the sex-specific effect of statins. In fact, these patients, independently of sex, have the same release of TNF α , IL1 β , IL6, and MCP1, which are decreased after treatment with SMV, fenofibrate, and SMV+fenofibrate [282].

In MDMs, the basal release of TNF α is influenced by OCs. Specifically, the basal release of TNF α is higher in women treated with OCs [68], while the release of TNF α induced by LPS is lower in OC users than in non-users [68]. The basal release of TNF α in MDMs is elevated in non-smoking men versus non-smoking women, but this sex difference is not present in MDMs obtained from smoking men and women [136]. The exposure to LPS increases the release of TNF α in non-smoking women and decreases it in non-smoking men, whereas no significant difference between sexes has been found in MDMs obtained from smoking people [136].

It is important to recall that 24-h LPS exposure modifies the expression and activity of ER α and ER β in MDMs in a sex-dependent manner. In particular, LPS up-regulates the ER α level especially in male cells. On the other hand, the ER β level is down-regulated only in female MDMs [283]. In addition, LPS increases ER α activation without any significant effect on ER β activation status [283]. The ability of TLR agonists, including LPS, to increase ER α phosphorylation has also been reported and is associated with an increase in the inflammatory response of mesangial cells [284]. Together, these data confirm the pivotal role played by ER α in the LPS-induced inflammatory response.

Macrophages can be subdivided according to their polarization into classically activated cells (M1) or non-classically activated cells (M2) [285]. M1 enhance and sustain inflammatory responses. Conversely, M2 secrete anti-inflammatory cytokines and promote the resolution of inflammation clearing of apoptotic cells, the dampening of immune responses, and tissue repair and healing. Premenopausal and postmenopausal women have similar M1/M2 ratios [286]. Both M1 and M2 are present at different stages of human atherosclerotic plaque development [245]. M1 express ER α and ER β , but only ER α is reduced by M1 agonists such as LPS [LPS]/interferon- γ (IFN γ) [286].

8.4 Human monocytes and macrophages and cell fate

The cellular fate of monocytes is predominantly regulated by chemokines and their receptors [287]. Granulocyte-macrophage colony-stimulating factor, macrophage colony-stimulating factors, and colony-stimulating factor-1 are relevant for the differentiation of monocytes into macrophages [288]. The inhibition of apoptosis contributes to the accumulation of macrophages and the persistence of an inflammatory milieu. Autophagy in macrophages processes bulky materials, participates in inflammatory responses [289-291], contributes to cholesterol homeostasis (lipophagy) [292-294] and facilitates efferocytosis (a process in which apoptotic cells are phagocytised and removed) [295, 296]. Some studies clearly show that the atheroprotective effect of macrophage autophagy is due to increased cholesterol efflux, reduced inflammation, and improved efferocytosis [291, 295].

8.4.1 Apoptosis

Monocytes are short-lived and undergo spontaneous apoptosis, and spontaneous apoptosis facilitates the resolution of the immune response [297, 298]. The apoptosis increases with each stage of atherosclerosis, but the largest increase is observed in the vulnerable plaque [299]. In fact, the necrotic core of plaques is derived from a combination of impaired phagocytosis of apoptotic cells and

accelerated macrophage cell death [300]. Female sex hormones delay apoptosis only in peripheral blood mononuclear cells obtained from women with normal menstrual cycles but do not inhibit TNF α production [301]. The paucity of data indicates that future detailed studies on the influences of sex on apoptosis in macrophages and monocytes will be necessary.

8.4.2 Autophagy

The deregulation of autophagy in monocytes and macrophages has only recently been the focus of studies [[302] and quoted literature]. Notably, lipid droplets can also be eliminated by autophagic-dependent phenomena [293]. Peripheral blood monocytes obtained from patients with acute coronary syndrome have lower levels of autophagic markers such as beclin-1 and ATG7 [303]. Unfortunately, the influence of sex on human monocyte and macrophage autophagy is still unknown. Therefore, there is an urgent need for studies of this specific point because SD are emerging in other human cells, but the differences are cell- and parameter-specific [304].

8.5 Human macrophages and foam cells

Macrophages are the first invaders of atherosclerotic lesions and the main component of atherosclerotic plaques [305]. The adherence of monocytes to ECs and their transmigration into the sub-endothelial space to differentiate into macrophages is an early step in atherogenesis, after which they accumulate lipids to form foam cells. The recruitment of circulating monocytes to the endothelium is facilitated by cell adhesion molecules (CAMs), including intercellular adhesion molecule-1 (ICAM1) and vascular cell adhesion molecule-1 (VCAM1). The levels of soluble ICAM1 in men are better predictors of the risk of myocardial infarction than those in women [306]. Exposure to sex hormones alters cholesterol metabolism in human MDMs obtained from young men and premenopausal women [307, 308]. Exposure of human MDMs to oestrogen or progesterone reduces the accumulation of cholesteryl ester

only in female cells [309]. Therefore, oestrogen and progesterone reduce lipid accumulation in a sex-specific manner. *In vitro*, dihydrotestosterone increases monocyte-endothelial adhesion, up-regulating VCAM1 in male cells but not in female cells [309, 310].

Notably, androgens affect the expression of genes involved in lipoprotein metabolism (lysosomal acid lipase, acyl CoA:cholesterol acyl transferase and cholesteryl ester hydrolysis) in male MDMs but not in female MDMs [308]. Cholesterol transport in macrophages plays a pivotal role in atherogenesis [311]. Scavenger receptors (SRs) of LDLs are the principal contributors to cholesterol uptake in macrophages [311], and numerous types of SRs are present [311]. Cholesterol efflux occurs through ABCA1, ABCG1, and apolipoprotein E [312-314]. SD in LDL receptors and lipoprotein uptake are primarily described in animal models [315-319], whereas only few studies have been conducted using human cells. However, oestrogens regulate cholesterol transport in MDMs obtained from postmenopausal women and age-matched men. Oestrogens appear to regulate cholesterol transport by inhibiting scavenger receptor A through heat shock protein 27 [320].

In particular, oestrogens lower cholesteryl ester levels in both female and male cells, but they do not affect cholesterol efflux [321]. The reduction of cholesterol esters in MDMs should mitigate foam cell formation. Finally, oestrogens, but not androgens, have a small effect on the expression of several genes involved in cholesterol transport, but they do not affect protein expression [321]. Moreover, oestrogen-and progesterone-treated macrophages from premenopausal female donors bind a significantly greater proportion of labelled acetylated LDL [307]. In male MDMs specifically, androgens elevate cholesteryl ester content [257], but this is not univocal data [139].

9. Human DCs

DCs are constituted by different subtypes (conventional/classical DCs (cDCs), plasmacytoid DCs (pDCs), Langerhans cells, and monocyte-derived CD11b⁺ inflammatory/migratory DCs). pDCs

express ER α and ER β as well as CD8 T and monocyte-derived DCs (Table 1) [322], whereas intracellular PRs (Table 1) are not expressed on resting DCs [27]. AR receptors are expressed in these cells [26], but it is not clear whether there are SD.

9.1 Human DCs and smoking

A paucity of data is present on the effect of smoking on DCs, and even less data are available on the interaction between smoking and sex on DCs. In particular, the number of pDCs in the human airway is not influenced by smoking, while the number of mDCs in the blood decreases in healthy male and female smokers; however, the effect of sex was not analysed [323]. Cigarette smoke can also impair the function of human monocyte-derived DCs, enhancing IL10 secretion and prostaglandin E2 release. Furthermore, cigarette smoke extract decreases DC-mediated priming of T cells, specifically inhibiting key Th1 cytokine production and favouring the development of Th2 responses. This results in a state of arrested DC maturation and reduced DC number [324]. The above data indicate that DCs are affected by smoking, but it is still unknown whether the effect of smoking is sex specific.

9.2 Function of human DCs

Very little data are available on sex effect in DCs. It has been shown that DCs from premenopausal women respond better to TLRs than those of men, producing a large amount of IFNs in response to TLR7 (including HIV-1) and TLR9 ligands [325-327]. The difference is not present when men and postmenopausal women are compared, but HTR restores the SD versus TLR7 responses [327], indicating that oestrogens can play a role. Blocking ERs *in vitro* during pDC differentiation reduces the frequency of cytokine-producing cells in response to TLR7 stimulation, suggesting that oestrogens may act as a cell-extrinsic factor to positively regulate the TLR7 responses of pDCs in women in a cell-intrinsic manner [328]. Furthermore, the production of IFN α is primarily regulated at the transcriptional

level by the IFN regulatory factors (IRF) family, and female cells express more IRF5 (1.6 times more) than male cells [329]. Importantly, the SD in IRF5 expression are not present in CD3⁺ T cells and in monocytes/cDCs [329], indicating a certain cell specificity. Notably, IRF5 elevates the transcription of TNF α , IL8, MIP1 α , and MIP1 β without SD, suggesting that sex influence is cytokine specific. Finally, IRF5 protein expression does not change with the use of OCs. In contrast, Seillet and colleagues [327] report an elevated frequency of TNF α -producing pDCs in women compared with men after stimulation with a TLR7 ligand. Finally, it is important to note that IRF5 appears to be regulated, at least in part, by ER α [329]. Cytokine specificity, *in vitro*, was confirmed by data from Segerer and collaborators [330]. They observed that progesterone and oestradiol do not affect the release of IL8, MCP1, and CCL5 from monocyte-derived DCs from healthy men and women. Finally, 17 β -oestradiol induces CD40 expression through the activation of p38 and JNK, which increases minichromosome maintenance protein 6 expression, which in turn induces CD40 expression [331].

During the so called “mini-puberty” (in first six months of life, infants have circulating levels of sex hormones near the levels seen during puberty), pDCs obtained from female infants, just as those obtained from women, generate more IFN α than those obtained from male infants in response to stimulation with a stimulator of TLRs 7/8. Interestingly, androgen signalling dose-dependently down-regulates IFN α production. In the same cohorts, TNF α production in pDCs does not vary between the sexes when the DCs are exposed to a stimulator of TLRs 7/8 in a dose-dependent fashion [332].

9.3 Human DCs and cell fate

Programmed cell death is an important element of the lifespan of DCs. However, SD in apoptosis and autophagy in XX and XY cells have not been thoroughly investigated because most cell-based studies have been performed without consideration of the male or female origin of the cells.

9.3.1 Apoptosis

It has been shown that dysregulation of apoptosis in DCs is involved in sepsis-induced immunosuppression [333]. As shown in [334], in mDCs obtained from healthy donors of both sexes, apoptosis is enhanced by progesterone and oestradiol *in vitro*; however, no analysis of sex was performed in this study.

9.3.2 Autophagy

To the best of our knowledge, no SD have been reported in the autophagic process in human DCs.

10. Human lymphocytes

Approximately 30% of white blood cells are lymphocytes. They comprise three types of cells: T lymphocytes (85-90% of circulating lymphocytes) release cytokines including IL2, IL4, IL10, IFN γ and TNF α and underlie cell-mediated adaptive immunity; B lymphocytes (5–15% of circulating lymphocytes) produce IgG and IgM antibodies and underlie antibody-driven adaptive immune responses; and natural killer cells (NKs), which are effectors of cell-mediated innate immunity. They are very well represented in atherosclerotic plaques, where T lymphocytes comprise 20% of the immune cells [14 and quoted literature]. Most of them are CD8⁺ cytotoxic T cells [335]. Formation of the atherosclerotic lesion and destabilization of the plaque begin with the activation of T lymphocytes and their Th2 versus the Th1 subset [245].

Human lymphocytes express both ER α and ER β [336] (Table 1). It has also been shown that, in CD4⁺ and CD8⁺ T lymphocytes, B lymphocytes and NKs, the ER α 46 isoform is the most represented [337]. Intracellular PRs are not expressed on resting lymphocytes, NKs, or DCs [27], but membrane-bound PRs are present on resting lymphocytes (Table 1) [27]. Interestingly, activated lymphocytes can up-regulate PRs [27]. Finally, the presence of ARs is still a controversial issue (Table 1) [27].

In B lymphocytes, 358 genes exhibit sexual dimorphism: compared to male-derived cells, 226 and 13 genes are found to be up- and down-regulated, respectively, in female-derived cells [338]. Furthermore, Hewagama and collaborators identified 1953 genes that exhibit sexual dimorphism in T lymphocytes [339]. Despite the genetic sex differences and the involvement of T and B lymphocytes in atherosclerotic plaques [340], the influence of sex is scarcely considered in studies of lymphocytes. Therefore, only few data are available.

10.1 Human lymphocyte counts

Men and women have similar numbers of lymphocytes [[32] and quoted literature]. This is not an univocal fact because some studies have shown higher levels of lymphocytes in male subjects [341]. However, when lymphocyte subtypes are considered, some authors report lower T lymphocyte counts in males than in females [205]. B lymphocytes are reported to be higher in females [341, 342], and studies show that women have a significantly higher percentage of CD3⁺ and CD4⁺ and a lower percentage of NKs than men. The lymphocyte count in women is influenced by menopausal status. The number of T and B lymphocytes and helper T cells are lower in postmenopausal women versus premenopausal women [32], but the total lymphocyte count and the subtype count do not vary throughout the menstrual cycle [228]. OCs do not affect the absolute numbers or subtype counts [228]. In contrast, HRT affects lymphocyte subtypes. The total lymphocyte counts and the percentages of T cells and Th lymphocytes are lower in HRT users than in non-users [228]. Ageing influences lymphocyte counts with T helper cells and cytotoxic T cells decreased primarily in men [202]. More older men exhibit an inverted CD4/CD8 T cell ratio compared to women [343]. In elderly women, there is an increase in the proportion of T cells that are positive for IFN γ , IL2, IL4, and IL10 and the proportion of cells presenting IL4 or IFN γ ; whereas, in old men, a higher percentage of T cells produce

IL2, IL4 and IL13 [344]. The changes induced by ageing are more evident in males, although statistical relevance is obtained only for CD8⁺ T cells and for effector memory cells [345].

In particular, the activation of the stress system changes the distribution of T and NKs (decreasing their ratio), independent of sex or menstrual cycle phase [346]. Globally, in women, oestrogens appear to play a role in the regulation of lymphocyte numbers.

10.2 Human lymphocyte count and the effect of smoking

Data regarding the influence of smoking on lymphocyte subsets are few and conflicting. However, one study showed that smoking status increases the lymphocyte counts only in women [136].

10.3 Human lymphocyte function

Women have more pronounced humoral and cellular immune responses to antigens than males [[32] and quoted literature]. SD in bacterial, viral and parasitic infections have been extensively reviewed by Ngo ST et al. [[32] and quoted literature]. In particular, women produce higher levels of CD4⁺ T cells in response to immunization, with higher levels of circulating antibodies after vaccination for influenza, hepatitis B, rubella and tetanus than men [[32] and quoted literature]. Women also have higher levels of IgM than men [[32] and quoted literature].

The culture of peripheral blood mononuclear cells results in significant increases in the percentages of B cells and total T cells among females and of NKs among males [341], indicating that culture conditions have different consequences between the sexes. As previously suggested by Franconi and collaborators [347], this result indicates that sex studies require peculiar attention to experimental conditions because the very same experimental condition may affect experimental outcomes in a sex-specific manner.

Sex appears to influence lymphocyte function in an age-dependent manner. T cells isolated from older men have a lower proliferative and cytokine secretion capacity than cells from older women. The ability of monocytes to secrete a chemoattractant for activated T cells, IFN- γ -inducible protein 10 (IP-10), is reduced in a similar fashion [26]. The production of IFN γ and IL17 in stimulated T cells from healthy older men is lower than in those from healthy young men. However, in T cells obtained from old and young women, the production of IFN γ and IL17 is not different [348]. T regulatory cells and Th2 CD4⁺ helper T cells produce the anti-inflammatory cytokine IL10 [349]. The plasma levels of IL10 decline with age, with a greater decline in men than in women [350]. Interestingly, elevated levels of IL10 are associated with stroke outcomes only in women [351].

10.4 Human lymphocytes and cell fate

Apoptosis participates in the regulation of the immune response. Nevertheless, death pathways in immune XX and XY cells have not been thoroughly investigated because most cell-based studies have been performed without consideration of the male or female origin of the cells. Autophagy has been reported in T and B cells [352]. In T cells, autophagy provides substrates and modulates NF κ B activation [352]. Less is known about the role of autophagy in B cells [352].

10.4.1 Apoptosis

In vitro, sex hormones delay apoptosis in B and T cells obtained from women with normal menstrual cycles [301, 353]. The activation of the stress system changes the distribution of T cells and NKs (decreasing their ratio); however, more interestingly, this is accomplished by an increase in apoptotic T helper cells independent of sex or menstrual cycle phase [346].

10.4.2 Autophagy

This cannibalistic process has been implicated in cellular survival and programmed cell death. Currently, to the best of our knowledge, it is not known whether lymphocyte autophagy is influenced by the sex of cells.

11. Human NKs

Human NKs consist of two subsets: CD56dim and CD56bright. CD56dim constitutes 90% of the total NK cell population in the peripheral blood and has high cytotoxic activity. The other 10% of NKs consist of CD56 [[354] and quoted literature]. These cells have the capacity to kill cells and to produce cytokines, and they play a central role in the innate immune response against tumours, parasites and infected cells. They also play an immunoregulatory role in the pathogenesis and progression of atherosclerosis, although it is not clear whether they are pro-atherotic or antiatherothic [355].

Both ER α and ER β (Table 1) are expressed in human NKs [322, 336], and it has also been shown that the ER α 46 isoform is the most represented [337]. Intracellular PRs (Table 1) are not expressed in NKs [27].

11.1 Human NK counts

NKs constitute approximately 5% of the leukocytes. The data on SD in NKs are scarce and discrepant. In particular, it has been reported that: a) the NK counts in males and females (postmenopausal or fertile) and OC users [[228] and quoted literature] do not differ, whereas they are higher in women with premature menopause [[32] and quoted literature]; b) the NK count varies during the menstrual cycle, increasing in the late secretory phase [[228] and quoted literature]; c) the NK count has been shown to be higher in males than in females in some studies [341], while other studies have provided evidence [356, 357] that the NK count is lower in men than in women; d) there are no SD in the subset

distributions of circulating NKs [358]. Furthermore, in older men, the proportion of positive and double-negative CD4⁺ cells is elevated and reduced, respectively [356]. In older individuals, a recent paper shows that there are not SD in NKs and T cells measured as a percentage of lymphocytes. However, more B cells are present in women than in men, and the ratio of CD56^{bright} to CD56^{dim} NKs is larger in older women than in older men [359], suggesting a sex difference in maturation of these cells. Indeed, the number of NKs increases in men and women > 60 years of age, but the increase is higher in women than in men [360]. Furthermore, during pregnancy, peripheral blood NKs are suppressed both in terms of number and activity [[228] and quoted literature], whereas mental stress increases the number of NKs in women but decreases them in men [346]. Globally, these results suggest that some of the discrepancies reported could depend on the age and hormonal status of women. The ARs and ERs are present on these cells, but sex stratification is currently missing (Table 1).

11.2 Human blood NKs and smoking

Smoking impairs the immune response and the peripheral blood leukocyte counts including the NK count, which is lower in smokers [361-363]. Moreover, an impairment in the production of IFN γ and TNF α is reported in NKs isolated from smokers [364]. In the previous studies, stratification for sex is missing, and therefore we do not know the sex effect.

11.3 Function of human blood NKs

Data on SD of NK activity is limited. In particular, NKs in men exhibit more cytotoxic activity than those in women with regular menstrual cycles or women using OCs, who have the lowest levels of activity [[32] and quoted literature]. The menstrual cycle has no significant effect on the activity levels of NKs [365]. However, others have shown that NKs are more cytotoxic in the follicular phase than in

the luteal phase and in postmenopausal women. Males exhibit cytotoxic activity similar to that observed in the follicular phase [[32, 228] and quoted literature]. In older individuals, mature NKs obtained from women have a more cytotoxic response and produce more MIP-1 β in response to a variety of stimuli [359]. These data show that sex influences NK activity in elderly individuals. The α -galactosylceramide-induced intracellular production of IFN γ , IL4, IL17 and TNF by CD4⁺ and DN⁺NKT cells is larger in women than in men [357]. Data on the *in vitro* effects of progesterone, oestrogens and androgens on human NKs are not univocal [366-368]. Finally, there are not significant SD in IL4 response, but cells obtained from men produce more IFN γ and MIP1 α than female cells [358]. Therefore, NKs may contribute to the sexual dimorphism.

11.4 Human NKs and cell fate

SD in apoptosis and autophagy in XX and XY cells have not been thoroughly investigated because most cell-based studies have been performed without consideration of the male or female origin of the cells.

11.4.1 Apoptosis

Notably, mental stress increases the apoptotic T helper cell percentage irrespective of sex or menstrual cycle phase [346]. It has been reported that NK death occurs via TNF family receptors [369], but no information is available on sex effects.

11.4.2 Autophagy

To the best of our knowledge, there is no data on the influences of sex on autophagy of human NKs.

12. Human mast cells

These cells are resident in a multitude of tissues and play a role in the defence against microorganisms [[32] and quoted literature]. Mast cells have pleiotropic roles in atherosclerosis as well as in associated complications. Indeed, the number of mast cells in humans with cardiometabolic diseases is increased compared with healthy adults [370]. Additionally, the molecules released from mast cells might contribute to the pathogenesis of CVD [370]. Mast cells contribute to plaque progression and destabilization [7]. Activated mast cells can have deleterious effects on the vessel wall, degrading the extracellular matrix, enhancing apoptosis, and recruiting inflammatory cells. The expression of ER α , ER β , both PRs and ARs (Table 1) has been demonstrated in mast cells, and oestrogens can enhance mast cell degranulation, whereas the influence of androgens remains largely unclear [371-373]. Both oestrogen and progesterone can activate mast cells [341].

12.1 Function of human mast cells

Mast cells are implicated in inflammation during atherogenesis and plaque destabilization. Activated mast cells increase vascular leakage and the influx of leukocytes into the plaque and induce intra-plaque haemorrhage causing plaque destabilization [374]. Mast cell number is a good marker for recent lesions, and it increases in male patients, particularly in older male patients [375]. Oestrogen or progesterone induces mast cell degranulation, whereas androgens are not effective [371]. There is limited knowledge of the involvement of mast cells in SD in atherosclerosis.

13. CONCLUSIONS

Globally, an analysis of the literature shows that sex aspects have been neglected in studies of cells involved in atherosclerosis with few exceptions because the majority of *in vitro* studies do not indicate the sex of the cells used and because most of the clinical studies are comprised of a majority of men. Additionally, many studies do not consider the methodological issues of sex and gender research such

as physiological aspects (age, menstruation, pregnancy, lifestyles, etc.) and the pre-analytical conditions [347]. This could explain the great deal of variability of the results. This variability increases the difficulty in the comparison of the results of sex studies among different laboratories. The comparison is, in fact, a very complicated and hard task because investigators use cells from individuals of different ages and with different lifestyles. Furthermore, *in vitro* studies are often performed using various different methodologies such as different hormonal concentrations and different times of incubation. This leads to an increase in variability, and the obvious consequence is the presence of controversial results.

The previous observations demonstrate the need for a clear definition of experimental conditions as has been previously suggested by Franconi and collaborators [347]. In our opinion, a mandatory questionnaire should be presented to cell donors, and the very same questionnaire should be used by all researchers in order to reduce variability among laboratories. A sample questionnaire has already been proposed by Franconi and collaborators [347].

Controversial data are even present when cell counts are considered. In view of the fact that an increase in some cell numbers is a risk factor for CVD, in our opinion, it is time to establish reference values for men and women considering, at least, age, phase of life of women, and some lifestyle choices such as smoking in order to appropriately and adequately evaluate CV risk for men and women. Considering the multiplicity of cell players and their complex interactions in atherosclerosis, the analysis of the literature indicated that there is an urgent need to extend the study of sex influences not only for single cells but also for interactions among cells such as PLT-neutrophil satellitism [376]. Therefore, it appears relevant to consider gender. In order to improve the diagnosis and treatment of CAD, the research community urgently needs to understand the SD of vessel wall cells and immune cells in the specific setting of atherosclerosis. The available data suggest that cells retain memory of sex and of the

life events of the donors. For example, treatment with OCs modifies macrophage function and expression and activity of ERs [68], indicating that cells not only have a sex but also have a gender.

REFERENCES

- [1] Libby P, Tabas I, Fredman G, Fisher EA. Inflammation and its resolution as determinants of acute coronary syndromes. *Circ Res*. 2014;114:1867-79.
- [2] Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. *Circulation*. 2002;105:1135-43.
- [3] Miller YI, Choi SH, Wiesner P, Fang L, Harkewicz R, Hartvigsen K, et al. Oxidation-specific epitopes are danger-associated molecular patterns recognized by pattern recognition receptors of innate immunity. *Circ Res*. 2011;108:235-48.
- [4] Stary HC, Chandler AB, Dinsmore RE, Fuster V, Glagov S, Insull W, Jr., et al. A definition of advanced types of atherosclerotic lesions and a histological classification of atherosclerosis. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Arterioscler Thromb Vasc Biol*. 1995;15:1512-31.
- [5] Chistiakov DA, Orekhov AN, Bobryshev YV. Endothelial barrier and its abnormalities in cardiovascular disease. *Front Physiol*. 2015;6:365-75.
- [6] Ahmadsei M, Lievens D, Weber C, von Hundelshausen P, Gerdes N. Immune-mediated and lipid-mediated platelet function in atherosclerosis. *Curr Opin Lipidol*. 2015;26:438-48.
- [7] Conti P, Shaik-Dasthagirisae Y. Atherosclerosis: a chronic inflammatory disease mediated by mast cells. *Cent Eur J Immunol*. 2015;40:380-6.
- [8] Wizemann T, Pardue M. Exploring the biological contributions to human health: does sex matter? . Washington: National Academy Press; 2001.
- [9] Legato M. Principles of gender-specific medicine. San Diego: Academic Press; 2010.
- [10] Taylor KE, Vallejo-Giraldo C, Schaible NS, Zakeri R, Miller VM. Reporting of sex as a variable in cardiovascular studies using cultured cells. *Biol Sex Differ*. 2011;2:11.
- [11] Addis R, Campesi I, Fois M, Capobianco G, Dessole S, Fenu G, et al. Human umbilical endothelial cells (HUVECs) have a sex: characterisation of the phenotype of male and female cells. *Biol Sex Differ*. 2014;5:18.
- [12] Banos G, Guarner V, Perez-Torres I. Sex steroid hormones, cardiovascular diseases and the metabolic syndrome. *Cardiovasc Hematol Agents Med Chem*. 2011;9:137-46.
- [13] Viridis A, Taddei S. Endothelial aging and gender. *Maturitas*. 2012;71:326-30.
- [14] Vitale C, Mendelsohn ME, Rosano GM. Gender differences in the cardiovascular effect of sex hormones. *Nat Rev Cardiol*. 2009;6:532-42.
- [15] Mumford SL, Dasharathy S, Pollack AZ, Schisterman EF. Variations in lipid levels according to menstrual cycle phase: clinical implications. *Clin Lipidol*. 2011;6:225-34.
- [16] Shchelkunova TA, Morozov IA, Rubtsov PM, Samokhodskaya LM, Andrianova IV, Sobenin IA, et al. Changes in levels of gene expression in human aortal intima during atherogenesis. *Biochemistry (Mosc)*. 2013;78:463-70.
- [17] Campesi I, Capobianco G, Dessole S, Occhioni S, Montella A, Franconi F. Estrogenic compounds have divergent effects on human endothelial progenitor cell migration according to sex of the donor. *J Vasc Res*. 2015;52:273-8.
- [18] Fadini GP, de Kreutzenberg S, Albiero M, Coracina A, Pagnin E, Baesso I, et al. Gender differences in endothelial progenitor cells and cardiovascular risk profile: the role of female estrogens. *Arterioscler Thromb Vasc Biol*. 2008;28:997-1004.
- [19] Getz GS, Reardon CA. Animal models of atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2012;32:1104-15.

- [20] Pellegrini M, Bulzomi P, Lecis M, Leone S, Campesi I, Franconi F, et al. Endocrine disruptors differently influence estrogen receptor beta and androgen receptor in male and female rat VSMC. *J Cell Physiol.* 2014;229:1061-8.
- [21] Ding Q, Gros R, Limbird LE, Chorazyczewski J, Feldman RD. Estradiol-mediated ERK phosphorylation and apoptosis in vascular smooth muscle cells requires GPR 30. *Am J Physiol Cell Physiol.* 2009;297:C1178-87.
- [22] Oertelt-Prigione S. The influence of sex and gender on the immune response. *Autoimmun Rev.* 2012;11:A479-85.
- [23] Oertelt-Prigione S. Immunology and the menstrual cycle. *Autoimmun Rev.* 2012;11:A486-92.
- [24] Trigunaite A, Dimo J, Jorgensen TN. Suppressive effects of androgens on the immune system. *Cell Immunol.* 2015;294:87-94.
- [25] Jiang W, Gilkeson G. Sex differences in monocytes and TLR4 associated immune responses; implications for systemic lupus erythematosus (SLE). *J Immunother Appl.* 2014;1:1-18.
- [26] Gubbels Bupp MR. Sex, the aging immune system, and chronic disease. *Cell Immunol.* 2015;294:102-10.
- [27] Faas M, de Vos P, Melgert B. Sex hormones and immunoregulation. In: Elenkov IJ, editor.: *BrainImmune*; 2011.
- [28] Korshunov VA, Berk BC. Smooth muscle apoptosis and vascular remodeling. *Curr Opin Hematol.* 2008;15:250-4.
- [29] Walsh K, Smith RC, Kim HS. Vascular cell apoptosis in remodeling, restenosis, and plaque rupture. *Circ Res.* 2000;87:184-8.
- [30] Mallat Z, Ohan J, Leseche G, Tedgui A. Colocalization of CPP-32 with apoptotic cells in human atherosclerotic plaques. *Circulation.* 1997;96:424-8.
- [31] Stoneman VE, Bennett MR. Role of apoptosis in atherosclerosis and its therapeutic implications. *Clin Sci (Lond).* 2004;107:343-54.
- [32] Ngo ST, Steyn FJ, McCombe PA. Gender differences in autoimmune disease. *Front Neuroendocrinol.* 2014;35:347-69.
- [33] Lavandero S, Troncoso R, Rothermel BA, Martinet W, Sadoshima J, Hill JA. Cardiovascular autophagy: concepts, controversies, and perspectives. *Autophagy.* 2013;9:1455-66.
- [34] Malorni W, Campesi I, Straface E, Vella S, Franconi F. Redox features of the cell: a gender perspective. *Antioxid Redox Signal.* 2007;9:1779-801.
- [35] Giergiel M, Lopucki M, Stachowicz N, Kankofer M. The influence of age and gender on antioxidant enzyme activities in humans and laboratory animals. *Aging Clin Exp Res.* 2012;24:561-9.
- [36] Appelman Y, van Rijn BB, Ten Haaf ME, Boersma E, Peters SA. Sex differences in cardiovascular risk factors and disease prevention. *Atherosclerosis.* 2015;241:211-8.
- [37] Yang Z, Ming XF. Recent advances in understanding endothelial dysfunction in atherosclerosis. *Clin Med Res.* 2006;4:53-65.
- [38] Simard M, Drolet R, Blomquist CH, Tremblay Y. Human type 2 17beta-hydroxysteroid dehydrogenase in umbilical vein and artery endothelial cells: differential inactivation of sex steroids according to the vessel type. *Endocrine.* 2011;40:203-11.
- [39] Annibalini G, Agostini D, Calcabrini C, Martinelli C, Colombo E, Guescini M, et al. Effects of sex hormones on inflammatory response in male and female vascular endothelial cells. *J Endocrinol Invest.* 2014;37:861-9.
- [40] Toth B, Saadat G, Geller A, Scholz C, Schulze S, Friese K, et al. Human umbilical vascular endothelial cells express estrogen receptor beta (ERbeta) and progesterone receptor A (PR-A), but not ERalpha and PR-B. *Histochem Cell Biol.* 2008;130:399-405.
- [41] Kim-Schulze S, McGowan KA, Hubchak SC, Cid MC, Martin MB, Kleinman HK, et al. Expression of an estrogen receptor by human coronary artery and umbilical vein endothelial cells. *Circulation.* 1996;94:1402-7.
- [42] Tu J, Jufri NF. Estrogen signaling through estrogen receptor beta and G-protein-coupled estrogen receptor 1 in human cerebral vascular endothelial cells: implications for cerebral aneurysms. *Biomed Res Int.* 2013;2013:524324.

- [43] Villablanca AC, Jayachandran M, Banka C. Atherosclerosis and sex hormones: current concepts. *Clin Sci (Lond)*. 2010;119:493-513.
- [44] Death AK, McGrath KC, Sader MA, Nakhla S, Jessup W, Handelsman DJ, et al. Dihydrotestosterone promotes vascular cell adhesion molecule-1 expression in male human endothelial cells via a nuclear factor-kappaB-dependent pathway. *Endocrinology*. 2004;145:1889-97.
- [45] McGrath KC, Hill MD, McRobb LS, Heather AK. The androgen receptor drives the sex-specific expression of vascular cell adhesion molecule-1 in endothelial cells but not lipid metabolism genes in monocyte-derived macrophages. *Horm Mol Biol Clin Investig*. 2010;2:203-9.
- [46] Rajendran P, Rengarajan T, Thangavel J, Nishigaki Y, Sakthisekaran D, Sethi G, et al. The vascular endothelium and human diseases. *Int J Biol Sci*. 2013;9:1057-69.
- [47] Mudau M, Genis A, Lochner A, Strijdom H. Endothelial dysfunction: the early predictor of atherosclerosis. *Cardiovasc J Afr*. 2012;23:222-31.
- [48] Sitia S, Tomasoni L, Atzeni F, Ambrosio G, Cordiano C, Catapano A, et al. From endothelial dysfunction to atherosclerosis. *Autoimmun Rev*. 2010;9:830-4.
- [49] Tabas I, Garcia-Cardena G, Owens GK. Recent insights into the cellular biology of atherosclerosis. *J Cell Biol*. 2015;209:13-22.
- [50] Hashimoto M, Akishita M, Eto M, Ishikawa M, Kozaki K, Toba K, et al. Modulation of endothelium-dependent flow-mediated dilatation of the brachial artery by sex and menstrual cycle. *Circulation*. 1995;92:3431-5.
- [51] Adams MR, Robinson J, Sorensen K, Deanfield J, Celermajer DS. Normal ranges for brachial artery flow-mediated dilation: a non-invasive ultrasound test of arterial endothelial function. *J Vasc Invest*. 1996;2:146-50.
- [52] Skaug EA, Aspenes ST, Oldervoll L, Morkedal B, Vatten L, Wisloff U, et al. Age and gender differences of endothelial function in 4739 healthy adults: the HUNT3 Fitness Study. *Eur J Prev Cardiol*. 2013;20:531-40.
- [53] Mizia-Stec K, Gasior Z, Mizia M, Haberka M, Holecki M, Zwolinska W, et al. Flow-mediated dilation and gender in patients with coronary artery disease: arterial size influences gender differences in flow-mediated dilation. *Echocardiography*. 2007;24:1051-7.
- [54] Steinberg HO, Paradisi G, Cronin J, Crowde K, Hempfling A, Hook G, et al. Type II diabetes abrogates sex differences in endothelial function in premenopausal women. *Circulation*. 2000;101:2040-6.
- [55] Williams MR, Westerman RA, Kingwell BA, Paige J, Blombery PA, Sudhir K, et al. Variations in endothelial function and arterial compliance during the menstrual cycle. *J Clin Endocrinol Metab*. 2001;86:5389-95.
- [56] English JL, Jacobs LO, Green G, Andrews TC. Effect of the menstrual cycle on endothelium-dependent vasodilation of the brachial artery in normal young women. *Am J Cardiol*. 1998;82:256-8.
- [57] Cicinelli E, Ignarro LJ, Lograno M, Galantino P, Balzano G, Schonauer LM. Circulating levels of nitric oxide in fertile women in relation to the menstrual cycle. *Fertil Steril*. 1996;66:1036-8.
- [58] Jilma B, Kastner J, Mensik C, Vondrovec B, Hildebrandt J, Krejcy K, et al. Sex differences in concentrations of exhaled nitric oxide and plasma nitrate. *Life Sci*. 1996;58:469-76.
- [59] Celermajer DS, Sorensen KE, Spiegelhalter DJ, Georgakopoulos D, Robinson J, Deanfield JE. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. *J Am Coll Cardiol*. 1994;24:471-6.
- [60] Halligan SC, Murtagh B, Lennon RJ, Pumper GM, Mathew V, Higano ST, et al. Effect of long-term hormone replacement therapy on coronary endothelial function in postmenopausal women. *Mayo Clin Proc*. 2004;79:1514-20.
- [61] Best PJ, Berger PB, Miller VM, Lerman A. The effect of estrogen replacement therapy on plasma nitric oxide and endothelin-1 levels in postmenopausal women. *Ann Intern Med*. 1998;128:285-8.
- [62] Forte P, Kneale BJ, Milne E, Chowienczyk PJ, Johnston A, Benjamin N, et al. Evidence for a difference in nitric oxide biosynthesis between healthy women and men. *Hypertension*. 1998;32:730-4.
- [63] Taylor DR, Mandhane P, Greene JM, Hancox RJ, Filsell S, McLachlan CR, et al. Factors affecting exhaled nitric oxide measurements: the effect of sex. *Respir Res*. 2007;8:82-90.

- [64] Torres-Estay V, Carreno DV, San Francisco IF, Sotomayor P, Godoy AS, Smith GJ. Androgen receptor in human endothelial cells. *J Endocrinol*. 2015;224:R131-7.
- [65] Caulin-Glaser T, Garcia-Cardena G, Sarrel P, Sessa WC, Bender JR. 17 beta-estradiol regulation of human endothelial cell basal nitric oxide release, independent of cytosolic Ca²⁺ mobilization. *Circ Res*. 1997;81:885-92.
- [66] Hishikawa K, Nakaki T, Marumo T, Suzuki H, Kato R, Saruta T. Up-regulation of nitric oxide synthase by estradiol in human aortic endothelial cells. *FEBS Lett*. 1995;360:291-3.
- [67] Brancaleone V, Vellecco V, Matassa DS, d'Emmanuele di Villa Bianca R, Sorrentino R, Ianaro A, et al. Crucial role of androgen receptor in vascular H₂S biosynthesis induced by testosterone. *Br J Pharmacol*. 2015;172:1505-15.
- [68] Campesi I, Sanna M, Zinellu A, Carru C, Rubattu L, Bulzomi P, et al. Oral contraceptives modify DNA methylation and monocyte-derived macrophage function. *Biol Sex Differ*. 2012;3:4.
- [69] Cai J, Hong Y, Weng C, Tan C, Imperato-McGinley J, Zhu YS. Androgen stimulates endothelial cell proliferation via an androgen receptor/VEGF/cyclin A-mediated mechanism. *Am J Physiol Heart Circ Physiol*. 2011;300:H1210-21.
- [70] Finch J, Conklin DJ. Air pollution-induced vascular dysfunction: Potential role of Endothelin-1 (ET-1) system. *Cardiovasc Toxicol*. 2015.
- [71] Polderman KH, Stehouwer CD, van Kamp GJ, Dekker GA, Verheugt FW, Gooren LJ. Influence of sex hormones on plasma endothelin levels. *Ann Intern Med*. 1993;118:429-32.
- [72] Pearson LJ, Yandle TG, Nicholls MG, Evans JJ. Regulation of endothelin-1 release from human endothelial cells by sex steroids and angiotensin-II. *Peptides*. 2008;29:1057-61.
- [73] Batres RO, Dupont J. Gender differences in prostacyclin and prostaglandin E₂ synthesis by human endothelial cells. *Prostaglandins Leukot Med*. 1986;22:159-71.
- [74] Lorenz M, Koschate J, Kaufmann K, Kreye C, Mertens M, Kuebler WM, et al. Does cellular sex matter? Dimorphic transcriptional differences between female and male endothelial cells. *Atherosclerosis*. 2015;240:61-72.
- [75] Oettel A, Lorenz M, Stangl V, Costa SD, Zenclussen AC, Schumacher A. Human Umbilical Vein Endothelial Cells foster conversion of CD4⁺CD25⁻Foxp3⁻ T cells into CD4⁺Foxp3⁺ Regulatory T Cells via Transforming Growth Factor-beta. *Sci Rep*. 2016;6:23278.
- [76] Zhang YL, Cao YJ, Zhang X, Liu HH, Tong T, Xiao GD, et al. The autophagy-lysosome pathway: a novel mechanism involved in the processing of oxidized LDL in human vascular endothelial cells. *Biochem Biophys Res Commun*. 2010;394:377-82.
- [77] Zhang Y, Xie Y, You S, Han Q, Cao Y, Zhang X, et al. Autophagy and apoptosis in the response of human vascular endothelial cells to oxidized low-density lipoprotein. *Cardiology*. 2015;132:27-33.
- [78] Ling S, Dai A, Williams MR, Myles K, Dilley RJ, Komesaroff PA, et al. Testosterone (T) enhances apoptosis-related damage in human vascular endothelial cells. *Endocrinology*. 2002;143:1119-25.
- [79] Seli E, Guzeloglu-Kayisli O, Cakmak H, Kayisli UA, Selam B, Arici A. Estradiol increases apoptosis in human coronary artery endothelial cells by up-regulating Fas and Fas ligand expression. *J Clin Endocrinol Metab*. 2006;91:4995-5001.
- [80] Zhang W, Iliff JJ, Campbell CJ, Wang RK, Hurn PD, Alkayed NJ. Role of soluble epoxide hydrolase in the sex-specific vascular response to cerebral ischemia. *J Cereb Blood Flow Metab*. 2009;29:1475-81.
- [81] Gupta NC, Davis CM, Nelson JW, Young JM, Alkayed NJ. Soluble epoxide hydrolase: sex differences and role in endothelial cell survival. *Arterioscler Thromb Vasc Biol*. 2012;32:1936-42.
- [82] Kayisli UA, Guzeloglu-Kayisli O, Guzel E, Arici A. Genistein inhibits cell proliferation and stimulates apoptosis in human coronary artery endothelial cells. *Gynecol Obstet Invest*. 2013;75:235-42.
- [83] Matarrese P, Colasanti T, Ascione B, Margutti P, Franconi F, Alessandri C, et al. Gender disparity in susceptibility to oxidative stress and apoptosis induced by autoantibodies specific to RLIP76 in vascular cells. *Antioxid Redox Signal*. 2011;15:2825-36.
- [84] Martinet W, De Meyer GR. Autophagy in atherosclerosis: a cell survival and death phenomenon with therapeutic potential. *Circ Res*. 2009;104:304-17.

- [85] Liu H, Cao Y, Tong T, Shi J, Zhang Y, Yang Y, et al. Autophagy in atherosclerosis: a phenomenon found in human carotid atherosclerotic plaques. *Chin Med J (Engl)*. 2015;128:69-74.
- [86] Torisu T, Torisu K, Lee IH, Liu J, Malide D, Combs CA, et al. Autophagy regulates endothelial cell processing, maturation and secretion of von Willebrand factor. *Nat Med*. 2013;19:1281-7.
- [87] Shan H, Guo D, Li X, Zhao X, Li W, Bai X. From autophagy to senescence and apoptosis in Angiotensin II-treated vascular endothelial cells. *APMIS*. 2014;122:985-92.
- [88] Chen F, Chen B, Xiao FQ, Wu YT, Wang RH, Sun ZW, et al. Autophagy protects against senescence and apoptosis via the RAS-mitochondria in high-glucose-induced endothelial cells. *Cell Physiol Biochem*. 2014;33:1058-74.
- [89] Tai S, Hu XQ, Peng DQ, Zhou SH, Zheng XL. The roles of autophagy in vascular smooth muscle cells. *Int J Cardiol*. 2016;211:1-6.
- [90] Rzuclido EM, Martin KA, Powell RJ. Regulation of vascular smooth muscle cell differentiation. *J Vasc Surg*. 2007;45 Suppl A:A25-32.
- [91] Lao KH, Zeng L, Xu Q. Endothelial and smooth muscle cell transformation in atherosclerosis. *Curr Opin Lipidol*. 2015;26:449-56.
- [92] Hodges YK, Richer JK, Horwitz KB, Horwitz LD. Variant estrogen and progesterone receptor messages in human vascular smooth muscle. *Circulation*. 1999;99:2688-93.
- [93] Mendelsohn ME, Karas RH. Estrogen and the blood vessel wall. *Curr Opin Cardiol*. 1994;9:619-26.
- [94] Hodges YK, Tung L, Yan XD, Graham JD, Horwitz KB, Horwitz LD. Estrogen receptors alpha and beta: prevalence of estrogen receptor beta mRNA in human vascular smooth muscle and transcriptional effects. *Circulation*. 2000;101:1792-8.
- [95] Christian RC, Liu PY, Harrington S, Ruan M, Miller VM, Fitzpatrick LA. Intimal estrogen receptor (ER)beta, but not ERalpha expression, is correlated with coronary calcification and atherosclerosis in pre- and postmenopausal women. *J Clin Endocrinol Metab*. 2006;91:2713-20.
- [96] Nakamura Y, Suzuki T, Miki Y, Tazawa C, Senzaki K, Moriya T, et al. Estrogen receptors in atherosclerotic human aorta: inhibition of human vascular smooth muscle cell proliferation by estrogens. *Mol Cell Endocrinol*. 2004;219:17-26.
- [97] Occhioni S, Campesi I, Montella A, Capobianco G, Dessole S, Franconi F. Sex differences in autophagy of VSMCs from human male and female neonates 37° Congresso Nazionale della Società Italiana di Farmacologia. Napoli, Italy2015.
- [98] Montague CR, Hunter MG, Gavrillin MA, Phillips GS, Goldschmidt-Clermont PJ, Marsh CB. Activation of estrogen receptor-alpha reduces aortic smooth muscle differentiation. *Circ Res*. 2006;99:477-84.
- [99] Nakamura Y, Suzuki T, Inoue T, Tazawa C, Ono K, Moriya T, et al. Progesterone receptor subtypes in vascular smooth muscle cells of human aorta. *Endocr J*. 2005;52:245-52.
- [100] Williams MR, Ling S, Dawood T, Hashimura K, Dai A, Li H, et al. Dehydroepiandrosterone inhibits human vascular smooth muscle cell proliferation independent of ARs and ERs. *J Clin Endocrinol Metab*. 2002;87:176-81.
- [101] Dubey RK, Gillespie DG, Imthurn B, Rosselli M, Jackson EK, Keller PJ. Phytoestrogens inhibit growth and MAP kinase activity in human aortic smooth muscle cells. *Hypertension*. 1999;33:177-82.
- [102] Thompson J, Khalil RA. Gender differences in the regulation of vascular tone. *Clin Exp Pharmacol Physiol*. 2003;30:1-15.
- [103] Natoli AK, Medley TL, Ahimastos AA, Drew BG, Thearle DJ, Dilley RJ, et al. Sex steroids modulate human aortic smooth muscle cell matrix protein deposition and matrix metalloproteinase expression. *Hypertension*. 2005;46:1129-34.
- [104] Nakamura Y, Suzuki T, Sasano H. Estrogen actions and in situ synthesis in human vascular smooth muscle cells and their correlation with atherosclerosis. *J Steroid Biochem Mol Biol*. 2005;93:263-8.
- [105] Jia G, Cheng G, Agrawal DK. Autophagy of vascular smooth muscle cells in atherosclerotic lesions. *Autophagy*. 2007;3:63-4.

- [106] Perrotta I, Aquila S. The role of oxidative stress and autophagy in atherosclerosis. *Oxid Med Cell Longev*. 2015;2015:130315.
- [107] Kolodgie FD, Narula J, Haider N, Virmani R. Apoptosis in atherosclerosis. Does it contribute to plaque instability? *Cardiol Clin*. 2001;19:127-39.
- [108] Chawla-Sarkar M, Lindner DJ, Liu YF, Williams BR, Sen GC, Silverman RH, et al. Apoptosis and interferons: role of interferon-stimulated genes as mediators of apoptosis. *Apoptosis*. 2003;8:237-49.
- [109] Kavurma MM, Tan NY, Bennett MR. Death receptors and their ligands in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2008;28:1694-702.
- [110] Seimon T, Tabas I. Mechanisms and consequences of macrophage apoptosis in atherosclerosis. *J Lipid Res*. 2009;50 Suppl:S382-7.
- [111] Swiader A, Nahapetyan H, Faccini J, D'Angelo R, Mucher E, Elbaz M, et al. Mitophagy acts as a safeguard mechanism against human vascular smooth muscle cell apoptosis induced by atherogenic lipids. *Oncotarget*. 2016.
- [112] Malorni W, Straface E, Matarrese P, Ascione B, Coinu R, Canu S, et al. Redox state and gender differences in vascular smooth muscle cells. *FEBS Lett*. 2008;582:635-42.
- [113] Straface E, Vona R, Campesi I, Franconi F. Mitochondria can orchestrate sex differences in cell fate of vascular smooth muscle cells from rats. *Biol Sex Differ*. 2015;6:34.
- [114] Straface E, Vona R, Gambardella L, Ascione B, Marino M, Bulzomi P, et al. Cell sex determines anoikis resistance in vascular smooth muscle cells. *FEBS Lett*. 2009;583:3448-54.
- [115] Salabei JK, Hill BG. Autophagic regulation of smooth muscle cell biology. *Redox Biol*. 2015;4:97-103.
- [116] Kiffin R, Bandyopadhyay U, Cuervo AM. Oxidative stress and autophagy. *Antioxid Redox Signal*. 2006;8:152-62.
- [117] Martinet W, Schrijvers DM, Timmermans JP, Bult H. Interactions between cell death induced by statins and 7-ketocholesterol in rabbit aorta smooth muscle cells. *Br J Pharmacol*. 2008;154:1236-46.
- [118] Campesi I, Occhioni S, Capobianco G, Fois M, Montella A, Dessole S, et al. Sex-specific pharmacological modulation of autophagic process in human umbilical artery smooth muscle cells. *Pharmacol Res*. 2016.
- [119] Lindemann S, Kramer B, Seizer P, Gawaz M. Platelets, inflammation and atherosclerosis. *J Thromb Haemost*. 2007;5 Suppl 1:203-11.
- [120] Gawaz M. Platelets in the onset of atherosclerosis. *Blood Cells Mol Dis*. 2006;36:206-10.
- [121] Khetawat G, Faraday N, Nealen ML, Vijayan KV, Bolton E, Noga SJ, et al. Human megakaryocytes and platelets contain the estrogen receptor beta and androgen receptor (AR): testosterone regulates AR expression. *Blood*. 2000;95:2289-96.
- [122] Jayachandran M, Miller VM. Human platelets contain estrogen receptor alpha, caveolin-1 and estrogen receptor associated proteins. *Platelets*. 2003;14:75-81.
- [123] Moro L, Reineri S, Piranda D, Pietrapiana D, Lova P, Bertoni A, et al. Nongenomic effects of 17beta-estradiol in human platelets: potentiation of thrombin-induced aggregation through estrogen receptor beta and Src kinase. *Blood*. 2005;105:115-21.
- [124] Kueviakoe IM, Segbena AY, Jouault H, Vovor A, Imbert M. Hematological reference values for healthy adults in togo. *ISRN Hematol*. 2011;2011:736062.
- [125] Jaremo P, Eriksson-Franzen M, Milovanovic M. Platelets, gender and acute cerebral infarction. *J Transl Med*. 2015;13:267-70.
- [126] Santimone I, Di Castelnuovo A, De Curtis A, Spinelli M, Cugino D, Gianfagna F, et al. White blood cell count, sex and age are major determinants of heterogeneity of platelet indices in an adult general population: results from the MOLI-SANI project. *Haematologica*. 2011;96:1180-8.
- [127] Segal JB, Moliterno AR. Platelet counts differ by sex, ethnicity, and age in the United States. *Ann Epidemiol*. 2006;16:123-30.
- [128] Butkiewicz AM, Kemoni H, Dymicka-Piekarska V, Matowicka-Karna J, Radziwon P, Lipska A. Platelet count, mean platelet volume and thrombocytopenic indices in healthy women and men. *Thromb Res*. 2006;118:199-204.

- [129] Sloan A, Gona P, Johnson AD. Cardiovascular correlates of platelet count and volume in the Framingham Heart Study. *Ann Epidemiol.* 2015;25:492-8.
- [130] Biino G, Santimone I, Minelli C, Sorice R, Frongia B, Traglia M, et al. Age- and sex-related variations in platelet count in Italy: a proposal of reference ranges based on 40987 subjects' data. *PLoS One.* 2013;8:e54289.
- [131] Al-Chalaby SH, Taib SM, Ahmed AF. The effect of oral contraceptive pills on haematological indices. *Tikrit Med J.* 2006;12:65-9.
- [132] Patti G, De Caterina R, Abbate R, Andreotti F, Biasucci LM, Calabro P, et al. Platelet function and long-term antiplatelet therapy in women: is there a gender-specificity? A 'state-of-the-art' paper. *Eur Heart J.* 2014;35:2213-23b.
- [133] Lawrence JB, Leifer DW, Moura GL, Southern P, Emery JD, Bodenheimer SL, et al. Sex differences in platelet adherence to subendothelium: relationship to platelet function tests and hematologic variables. *Am J Med Sci.* 1995;309:201-7.
- [134] Juan P, Stefano G, Antonella S, Albana C. Platelets in pregnancy. *J Prenat Med.* 2011;5:90-2.
- [135] Chu SG, Becker RC, Berger PB, Bhatt DL, Eikelboom JW, Konkle B, et al. Mean platelet volume as a predictor of cardiovascular risk: a systematic review and meta-analysis. *J Thromb Haemost.* 2010;8:148-56.
- [136] Campesi I, Carru C, Zinellu A, Occhioni S, Sanna M, Palermo M, et al. Regular cigarette smoking influences the transsulfuration pathway, endothelial function, and inflammation biomarkers in a sex-gender specific manner in healthy young humans. *Am J Transl Res.* 2013;5:497-509.
- [137] Green ED, Green P. Sequence-tagged site (STS) content mapping of human chromosomes: theoretical considerations and early experiences. *PCR Methods Appl.* 1991;1:77-90.
- [138] Aghaji M, Nnabuko R, Uzuegbunam C, Oyeka IC. The relationship of white blood cell and platelet counts to cigarette smoking in adult Nigerians. *Cent Afr J Med.* 1990;36:273-8.
- [139] Wang TY, Angiolillo DJ, Cushman M, Sabatine MS, Bray PF, Smyth SS, et al. Platelet biology and response to antiplatelet therapy in women: implications for the development and use of antiplatelet pharmacotherapies for cardiovascular disease. *J Am Coll Cardiol.* 2012;59:891-900.
- [140] Schubert P, Coupland D, Nombalais M, G MW, Devine DV. RhoA/ROCK signaling contributes to sex differences in the activation of human platelets. *Thromb Res.* 2016;139:50-5.
- [141] Johnson M, Ramey E, Ramwell PW. Sex and age differences in human platelet aggregation. *Nature.* 1975;253:355-7.
- [142] Faraday N, Goldschmidt-Clermont PJ, Bray PF. Gender differences in platelet GPIIb-IIIa activation. *Thromb Haemost.* 1997;77:748-54.
- [143] Gremmel T, Kopp CW, Eichelberger B, Koppensteiner R, Panzer S. Sex differences of leukocyte-platelet interactions and on-treatment platelet reactivity in patients with atherosclerosis. *Atherosclerosis.* 2014;237:692-5.
- [144] Samad Z, Boyle S, Ersboll M, Vora AN, Zhang Y, Becker RC, et al. Sex differences in platelet reactivity and cardiovascular and psychological response to mental stress in patients with stable ischemic heart disease: insights from the REMIT study. *J Am Coll Cardiol.* 2014;64:1669-78.
- [145] Cho YU, Chi HS, Jang S, Park CJ. Reconfirmation of preanalytical variables and establishment of reference intervals of platelet function analyzer-100 closure times in Korean adults. *Korean J Lab Med.* 2007;27:318-23.
- [146] Sestito A, Sciahbasi A, Landolfi R, Maseri A, Lanza GA, Andreotti F. A simple assay for platelet-mediated hemostasis in flowing whole blood (PFA-100): reproducibility and effects of sex and age. *Cardiologia.* 1999;44:661-5.
- [147] Basili S, Raparelli V, Proietti M, Tanzilli G, Franconi F. Impact of sex and gender on the efficacy of antiplatelet therapy: the female perspective. *J Atheroscler Thromb.* 2015;22:109-25.
- [148] Gambardella L, Vona R, Pichini S, Pacifici R, Malorni W, Straface E. Gender difference in platelet aggregation and reactivity induced by recombinant human erythropoietin. *Br J Clin Pharmacol.* 2015.
- [149] Gawaz M, Langer H, May AE. Platelets in inflammation and atherogenesis. *J Clin Invest.* 2005;115:3378-84.

- [150] Huo Y, Ley KF. Role of platelets in the development of atherosclerosis. *Trends Cardiovasc Med*. 2004;14:18-22.
- [151] Koupenova M, Mick E, Mikhalev E, Benjamin EJ, Tanriverdi K, Freedman JE. Sex differences in platelet toll-like receptors and their association with cardiovascular risk factors. *Arterioscler Thromb Vasc Biol*. 2015;35:1030-7.
- [152] Duman RS. Novel therapeutic approaches beyond the serotonin receptor. *Biol Psychiatry*. 1998;44:324-35.
- [153] Kaess BM, Preis SR, Lieb W, Beiser AS, Yang Q, Chen TC, et al. Circulating brain-derived neurotrophic factor concentrations and the risk of cardiovascular disease in the community. *J Am Heart Assoc*. 2015;4:e001544.
- [154] Lommatzsch M, Zingler D, Schuhbaeck K, Schloetcke K, Zingler C, Schuff-Werner P, et al. The impact of age, weight and gender on BDNF levels in human platelets and plasma. *Neurobiol Aging*. 2005;26:115-23.
- [155] Williams MS, Ngongang CK, Ouyang P, Betoudji F, Harrer C, Wang NY, et al. Gender differences in platelet brain derived neurotrophic factor in patients with cardiovascular disease and depression. *J Psychiatr Res*. 2016;78:72-7.
- [156] Kurrelmeyer K, Becker L, Becker D, Yanek L, Goldschmidt-Clermont P, Bray PF. Platelet hyperreactivity in women from families with premature atherosclerosis. *J Am Med Womens Assoc*. 2003;58:272-7.
- [157] Roell A, Schueller P, Schultz A, Losel R, Wehling M, Christ M, et al. Effect of oral contraceptives and ovarian cycle on platelet function. *Platelets*. 2007;18:165-70.
- [158] Tarantino MD, Kunicki TJ, Nugent DJ. The estrogen receptor is present in human megakaryocytes. *Ann N Y Acad Sci*. 1994;714:293-6.
- [159] Eidelman O, Jozwik C, Huang W, Srivastava M, Rothwell SW, Jacobowitz DM, et al. Gender dependence for a subset of the low-abundance signaling proteome in human platelets. *Hum Genomics Proteomics*. 2010;2010:164906.
- [160] Jones SB, Bylund DB, Rieser CA, Shekim WO, Byer JA, Carr GW. alpha 2-Adrenergic receptor binding in human platelets: alterations during the menstrual cycle. *Clin Pharmacol Ther*. 1983;34:90-6.
- [161] Yee DL, Sun CW, Bergeron AL, Dong JF, Bray PF. Aggregometry detects platelet hyperreactivity in healthy individuals. *Blood*. 2005;106:2723-9.
- [162] Norris LA, Bonnar J. Effect of oestrogen dose on whole blood platelet activation in women taking new low dose oral contraceptives. *Thromb Haemost*. 1994;72:926-30.
- [163] Braunstein JB, Kershner DW, Bray P, Gerstenblith G, Schulman SP, Post WS, et al. Interaction of hemostatic genetics with hormone therapy: new insights to explain arterial thrombosis in postmenopausal women. *Chest*. 2002;121:906-20.
- [164] Thijs A, van Baal WM, van der Mooren MJ, Kenemans P, Drager AM, Huijgens PC, et al. Effects of hormone replacement therapy on blood platelets. *Eur J Clin Invest*. 2002;32:613-8.
- [165] Teede HJ, McGrath BP, Turner A, Majewski H. Effects of oral combined hormone replacement therapy on platelet aggregation in postmenopausal women. *Clin Sci (Lond)*. 2001;100:207-13.
- [166] Brenner B. Haemostatic changes in pregnancy. *Thromb Res*. 2004;114:409-14.
- [167] Kazmi RS, Cooper AJ, Lwaleed BA. Platelet function in pre-eclampsia. *Semin Thromb Hemost*. 2011;37:131-6.
- [168] Badrnya S, Schrottmaier WC, Kral JB, Yaiw KC, Volf I, Schabbauer G, et al. Platelets mediate oxidized low-density lipoprotein-induced monocyte extravasation and foam cell formation. *Arterioscler Thromb Vasc Biol*. 2014;34:571-80.
- [169] Daub K, Seizer P, Stellos K, Kramer BF, Bigalke B, Schaller M, et al. Oxidized LDL-activated platelets induce vascular inflammation. *Semin Thromb Hemost*. 2010;36:146-56.
- [170] Thushara RM, Hemshekhar M, Basappa, Kemparaju K, Rangappa KS, Girish KS. Biologicals, platelet apoptosis and human diseases: An outlook. *Crit Rev Oncol Hematol*. 2015;93:149-58.
- [171] You T, Wang Q, Zhu L. Role of autophagy in megakaryocyte differentiation and platelet formation. *Int J Physiol Pathophysiol Pharmacol*. 2016;8:28-34.

- [172] Lee SH, Du J, Stitham J, Atteya G, Lee S, Xiang Y, et al. Inducing mitophagy in diabetic platelets protects against severe oxidative stress. *EMBO Mol Med.* 2016;8:779-95.
- [173] Ouseph MM, Huang Y, Banerjee M, Joshi S, MacDonald L, Zhong Y, et al. Autophagy is induced upon platelet activation and is essential for hemostasis and thrombosis. *Blood.* 2015;126:1224-33.
- [174] Blum A. Endothelial progenitor cells are affected by medications and estrogen. *Isr Med Assoc J.* 2015;17:578-80.
- [175] Foresta C, Zuccarello D, Biagioli A, De Toni L, Prana E, Nicoletti V, et al. Oestrogen stimulates endothelial progenitor cells via oestrogen receptor-alpha. *Clin Endocrinol (Oxf).* 2007;67:520-5.
- [176] Foresta C, De Toni L, Di Mambro A, Ferlin A, Perilli L, Bertuzzi I, et al. Role of estrogen receptors in menstrual cycle-related neoangiogenesis and their influence on endothelial progenitor cell physiology. *Fertil Steril.* 2010;93:220-8.
- [177] Foresta C, Zuccarello D, De Toni L, Garolla A, Caretta N, Ferlin A. Androgens stimulate endothelial progenitor cells through an androgen receptor-mediated pathway. *Clin Endocrinol (Oxf).* 2008;68:284-9.
- [178] Rehman J, Li J, Orschell CM, March KL. Peripheral blood "endothelial progenitor cells" are derived from monocyte/macrophages and secrete angiogenic growth factors. *Circulation.* 2003;107:1164-9.
- [179] Kikuchi-Taura A, Soma T, Matsuyama T, Stern DM, Taguchi A. A new protocol for quantifying CD34(+) cells in peripheral blood of patients with cardiovascular disease. *Tex Heart Inst J.* 2006;33:427-9.
- [180] Rousseau A, Ayoubi F, Deveaux C, Charbit B, Delmau C, Christin-Maitre S, et al. Impact of age and gender interaction on circulating endothelial progenitor cells in healthy subjects. *Fertil Steril.* 2010;93:843-6.
- [181] Jiang YP, Zeng GF, Ren Z, Zeng HT, Yang Z. Sex differences in the number and activity of circulating endothelial progenitor cells in prehypertension. *Chin J Tissue Eng Res* 2015;19:3061-6.
- [182] Zhen Y, Xiao S, Ren Z, Shen HW, Su H, Tang YB, et al. Increased endothelial progenitor cells and nitric oxide in young prehypertensive women. *J Clin Hypertens (Greenwich).* 2015;17:298-305.
- [183] Ruszkowska-Ciastek B, Sokup A, Leszcz M, Drela E, Stankowska K, Boinska J, et al. The number of circulating endothelial progenitor cells in healthy individuals--effect of some anthropometric and environmental factors (a pilot study). *Adv Med Sci.* 2015;60:58-63.
- [184] Stauffer BL, Maceneaney OJ, Kushner EJ, Cech JN, Greiner JJ, Westby CM, et al. Gender and endothelial progenitor cell number in middle-aged adults. *Artery Res.* 2008;2:156-60.
- [185] Chen CH, Cheng BC, Leu S, Sun CK, Chua S, Yen CH, et al. Circulating level of endothelial progenitor cells in healthy Taiwanese. *Acta Cardiol Sin.* 2010;26:94-101.
- [186] Ricottini E, Madonna R, Grieco D, Zoccoli A, Stampachiacchiere B, Patti G, et al. Effect of high-dose Atorvastatin reload on the release of endothelial progenitor cells in patients on long-term statin treatment who underwent percutaneous coronary intervention (from the ARMYDA-EPC Study). *Am J Cardiol.* 2016;117:165-71.
- [187] Oikonomou E, Siasos G, Zaromitidou M, Hatzis G, Mourouzis K, Chrysohoou C, et al. Atorvastatin treatment improves endothelial function through endothelial progenitor cells mobilization in ischemic heart failure patients. *Atherosclerosis.* 2015;238:159-64.
- [188] Lin CP, Lin FY, Huang PH, Chen YL, Chen WC, Chen HY, et al. Endothelial progenitor cell dysfunction in cardiovascular diseases: role of reactive oxygen species and inflammation. *Biomed Res Int.* 2013;2013:845037.
- [189] Pelliccia F, Pasceri V, Cianfrocca C, Vitale C, Meoni G, Pristipino C, et al. Circulating endothelial progenitor cells in postmenopausal women with and without coronary artery disease. *Climacteric.* 2009;12:259-65.
- [190] Yue WS, Wang M, Yan GH, Yiu KH, Yin L, Lee SW, et al. Smoking is associated with depletion of circulating endothelial progenitor cells and elevated pulmonary artery systolic pressure in patients with coronary artery disease. *Am J Cardiol.* 2010;106:1248-54.
- [191] Werner N, Kosiol S, Schiegl T, Ahlers P, Walenta K, Link A, et al. Circulating endothelial progenitor cells and cardiovascular outcomes. *N Engl J Med.* 2005;353:999-1007.
- [192] Mobarrez F, Antoniewicz L, Bosson JA, Kuhl J, Pisetsky DS, Lundback M. The effects of smoking on levels of endothelial progenitor cells and microparticles in the blood of healthy volunteers. *PLoS One.* 2014;9:e90314.

- [193] Kondo T, Hayashi M, Takeshita K, Numaguchi Y, Kobayashi K, Iino S, et al. Smoking cessation rapidly increases circulating progenitor cells in peripheral blood in chronic smokers. *Arterioscler Thromb Vasc Biol.* 2004;24:1442-7.
- [194] Hoetzer GL, MacEneaney OJ, Irmiger HM, Keith R, Van Guilder GP, Stauffer BL, et al. Gender differences in circulating endothelial progenitor cell colony-forming capacity and migratory activity in middle-aged adults. *Am J Cardiol.* 2007;99:46-8.
- [195] Lemieux C, Cloutier I, Tanguay JF. Menstrual cycle influences endothelial progenitor cell regulation: a link to gender differences in vascular protection? *Int J Cardiol.* 2009;136:200-10.
- [196] Liao CH, Wu YN, Lin FY, Tsai WK, Liu SP, Chiang HS. Testosterone replacement therapy can increase circulating endothelial progenitor cell number in men with late onset hypogonadism. *Andrology.* 2013;1:563-9.
- [197] Aicher A, Zeiher AM, Dimmeler S. Mobilizing endothelial progenitor cells. *Hypertension.* 2005;45:321-5.
- [198] Patel PD, Arora RR. Endothelial dysfunction: a potential tool in gender related cardiovascular disease. *Ther Adv Cardiovasc Dis.* 2008;2:89-100.
- [199] Bacon SL, Lavoie KL, Arsenault A, Dupuis J, Pilote L, Laurin C, et al. The research on endothelial function in women and men at risk for cardiovascular disease (REWARD) study: methodology. *BMC Cardiovasc Disord.* 2011;11:50-8.
- [200] Apostolakis S, Spandidos D. Chemokines and atherosclerosis: focus on the CX3CL1/CX3CR1 pathway. *Acta Pharmacol Sin.* 2013;34:1251-6.
- [201] Leung J, Jayachandran M, Kendall-Thomas J, Behrenbeck T, Araoz P, Miller VM. Pilot study of sex differences in chemokine/cytokine markers of atherosclerosis in humans. *Gend Med.* 2008;5:44-52.
- [202] Mansfield AS, Nevala WK, Dronca RS, Leontovich AA, Shuster L, Markovic SN. Normal ageing is associated with an increase in Th2 cells, MCP-1 (CCL1) and RANTES (CCL5), with differences in sCD40L and PDGF-AA between sexes. *Clin Exp Immunol.* 2012;170:186-93.
- [203] Li J, Wang Y, Lin J, Wang D, Wang A, Zhao X, et al. Soluble CD40L is a useful marker to predict future strokes in patients with minor stroke and transient ischemic attack. *Stroke.* 2015;46:1990-2.
- [204] Sirois-Gagnon D, Chamberland A, Perron S, Brisson D, Gaudet D, Laprise C. Association of common polymorphisms in the fractalkine receptor (CX3CR1) with obesity. *Obesity (Silver Spring).* 2011;19:222-7.
- [205] Hsieh MM, Everhart JE, Byrd-Holt DD, Tisdale JF, Rodgers GP. Prevalence of neutropenia in the U.S. population: age, sex, smoking status, and ethnic differences. *Ann Intern Med.* 2007;146:486-92.
- [206] Madhura M, Doniya J. Cyclical variation of leucocyte profile in healthy females: a comparison with males. *Inter J Biomed Res* 2014;5:257-9.
- [207] Nilsson G, Hedberg P, Ohrvik J. White blood cell count in elderly is clinically useful in predicting long-term survival. *J Aging Res.* 2014;2014:475093.
- [208] Chmielewski PP, Borysławski K, Chmielowiec K, Chmielowiec J, Strzelec B. The association between total leukocyte count and longevity: Evidence from longitudinal and cross-sectional data *Ann Anat.* 2016;204:1-10.
- [209] Smith MR, Kinmonth AL, Luben RN, Bingham S, Day NE, Wareham NJ, et al. Smoking status and differential white cell count in men and women in the EPIC-Norfolk population. *Atherosclerosis.* 2003;169:331-7.
- [210] Yarnell JW, Sweetnam PM, Rogers S, Elwood PC, Bainton D, Baker IA, et al. Some long term effects of smoking on the haemostatic system: a report from the Caerphilly and Speedwell Collaborative Surveys. *J Clin Pathol.* 1987;40:909-13.
- [211] Jensen EJ, Pedersen B, Frederiksen R, Dahl R. Prospective study on the effect of smoking and nicotine substitution on leucocyte blood counts and relation between blood leucocytes and lung function. *Thorax.* 1998;53:784-9.
- [212] Pratley RE, Wilson C, Bogardus C. Relation of the white blood cell count to obesity and insulin resistance: effect of race and gender. *Obes Res.* 1995;3:563-71.
- [213] Ruggiero C, Metter EJ, Cherubini A, Maggio M, Sen R, Najjar SS, et al. White blood cell count and mortality in the Baltimore Longitudinal Study of Aging. *J Am Coll Cardiol.* 2007;49:1841-50.

- [214] Gwak MS, Choi SJ, Kim JA, Ko JS, Kim TH, Lee SM, et al. Effects of gender on white blood cell populations and neutrophil-lymphocyte ratio following gastrectomy in patients with stomach cancer. *J Korean Med Sci.* 2007;22 Suppl:S104-8.
- [215] Orysiak J, Witek K, Zmijewski P, Gajewski J. White blood cells in Polish athletes of various sports disciplines. *Biol Sport.* 2012;29:101-5.
- [216] McGarry J. Letter: Smoking and leukocyte counts in pregnancy. *Br Med J.* 1974;1:160.
- [217] Rana JS, Boekholdt SM, Ridker PM, Jukema JW, Luben R, Bingham SA, et al. Differential leucocyte count and the risk of future coronary artery disease in healthy men and women: the EPIC-Norfolk Prospective Population Study. *J Intern Med.* 2007;262:678-89.
- [218] Molero L, Garcia-Duran M, Diaz-Recasens J, Rico L, Casado S, Lopez-Farre A. Expression of estrogen receptor subtypes and neuronal nitric oxide synthase in neutrophils from women and men: regulation by estrogen. *Cardiovasc Res.* 2002;56:43-51.
- [219] Mantalaris A, Panoskaltis N, Sakai Y, Bourne P, Chang C, Messing EM, et al. Localization of androgen receptor expression in human bone marrow. *J Pathol.* 2001;193:361-6.
- [220] Walker AE, Seibert SM, Donato AJ, Pierce GL, Seals DR. Vascular endothelial function is related to white blood cell count and myeloperoxidase among healthy middle-aged and older adults. *Hypertension.* 2010;55:363-9.
- [221] Guasti L, Dentali F, Castiglioni L, Maroni L, Marino F, Squizzato A, et al. Neutrophils and clinical outcomes in patients with acute coronary syndromes and/or cardiac revascularisation. A systematic review on more than 34,000 subjects. *Thromb Haemost.* 2011;106:591-9.
- [222] Wheeler JG, Mussolino ME, Gillum RF, Danesh J. Associations between differential leucocyte count and incident coronary heart disease: 1764 incident cases from seven prospective studies of 30,374 individuals. *Eur Heart J.* 2004;25:1287-92.
- [223] Arruda-Olson AM, Reeder GS, Bell MR, Weston SA, Roger VL. Neutrophilia predicts death and heart failure after myocardial infarction: a community-based study. *Circ Cardiovasc Qual Outcomes.* 2009;2:656-62.
- [224] Chen J, Chen MH, Li S, Guo YL, Zhu CG, Xu RX, et al. Usefulness of the neutrophil-to-lymphocyte ratio in predicting the severity of coronary artery disease: a Gensini score assessment. *J Atheroscler Thromb.* 2014;21:1271-82.
- [225] Casimir GJ, Mulier S, Hanssens L, Zylberberg K, Duchateau J. Gender differences in inflammatory markers in children. *Shock.* 2010;33:258-62.
- [226] Bain BJ. Ethnic and sex differences in the total and differential white cell count and platelet count. *J Clin Pathol.* 1996;49:664-6.
- [227] Bain BJ, England JM. Normal haematological values: sex difference in neutrophil count. *Br Med J.* 1975;1:306-9.
- [228] Bouman A, Heineman MJ, Faas MM. Sex hormones and the immune response in humans. *Hum Reprod Update.* 2005;11:411-23.
- [229] Efrati P, Presentey B, Margalith M, Rozenszajn L. Leukocytes of normal pregnant women. *Obstet Gynecol.* 1964;23:429-32.
- [230] Watson RW, O'Neill A, Brannigan AE, Coffey R, Marshall JC, Brady HR, et al. Regulation of Fas antibody induced neutrophil apoptosis is both caspase and mitochondrial dependent. *FEBS Lett.* 1999;453:67-71.
- [231] Tulchinsky D, Hobel CJ. Plasma human chorionic gonadotropin, estrone, estradiol, estriol, progesterone, and 17 alpha-hydroxyprogesterone in human pregnancy. 3. Early normal pregnancy. *Am J Obstet Gynecol.* 1973;117:884-93.
- [232] von Dadelszen P, Watson RW, Noorwali F, Marshall JC, Parodo J, Farine D, et al. Maternal neutrophil apoptosis in normal pregnancy, preeclampsia, and normotensive intrauterine growth restriction. *Am J Obstet Gynecol.* 1999;181:408-14.
- [233] Pende A, Artom N, Bertolotto M, Montecucco F, Dallegri F. Role of neutrophils in atherogenesis: an update. *Eur J Clin Invest.* 2015.

- [234] Tavora FR, Ripple M, Li L, Burke AP. Monocytes and neutrophils expressing myeloperoxidase occur in fibrous caps and thrombi in unstable coronary plaques. *BMC Cardiovasc Disord.* 2009;9:27.
- [235] Larionov S, Dedek O, Birkenmeier G, Thal DR. Expression of alpha2-macroglobulin, neutrophil elastase, and interleukin-1alpha differs in early-stage and late-stage atherosclerotic lesions in the arteries of the circle of Willis. *Acta Neuropathol.* 2007;113:33-43.
- [236] Ionita MG, van den Borne P, Catanzariti LM, Moll FL, de Vries JP, Pasterkamp G, et al. High neutrophil numbers in human carotid atherosclerotic plaques are associated with characteristics of rupture-prone lesions. *Arterioscler Thromb Vasc Biol.* 2010;30:1842-8.
- [237] Aomatsu M, Kato T, Kasahara E, Kitagawa S. Gender difference in tumor necrosis factor-alpha production in human neutrophils stimulated by lipopolysaccharide and interferon-gamma. *Biochem Biophys Res Commun.* 2013;441:220-5.
- [238] Aoyama M, Kotani J, Usami M. Gender difference in granulocyte dynamics and apoptosis and the role of IL-18 during endotoxin-induced systemic inflammation. *Shock.* 2009;32:401-9.
- [239] Moxley G, Stern AG, Carlson P, Estrada E, Han J, Benson LL. Premenopausal sexual dimorphism in lipopolysaccharide-stimulated production and secretion of tumor necrosis factor. *J Rheumatol.* 2004;31:686-94.
- [240] Imahara SD, Jelacic S, Junker CE, O'Keefe GE. The influence of gender on human innate immunity. *Surgery.* 2005;138:275-82.
- [241] Bauer I, Bauer M, Raddatz A, Luedtke C, Werth M, Silomon M, et al. [Influence of gender on stimulated cytokine response in patients with severe sepsis]. *Anaesthesist.* 2006;55:515-27.
- [242] Casimir GJ, Heldenbergh F, Hanssens L, Mulier S, Heinrichs C, Lefevre N, et al. Gender differences and inflammation: an in vitro model of blood cells stimulation in prepubescent children. *J Inflamm (Lond).* 2010;7:28.
- [243] Pergola C, Schaible AM, Nikels F, Dodt G, Northoff H, Werz O. Progesterone rapidly down-regulates the biosynthesis of 5-lipoxygenase products in human primary monocytes. *Pharmacol Res.* 2015;94:42-50.
- [244] Pergola C, Dodt G, Rossi A, Neunhoeffler E, Lawrenz B, Northoff H, et al. ERK-mediated regulation of leukotriene biosynthesis by androgens: a molecular basis for gender differences in inflammation and asthma. *Proc Natl Acad Sci U S A.* 2008;105:19881-6.
- [245] Back M, Weber C, Lutgens E. Regulation of atherosclerotic plaque inflammation. *J Intern Med.* 2015;278:462-82.
- [246] Garcia-Duran M, de Frutos T, Diaz-Recasens J, Garcia-Galvez G, Jimenez A, Monton M, et al. Estrogen stimulates neuronal nitric oxide synthase protein expression in human neutrophils. *Circ Res.* 1999;85:1020-6.
- [247] de Coupade C, Gear RW, Dazin PF, Sroussi HY, Green PG, Levine JD. Beta 2-adrenergic receptor regulation of human neutrophil function is sexually dimorphic. *Br J Pharmacol.* 2004;143:1033-41.
- [248] Molloy EJ, O'Neill AJ, Grantham JJ, Sheridan-Pereira M, Fitzpatrick JM, Webb DW, et al. Sex-specific alterations in neutrophil apoptosis: the role of estradiol and progesterone. *Blood.* 2003;102:2653-9.
- [249] Martin KR, Ohayon D, Witko-Sarsat V. Promoting apoptosis of neutrophils and phagocytosis by macrophages: novel strategies in the resolution of inflammation. *Swiss Med Wkly.* 2015;145:w14056.
- [250] Chargui A, El May MV. Autophagy mediates neutrophil responses to bacterial infection. *Apmis.* 2014;122:1047-58.
- [251] Ramachandran G, Gade P, Tsai P, Lu W, Kalvakolanu DV, Rosen GM, et al. Potential role of autophagy in the bactericidal activity of human PMNs for *Bacillus anthracis*. *Pathog Dis.* 2015;73:ftv080.
- [252] Kavathia N, Jain A, Walston J, Beamer BA, Fedarko NS. Serum markers of apoptosis decrease with age and cancer stage. *Aging (Albany NY).* 2009;1:652-63.
- [253] Wu G, Wei G, Huang J, Pang S, Liu L, Yan B. Decreased gene expression of LC3 in peripheral leucocytes of patients with coronary artery disease. *Eur J Clin Invest.* 2011;41:958-63.
- [254] Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol.* 2013;13:709-21.

- [255] Phiel KL, Henderson RA, Adelman SJ, Elloso MM. Differential estrogen receptor gene expression in human peripheral blood mononuclear cell populations. *Immunol Lett.* 2005;97:107-13.
- [256] Pelekanou V, Kampa M, Kiagiadaki F, Deli A, Theodoropoulos P, Agrogiannis G, et al. Estrogen anti-inflammatory activity on human monocytes is mediated through cross-talk between estrogen receptor ERalpha36 and GPR30/GPER1. *J Leukoc Biol.* 2015.
- [257] McCrohon JA, Death AK, Nakhla S, Jessup W, Handelsman DJ, Stanley KK, et al. Androgen receptor expression is greater in macrophages from male than from female donors. A sex difference with implications for atherogenesis. *Circulation.* 2000;101:224-6.
- [258] Jensen AL, Pioli PA. Estrogen induces the expression of estrogen receptor in human macrophages. *FASEB J.* 2008;22:672-43.
- [259] Murphy AJ, Guyre PM, Wira CR, Pioli PA. Estradiol regulates expression of estrogen receptor ERalpha46 in human macrophages. *PLoS One.* 2009;4:e5539.
- [260] Blasko E, Haskell CA, Leung S, Gualtieri G, Halks-Miller M, Mahmoudi M, et al. Beneficial role of the GPR30 agonist G-1 in an animal model of multiple sclerosis. *J Neuroimmunol.* 2009;214:67-77.
- [261] Olivares R, Ducimetiere P, Claude JR. Monocyte count: a risk factor for coronary heart disease? *Am J Epidemiol.* 1993;137:49-53.
- [262] Johnsen SH, Fosse E, Joakimsen O, Mathiesen EB, Stensland-Bugge E, Njolstad I, et al. Monocyte count is a predictor of novel plaque formation: a 7-year follow-up study of 2610 persons without carotid plaque at baseline the Tromso Study. *Stroke.* 2005;36:715-9.
- [263] Starr JM, Deary IJ. Sex differences in blood cell counts in the Lothian Birth Cohort 1921 between 79 and 87 years. *Maturitas.* 2011;69:373-6.
- [264] Mathur S, Mathur RS, Goust JM, Williamson HO, Fudenberg HH. Cyclic variations in white cell subpopulations in the human menstrual cycle: correlations with progesterone and estradiol. *Clin Immunol Immunopathol.* 1979;13:246-53.
- [265] Lefevre N, Corazza F, Duchateau J, Desir J, Casimir G. Sex differences in inflammatory cytokines and CD99 expression following in vitro lipopolysaccharide stimulation. *Shock.* 2012;38:37-42.
- [266] Gavello D, Carbone E, Carabelli V. Leptin-mediated ion channel regulation: PI3K pathways, physiological role, and therapeutic potential. *Channels (Austin).* 2016:1-15.
- [267] Cannon JG, Sharma G, Sloan G, Dimitropoulou C, Baker RR, Mazzoli A, et al. Leptin regulates CD16 expression on human monocytes in a sex-specific manner. *Physiol Rep.* 2014;2.
- [268] Verdugo RA, Zeller T, Rotival M, Wild PS, Munzel T, Lackner KJ, et al. Graphical modeling of gene expression in monocytes suggests molecular mechanisms explaining increased atherosclerosis in smokers. *PLoS One.* 2013;8:e50888.
- [269] Dumeaux V, Olsen KS, Nuel G, Paulssen RH, Borresen-Dale AL, Lund E. Deciphering normal blood gene expression variation--The NOWAC postgenome study. *PLoS Genet.* 2010;6:e1000873.
- [270] Charlesworth JC, Curran JE, Johnson MP, Goring HH, Dyer TD, Diego VP, et al. Transcriptomic epidemiology of smoking: the effect of smoking on gene expression in lymphocytes. *BMC Med Genomics.* 2010;3:29.
- [271] Zeller T, Wild P, Szymczak S, Rotival M, Schillert A, Castagne R, et al. Genetics and beyond--the transcriptome of human monocytes and disease susceptibility. *PLoS One.* 2010;5:e10693.
- [272] Amoruso A, Bardelli C, Fresu LG, Palma A, Vidali M, Ferrero V, et al. Enhanced peroxisome proliferator-activated receptor-gamma expression in monocyte/macrophages from coronary artery disease patients and possible gender differences. *J Pharmacol Exp Ther.* 2009;331:531-8.
- [273] Pergola C, Rogge A, Dodt G, Northoff H, Weinigel C, Barz D, et al. Testosterone suppresses phospholipase D, causing sex differences in leukotriene biosynthesis in human monocytes. *Faseb J.* 2011;25:3377-87.
- [274] Schaible AM, Koeberle A, Northoff H, Lawrenz B, Weinigel C, Barz D, et al. High capacity for leukotriene biosynthesis in peripheral blood during pregnancy. *Prostaglandins Leukot Essent Fatty Acids.* 2013;89:245-55.
- [275] Kramer PR, Winger V, Kramer SF. 17beta-Estradiol utilizes the estrogen receptor to regulate CD16 expression in monocytes. *Mol Cell Endocrinol.* 2007;279:16-25.

- [276] Konstantinidis D, Paletas K, Koliakos G, Kaloyianni M. Signaling components involved in leptin-induced amplification of the atherosclerosis-related properties of human monocytes. *J Vasc Res.* 2009;46:199-208.
- [277] Jia SJ, Niu PP, Cong JZ, Zhang BK, Zhao M. TLR4 signaling: a potential therapeutic target in ischemic coronary artery disease. *Int Immunopharmacol.* 2014;23:54-9.
- [278] Wege N, Schutkowski A, Boenn M, Bialek J, Schlitt A, Noack F, et al. Men and women differ in their diurnal expression of monocyte peroxisome proliferator-activated receptor-alpha in the fed but not in the fasted state. *Faseb J.* 2015;29:2905-11.
- [279] Trottier J, Caron P, Straka RJ, Barbier O. Profile of serum bile acids in noncholestatic volunteers: gender-related differences in response to fenofibrate. *Clin Pharmacol Ther.* 2011;90:279-86.
- [280] Ginsberg HN, Elam MB, Lovato LC, Crouse JR, 3rd, Leiter LA, Linz P, et al. Effects of combination lipid therapy in type 2 diabetes mellitus. *N Engl J Med.* 2010;362:1563-74.
- [281] Ruggieri A, Gambardella L, Maselli A, Vona R, Anticoli S, Panusa A, et al. Statin-induced impairment of monocyte migration is gender-related. *J Cell Physiol.* 2014;229:1990-8.
- [282] Krysiak R, Gdula-Dymek A, Marek B, Okopien B. Comparison of the effects of hypolipidemic treatment on monocyte proinflammatory cytokine release in men and women with type 2 diabetes and atherogenic dyslipidemia. *Endokrynol Pol.* 2015;66:224-30.
- [283] Campesi I, Marino M, Montella A, Pais S, Franconi F. Sex differences in estrogen receptor alpha and beta levels and activation status in LPS-stimulated human macrophages. *J Cell Physiol.* 2016.
- [284] Svenson J, Cunningham M, Dasgupta S, Gilkeson GS. Estrogen receptor alpha modulates mesangial cell responses to toll-like receptor ligands. *Am J Med Sci.* 2014;348:492-500.
- [285] Martinez FO, Gordon S, Locati M, Mantovani A. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *J Immunol.* 2006;177:7303-11.
- [286] Toniolo A, Fadini GP, Tedesco S, Cappellari R, Vegeto E, Maggi A, et al. Alternative activation of human macrophages is rescued by estrogen treatment in vitro and impaired by menopausal status. *J Clin Endocrinol Metab.* 2015;100:E50-8.
- [287] Drechsler M, Duchene J, Soehnlein O. Chemokines control mobilization, recruitment, and fate of monocytes in atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2015;35:1050-5.
- [288] Wiktor-Jedrzejczak W, Gordon S. Cytokine regulation of the macrophage (M phi) system studied using the colony stimulating factor-1-deficient op/op mouse. *Physiol Rev.* 1996;76:927-47.
- [289] Saitoh T, Fujita N, Jang MH, Uematsu S, Yang BG, Satoh T, et al. Loss of the autophagy protein Atg16L1 enhances endotoxin-induced IL-1beta production. *Nature.* 2008;456:264-8.
- [290] Harris J, Hartman M, Roche C, Zeng SG, O'Shea A, Sharp FA, et al. Autophagy controls IL-1beta secretion by targeting pro-IL-1beta for degradation. *J Biol Chem.* 2011;286:9587-97.
- [291] Razani B, Feng C, Coleman T, Emanuel R, Wen H, Hwang S, et al. Autophagy links inflammasomes to atherosclerotic progression. *Cell Metab.* 2012;15:534-44.
- [292] Singh R, Kaushik S, Wang Y, Xiang Y, Novak I, Komatsu M, et al. Autophagy regulates lipid metabolism. *Nature.* 2009;458:1131-5.
- [293] Ouimet M, Franklin V, Mak E, Liao X, Tabas I, Marcel YL. Autophagy regulates cholesterol efflux from macrophage foam cells via lysosomal acid lipase. *Cell Metab.* 2011;13:655-67.
- [294] Ouimet M, Marcel YL. Regulation of lipid droplet cholesterol efflux from macrophage foam cells. *Arterioscler Thromb Vasc Biol.* 2012;32:575-81.
- [295] Liao X, Sluimer JC, Wang Y, Subramanian M, Brown K, Pattison JS, et al. Macrophage autophagy plays a protective role in advanced atherosclerosis. *Cell Metab.* 2012;15:545-53.
- [296] Bonilla DL, Bhattacharya A, Sha Y, Xu Y, Xiang Q, Kan A, et al. Autophagy regulates phagocytosis by modulating the expression of scavenger receptors. *Immunity.* 2013;39:537-47.
- [297] Fahy RJ, Doseff AI, Wewers MD. Spontaneous human monocyte apoptosis utilizes a caspase-3-dependent pathway that is blocked by endotoxin and is independent of caspase-1. *J Immunol.* 1999;163:1755-62.
- [298] Savill J, Fadok V. Corpse clearance defines the meaning of cell death. *Nature.* 2000;407:784-8.

- [299] Cominacini L, Garbin U, Mozzini C, Stranieri C, Pasini A, Solani E, et al. The atherosclerotic plaque vulnerability: focus on the oxidative and endoplasmic reticulum stress in orchestrating the macrophage apoptosis in the formation of the necrotic core. *Curr Med Chem*. 2015;22:1565-72.
- [300] Linton MRF, Yancey PG, Davies SS, Vickers KC, Jerome WGJ, Linton EF. The role of lipids and lipoproteins in atherosclerosis. South Dartmouth (MA): Endotext; 2000.
- [301] Evans MJ, MacLaughlin S, Marvin RD, Abdou NI. Estrogen decreases in vitro apoptosis of peripheral blood mononuclear cells from women with normal menstrual cycles and decreases TNF-alpha production in SLE but not in normal cultures. *Clin Immunol Immunopathol*. 1997;82:258-62.
- [302] Sergin I, Razani B. Self-eating in the plaque: what macrophage autophagy reveals about atherosclerosis. *Trends Endocrinol Metab*. 2014;25:225-34.
- [303] Cheng J, Qiao L, Xu X, Zhai C, Zhao K, Ji X, et al. Lower AMP-activated protein kinase level is associated with the vulnerability of coronary atherosclerotic plaques by attenuating the expression of monocyte autophagy. *Coron Artery Dis*. 2015;26:322-7.
- [304] Lista P, Straface E, Brunelleschi S, Franconi F, Malorni W. On the role of autophagy in human diseases: a gender perspective. *J Cell Mol Med*. 2011;15:1443-57.
- [305] Gerrity RG, Naito HK, Richardson M, Schwartz CJ. Dietary induced atherogenesis in swine. Morphology of the intima in prelesion stages. *Am J Pathol*. 1979;95:775-92.
- [306] Ridker PM, Hennekens CH, Roitman-Johnson B, Stampfer MJ, Allen J. Plasma concentration of soluble intercellular adhesion molecule 1 and risks of future myocardial infarction in apparently healthy men. *Lancet*. 1998;351:88-92.
- [307] McCrohon JA, Nakhla S, Jessup W, Stanley KK, Celermajer DS. Estrogen and progesterone reduce lipid accumulation in human monocyte-derived macrophages: a sex-specific effect. *Circulation*. 1999;100:2319-25.
- [308] Ng MK, Quinn CM, McCrohon JA, Nakhla S, Jessup W, Handelsman DJ, et al. Androgens up-regulate atherosclerosis-related genes in macrophages from males but not females: molecular insights into gender differences in atherosclerosis. *J Am Coll Cardiol*. 2003;42:1306-13.
- [309] McCrohon JA, Jessup W, Handelsman DJ, Celermajer DS. Androgen exposure increases human monocyte adhesion to vascular endothelium and endothelial cell expression of vascular cell adhesion molecule-1. *Circulation*. 1999;99:2317-22.
- [310] Mukherjee TK, Dinh H, Chaudhuri G, Nathan L. Testosterone attenuates expression of vascular cell adhesion molecule-1 by conversion to estradiol by aromatase in endothelial cells: implications in atherosclerosis. *Proc Natl Acad Sci U S A*. 2002;99:4055-60.
- [311] Sorci-Thomas MG, Thomas MJ. Microdomains, inflammation, and atherosclerosis. *Circ Res*. 2016;118:679-91.
- [312] Cullen P, Cignarella A, Brennhansen B, Mohr S, Assmann G, von Eckardstein A. Phenotype-dependent differences in apolipoprotein E metabolism and in cholesterol homeostasis in human monocyte-derived macrophages. *J Clin Invest*. 1998;101:1670-7.
- [313] Wang X, Collins HL, Ranalletta M, Fuki IV, Billheimer JT, Rothblat GH, et al. Macrophage ABCA1 and ABCG1, but not SR-BI, promote macrophage reverse cholesterol transport in vivo. *J Clin Invest*. 2007;117:2216-24.
- [314] Out R, Hoekstra M, Habets K, Meurs I, de Waard V, Hildebrand RB, et al. Combined deletion of macrophage ABCA1 and ABCG1 leads to massive lipid accumulation in tissue macrophages and distinct atherosclerosis at relatively low plasma cholesterol levels. *Arterioscler Thromb Vasc Biol*. 2008;28:258-64.
- [315] Stahlberg N, Rico-Bautista E, Fisher RM, Wu X, Cheung L, Flores-Morales A, et al. Female-predominant expression of fatty acid translocase/CD36 in rat and human liver. *Endocrinology*. 2004;145:1972-9.
- [316] Brodeur MR, Luangrath V, Bourret G, Falstraull L, Brissette L. Physiological importance of SR-BI in the in vivo metabolism of human HDL and LDL in male and female mice. *J Lipid Res*. 2005;46:687-96.
- [317] Wang H, Liu Y, Zhu L, Wang W, Wan Z, Chen F, et al. 17beta-estradiol promotes cholesterol efflux from vascular smooth muscle cells through a liver X receptor alpha-dependent pathway. *Int J Mol Med*. 2014;33:550-8.

- [318] Segatto M, Trapani L, Marino M, Pallottini V. Age- and sex-related differences in extra-hepatic low-density lipoprotein receptor. *J Cell Physiol.* 2011;226:2610-6.
- [319] Lorbek G, Perse M, Horvat S, Bjorkhem I, Rozman D. Sex differences in the hepatic cholesterol sensing mechanisms in mice. *Molecules.* 2013;18:11067-85.
- [320] Rayner K, Chen YX, Siebert T, O'Brien ER. Heat shock protein 27: clue to understanding estrogen-mediated atheroprotection? *Trends Cardiovasc Med.* 2010;20:54-8.
- [321] Corcoran MP, Lichtenstein AH, Meydani M, Dillard A, Schaefer EJ, Lamon-Fava S. The effect of 17beta-estradiol on cholesterol content in human macrophages is influenced by the lipoprotein milieu. *J Mol Endocrinol.* 2011;47:109-17.
- [322] Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol.* 2015;294:63-9.
- [323] Lommatzsch M, Bratke K, Knappe T, Bier A, Dreschler K, Kuepper M, et al. Acute effects of tobacco smoke on human airway dendritic cells in vivo. *Eur Respir J.* 2010;35:1130-6.
- [324] Vassallo R, Tamada K, Lau JS, Kroening PR, Chen L. Cigarette smoke extract suppresses human dendritic cell function leading to preferential induction of Th-2 priming. *J Immunol.* 2005;175:2684-91.
- [325] Meier A, Chang JJ, Chan ES, Pollard RB, Sidhu HK, Kulkarni S, et al. Sex differences in the Toll-like receptor-mediated response of plasmacytoid dendritic cells to HIV-1. *Nat Med.* 2009;15:955-9.
- [326] Berghofer B, Frommer T, Haley G, Fink L, Bein G, Hackstein H. TLR7 ligands induce higher IFN-alpha production in females. *J Immunol.* 2006;177:2088-96.
- [327] Seillet C, Rouquie N, Foulon E, Douin-Echinard V, Krust A, Chambon P, et al. Estradiol promotes functional responses in inflammatory and steady-state dendritic cells through differential requirement for activation function-1 of estrogen receptor alpha. *J Immunol.* 2013;190:5459-70.
- [328] Laffont S, Rouquie N, Azar P, Seillet C, Plumas J, Asford C, et al. X-Chromosome complement and estrogen receptor signaling independently contribute to the enhanced TLR7-mediated IFN-alpha production of plasmacytoid dendritic cells from women. *J Immunol.* 2014;193:5444-52.
- [329] Griesbeck M, Ziegler S, Laffont S, Smith N, Chauveau L, Tomezsko P, et al. Sex differences in plasmacytoid dendritic cell levels of IRF5 drive higher IFN-alpha production in women. *J Immunol.* 2015;195:5327-36.
- [330] Segerer SE, Muller N, van den Brandt J, Kapp M, Dietl J, Reichardt HM, et al. Impact of female sex hormones on the maturation and function of human dendritic cells. *Am J Reprod Immunol.* 2009;62:165-73.
- [331] Xie H, Hua C, Sun L, Zhao X, Fan H, Dou H, et al. 17beta-estradiol induces CD40 expression in dendritic cells via MAPK signaling pathways in a minichromosome maintenance protein 6-dependent manner. *Arthritis Rheum.* 2011;63:2425-35.
- [332] Wang JP, Zhang L, Madera RF, Woda M, Libraty DH. Plasmacytoid dendritic cell interferon-alpha production to R-848 stimulation is decreased in male infants. *BMC Immunol.* 2012;13:35.
- [333] Hotchkiss RS, Karl IE. The pathophysiology and treatment of sepsis. *N Engl J Med.* 2003;348:138-50.
- [334] Kyurkchiev D, Ivanova-Todorova E, Hayrabedian S, Altankova I, Kyurkchiev S. Female sex steroid hormones modify some regulatory properties of monocyte-derived dendritic cells. *Am J Reprod Immunol.* 2007;58:425-33.
- [335] Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol.* 2011;12:204-12.
- [336] Blesson CS. Estrogen Receptors in Leukocytes - Possible Impact on Inflammatory Processes in the Female Reproductive System, Update on Mechanisms of Hormone Action In: Aimaretti G, editor. *Focus on Metabolism, Growth and Reproduction.* Rijeka, Croatia InTech; 2011.
- [337] Pierdominici M, Maselli A, Colasanti T, Giammarioli AM, Delunardo F, Vacirca D, et al. Estrogen receptor profiles in human peripheral blood lymphocytes. *Immunol Lett.* 2010;132:79-85.
- [338] Fan H, Dong G, Zhao G, Liu F, Yao G, Zhu Y, et al. Gender differences of B cell signature in healthy subjects underlie disparities in incidence and course of SLE related to estrogen. *J Immunol Res.* 2014;2014:814598.
- [339] Hewagama A, Patel D, Yarlagadda S, Strickland FM, Richardson BC. Stronger inflammatory/cytotoxic T-cell response in women identified by microarray analysis. *Genes Immun.* 2009;10:509-16.

- [340] Bobryshev YV, Watanabe T. Ultrastructural evidence for association of vascular dendritic cells with T-lymphocytes and with B-cells in human atherosclerosis. *J Submicrosc Cytol Pathol.* 1997;29:209-21.
- [341] Abdullah M, Chai PS, Chong MY, Tohit ER, Ramasamy R, Pei CP, et al. Gender effect on in vitro lymphocyte subset levels of healthy individuals. *Cell Immunol.* 2012;272:214-9.
- [342] Garcia-Dabrio MC, Pujol-Moix N, Martinez-Perez A, Fontcuberta J, Souto JC, Soria JM, et al. Influence of age, gender and lifestyle in lymphocyte subsets: report from the Spanish Gait-2 Study. *Acta Haematol.* 2012;127:244-9.
- [343] Strindhall J, Skog M, Ernerudh J, Bengner M, Lofgren S, Matussek A, et al. The inverted CD4/CD8 ratio and associated parameters in 66-year-old individuals: the Swedish HEXA immune study. *Age (Dordr).* 2013;35:985-91.
- [344] Pietschmann P, Gollob E, Brosch S, Hahn P, Kudlacek S, Willheim M, et al. The effect of age and gender on cytokine production by human peripheral blood mononuclear cells and markers of bone metabolism. *Exp Gerontol.* 2003;38:1119-27.
- [345] Yan J, Greer JM, Hull R, O'Sullivan JD, Henderson RD, Read SJ, et al. The effect of ageing on human lymphocyte subsets: comparison of males and females. *Immun Ageing.* 2010;7:4-13.
- [346] Pehlivanoglu B, Bayrak S, Gurel EI, Balkanci ZD. Effect of gender and menstrual cycle on immune system response to acute mental stress: apoptosis as a mediator. *Neuroimmunomodulation.* 2012;19:25-32.
- [347] Franconi F, Rosano G, Campesi I. Need for gender-specific pre-analytical testing: the dark side of the moon in laboratory testing. *Int J Cardiol.* 2015;179:514-35.
- [348] Goetzl EJ, Huang MC, Kon J, Patel K, Schwartz JB, Fast K, et al. Gender specificity of altered human immune cytokine profiles in aging. *Faseb J.* 2010;24:3580-9.
- [349] Iadecola C, Anrather J. The immunology of stroke: from mechanisms to translation. *Nat Med.* 2011;17:796-808.
- [350] Bartlett DB, Firth CM, Phillips AC, Moss P, Baylis D, Syddall H, et al. The age-related increase in low-grade systemic inflammation (Inflammaging) is not driven by cytomegalovirus infection. *Aging Cell.* 2012;11:912-5.
- [351] Conway SE, Roy-O'Reilly M, Friedler B, Staff I, Fortunato G, McCullough LD. Sex differences and the role of IL-10 in ischemic stroke recovery. *Biol Sex Differ.* 2015;6:17.
- [352] Cuervo AM, Macian F. Autophagy and the immune function in aging. *Curr Opin Immunol.* 2014;29:97-104.
- [353] Grimaldi CM, Cleary J, Dagtas AS, Moussai D, Diamond B. Estrogen alters thresholds for B cell apoptosis and activation. *J Clin Invest.* 2002;109:1625-33.
- [354] Mandal A, Viswanathan C. Natural killer cells: In health and disease. *Hematol Oncol Stem Cell Ther.* 2015;8:47-55.
- [355] Bonaccorsi I, De Pasquale C, Campana S, Barberi C, Cavaliere R, Benedetto F, et al. Natural killer cells in the innate immunity network of atherosclerosis. *Immunol Lett.* 2015;168:51-7.
- [356] Kee SJ, Park YW, Cho YN, Jin HM, Kim MJ, Lee SJ, et al. Age- and gender-related differences in circulating natural killer T cells and their subset levels in healthy Korean adults. *Hum Immunol.* 2012;73:1011-6.
- [357] Bernin H, Fehling H, Marggraff C, Tannich E, Lotter H. The cytokine profile of human NKT cells and PBMCs is dependent on donor sex and stimulus. *Med Microbiol Immunol.* 2016.
- [358] Snyder-Cappione JE, Tincati C, Eccles-James IG, Cappione AJ, Ndhlovu LC, Koth LL, et al. A comprehensive ex vivo functional analysis of human NKT cells reveals production of MIP1-alpha and MIP1-beta, a lack of IL-17, and a Th1-bias in males. *PLoS One.* 2010;5:e15412.
- [359] Al-Attar A, Presnell SR, Peterson CA, Thomas DT, Lutz CT. The effect of sex on immune cells in healthy aging: Elderly women have more robust natural killer lymphocytes than do elderly men. *Mech Ageing Dev.* 2016;156:25-33.
- [360] Hirokawa K, Utsuyama M, Hayashi Y, Kitagawa M, Makinodan T, Fulop T. Slower immune system aging in women versus men in the Japanese population. *Immun Ageing.* 2013;10:19.
- [361] Holt PG. Immune and inflammatory function in cigarette smokers. *Thorax.* 1987;42:241-9.

- [362] Tollerud DJ, Clark JW, Brown LM, Neuland CY, Mann DL, Pankiw-Trost LK, et al. Association of cigarette smoking with decreased numbers of circulating natural killer cells. *Am Rev Respir Dis.* 1989;139:194-8.
- [363] Inoue C, Takeshita T, Kondo H, Morimoto K. Cigarette smoking is associated with the reduction of lymphokine-activated killer cell and natural killer cell activities. *Environ Health Prev Med.* 1996;1:14-9.
- [364] Mian MF, Lauzon NM, Stampfli MR, Mossman KL, Ashkar AA. Impairment of human NK cell cytotoxic activity and cytokine release by cigarette smoke. *J Leukoc Biol.* 2008;83:774-84.
- [365] Yovel G, Shakhar K, Ben-Eliyahu S. The effects of sex, menstrual cycle, and oral contraceptives on the number and activity of natural killer cells. *Gynecol Oncol.* 2001;81:254-62.
- [366] Gibson DA, Greaves E, Critchley HO, Saunders PT. Estrogen-dependent regulation of human uterine natural killer cells promotes vascular remodelling via secretion of CCL2. *Hum Reprod.* 2015;30:1290-301.
- [367] Mandler RN, Seamer LC, Domalewski MD, Bankhurst AD. Progesterone but not estrogen depolarizes natural killer cells. *Nat Immun.* 1993;12:128-35.
- [368] Sulke AN, Jones DB, Wood PJ. Hormonal modulation of human natural killer cell activity in vitro. *J Reprod Immunol.* 1985;7:105-10.
- [369] Baltz KM, Krusch M, Baessler T, Schmiedel BJ, Bringmann A, Brossart P, et al. Neutralization of tumor-derived soluble glucocorticoid-induced TNFR-related protein ligand increases NK cell anti-tumor reactivity. *Blood.* 2008;112:3735-43.
- [370] Shi GP, Bot I, Kovanen PT. Mast cells in human and experimental cardiometabolic diseases. *Nat Rev Cardiol.* 2015;12:643-58.
- [371] Chen W, Beck I, Schober W, Brockow K, Effner R, Buters JT, et al. Human mast cells express androgen receptors but treatment with testosterone exerts no influence on IgE-independent mast cell degranulation elicited by neuromuscular blocking agents. *Exp Dermatol.* 2010;19:302-4.
- [372] Bakry OA, Samaka RM, Shoeib MA, Maher A. Immunolocalization of androgen receptor and estrogen receptors in skin tags. *Ultrastruct Pathol.* 2014;38:344-57.
- [373] Zierau O, Zenclussen AC, Jensen F. Role of female sex hormones, estradiol and progesterone, in mast cell behavior. *Front Immunol.* 2012;3:169.
- [374] Levick SP, Melendez GC, Plante E, McLarty JL, Brower GL, Janicki JS. Cardiac mast cells: the centrepiece in adverse myocardial remodelling. *Cardiovasc Res.* 2011;89:12-9.
- [375] Ferraz ML, Nascimento DM, Rorato JP, Espindula AP, Oliveira LF, Ramalho LS, et al. Correlation of lifetime progress of atherosclerosis and morphologic markers of severity in humans: new tools for a more sensitive evaluation. *Clinics (Sao Paulo).* 2012;67:1071-5.
- [376] Page C, Pitchford S. Neutrophil and platelet complexes and their relevance to neutrophil recruitment and activation. *Int Immunopharmacol.* 2013;17:1176-84.

Table 1. Protein expression of sex hormone receptors in human cells

Cell types	ERα	ERβ	AR	PR
ECs	M = F [11]	M = F [11]	M = F [11, 39] M > F [44, 45]	
hVSMCs	M = F [97] M < F (starved HAEC)[98]	M > F [97]	M = F [97]	Present; no gender analysis [92, 93]
PLTs	M = F [122]	M = F [122]	Present (no sex analysis) [121]	
EPCs	M = F [17, 18] = during ovulatory than follicular and luteal phases, and vs men and postmenopausal women [176]	M = F [17] > during ovulatory than follicular and luteal phases, and vs men and postmenopausal women [176]		
Neutrophils	Present (no sex	Present (no sex	M = F [219]	ND [27]

	analysis) [218]	analysis) [218]		
Monocytes	M = F [258]	M = F [258]	M > F [257]	ND [27]
Macrophages	M < F [283]	M < F [283]		
	M = F [258]	M = F [258]		
	F < F+OCs [68]	F > F+OCs [68]	F = F+OCs [68]	
Lymphocytes	Present (no sex analysis) [336]	Present (no sex analysis) [336]		Only membrane-bound PR [27]
NKs	Present (no sex analysis) [322]	Present (no sex analysis) [322]		ND [27]
DCs	Present (no sex analysis) [322, 327]	Present (no sex analysis); [322, 327]		ND [27]
Mast cells	Present (no sex analysis) [372, 373]	Present (no sex analysis) [372, 373]	Present (no sex analysis) [371]	Present (no sex analysis) [373]

ND: not detected