

Nasal chitosan microparticles target a zidovudine prodrug to brain HIV sanctuaries

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1 **Nasal chitosan microparticles target a zidovudine prodrug to brain HIV sanctuaries**

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1 **ABSTRACT**

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4 3 Zidovudine (AZT) is an antiretroviral drug that is a substrate of active efflux transporters
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7 4 (AETs) that extrude the drug from the central nervous system (CNS) and macrophages, which
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10 5 are considered to be sanctuaries of HIV. The conjugation of AZT to ursodeoxycholic acid is
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12 6 known to produce a prodrug (UDCA-AZT) that is able to elude the AET systems, indicating
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14 7 the potential ability of this prodrug to act as a carrier of AZT in the CNS and in macrophages.
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17 8 Here, we demonstrate that UDCA-AZT is able to permeate and remain in murine
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19 9 macrophages with an efficiency twenty times higher than that of AZT. Moreover, we propose
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21 10 the nasal administration of this prodrug in order to induce its uptake into the CNS. Chitosan
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23 11 chloride-based microparticles (CP) were prepared by spray-drying and were characterized
24
25 12 with respect to size, morphology, density, water uptake and the dissolution profile of UDCA-
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27 13 AZT. The CP sample was then nasally administered to rats. *All in vitro* and *in vivo*
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29 14 measurements were also performed for a CP parent physical mixture. The CP sample was able
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31 15 to increase the dissolution rate of UDCA-AZT and to reduce water uptake with respect to its
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33 16 parent physical mixture, inducing better uptake of UDCA-AZT into the cerebrospinal fluid of
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35 17 rats, where the prodrug can act as an AZT carrier in macrophages.
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46 20 *Keywords:* Zidovudine prodrug, HIV treatment, macrophage, brain targeting, nasal
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48 21 formulation, chitosan microparticle
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53 23 *Chemical compounds studied in this article:*

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56 24 Zidovudine (PubChem CID: 35370); Chitosan (PubChem CID: 21896651)
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1. Introduction

Combination antiretroviral therapy (cART) has made a significant impact against HIV infection. However, the efficacy of this therapy is limited by the poor bioavailability of anti-HIV drugs at viral reservoir sites, such as the central nervous system (CNS) (Cunningham et al., 2000; Kolson and Gonzalez-Scarano, 2000), particularly in cerebrospinal fluid (CSF) subarachnoid spaces that contain macrophages, which constitute the only site of HIV replication in the brain (Cunningham et al., 1997; Gherzi-Egea et al., 1996). The lack of drug penetration into these “HIV sanctuaries” occurs primarily due to the expression of active efflux transporters (AETs) on the blood–brain (BBB) and blood–cerebrospinal fluid (BCSFB) barriers (Namanja et al., 2012; Pavan and Dalpiaz, 2011). *In vivo*, these transporters induce the asymmetric transport of anti-HIV drugs across these barriers, where the rate of drug efflux from the CNS to the blood is greater than the influx rate (Wang and Sawchuk, 1995).

We recently proved that the conjugation of zidovudine (AZT), which is an antiretroviral drug used in cART protocols (De Clercq, 2009), with the bile acid ursodeoxycholic acid (UDCA) generates a prodrug (UDCA–AZT) that can elude the AET systems (Dalpiaz et al., 2012). We have therefore proposed that when this prodrug is taken up in the CNS, it should not be extruded into the bloodstream because it is able to elude the AET systems. Accordingly, we demonstrated that the nasal administration of solid lipid microparticles loaded with the UDCA-AZT prodrug facilitates its uptake into the CSF of rats (Dalpiaz et al., 2014). Nasal administration constitutes a potentially efficacious way of achieving the brain uptake of neuroactive agents (Fine et al, 2014, 2015; Illum, 2000; Vyas et al., 2005). Indeed, drugs deposited on the olfactory epithelium of the nose can gain direct access to the CNS, particularly the CSF, via transcellular transport through olfactory epithelial cells. The drugs absorbed in the CSF can then diffuse into the interstitial fluid (ISF) and subsequently penetrate the brain parenchyma (Illum, 2000, 2004; Thorne and Frey, 2001). Moreover, drugs

1 deposited on the olfactory epithelium can be transported into the brain parenchyma by
2 olfactory neurons or trigeminal nerves that reach the nasal cavity (Finger et al., 1990; Illum,
3 2000; Johnson et al., 2010). Finally, nasally administered drugs can be absorbed into the
4 systemic circulation from the respiratory epithelium (Cho et al, 2014) and can then reach the
5 CNS if they are able to cross the BBB (Illum, 2000).

6 In general, appropriate strategies are required to improve the brain delivery of drugs,
7 such as the addition of penetration enhancers and mucoadhesive materials to formulations or
8 the preparation of micro- and nano-particulate delivery systems (Casettari and Illum, 2014;
9 Dalpiaz et al., 2008; Horvát et al., 2009; Mistry et al., 2009; Rassa et al., 2015a). In this
10 regard, chitosan is a polysaccharide derived from the alkaline deacetylation of chitin that is
11 used in different formulations for the nose-to-brain delivery of drugs (Casettari and Illum,
12 2014) due to its biocompatibility, nontoxicity and high charge density, which confers
13 mucoadhesive properties (Bernkop-Schnürch and Dünnhaupt, 2012; Sinha et al, 2004).
14 Chitosan is poorly soluble in water at physiologic pH values, but it forms salts with inorganic
15 or organic acids, such as hydrochloride and glutamic acid, that are soluble in water and
16 possess better characteristics than chitosan itself, such as mucoadhesiveness and the ability to
17 enhance the penetration of neuroactive agents into the CNS (Dalpiaz et al., 2008; Gavini et
18 al., 2011; Maestrelli et al., 2004). Chitosan is also characterized by the ability to reversibly
19 open tight junctions, which is associated with a potential increase in the transcellular transport
20 of drugs across the olfactory mucosa (Durhia et al, 2010). Nasal formulations obtained in the
21 presence of water have drawbacks, such as the risk of chemical and physical instability and
22 microbiological growth. In addition, the residence time of the liquid formulation in the nasal
23 cavity is short, as the liquid is often rinsed into the gastrointestinal tract (GIT) or out of the
24 nose (Kublik and Vidgren, 1998). These disadvantages can be overcome by using powder-
25 based formulations (Marttin et al., 1997; Rassa et al., 2015b).

26 The purpose of the present work was first to demonstrate that UDCA-AZT can

1 permeate and remain in macrophages more efficiently than its parent drug AZT and then to
2 prepare microspheres based on chitosan chloride in order to increase the nose-to-brain
3 delivery of UDCA-AZT by using a particulate formulation that is suitable for nasal
4 administration. The characterization of microparticles was performed *in vitro* and *in vivo*. All
5 *in vitro* and *in vivo* measurements performed for the loaded microparticles were also
6 performed for a parent physical mixture of chitosan chloride and UDCA-AZT in order to
7 verify the efficacy of the microparticulate system for the brain delivery of the prodrug.

10 **2. Materials and methods**

12 *2.1. Materials*

14 The prodrug UDCA–AZT was synthesized as described previously (Dalpiaz et al.,
15 2012). Chitosan chloride (Protasan UP CL 113, molecular weight and degree of deacetylation
16 of 160,000 g/mol and 83%, respectively) was purchased from FMC BioPolymer AS
17 (Drammen, Norway). Dow Corning 345, which is a blend of polydimethylcyclosiloxane, was
18 obtained from Dow Corning (Brussels, Belgium). AZT, 7-n-propylxanthine (7n-PX), and
19 bovine serum albumin (BSA) were obtained from Sigma Aldrich Italy (Milan, Italy).
20 Methanol, acetonitrile, ethyl acetate and water were high performance liquid chromatography
21 (HPLC) grade and obtained from Sigma Aldrich Italy. Monopowder P[®] insufflators were
22 furnished by Valois Dispray (Mezzovico, Switzerland). Male Wistar rats were purchased from
23 Harlan SRC (Milan, Italy). All other reagents and solvents were of analytical grade (Sigma-
24 Aldrich).

26 *2.2. Uptake of AZT and UDCA-AZT in macrophages*

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2 The J774A.1 murine macrophage cell line was obtained from the American Tissue Type
3 Culture Collection (LGC Standards, Milan, Italy). J774A.1 macrophages were grown as
4 adherently cultured cells in Dulbecco's modified Eagle's medium + Glutamax supplemented
5 with 10% fetal bovine serum (FBS), 100 U/mL penicillin, and 100 µg/mL streptomycin at 37
6 °C. All cell culture reagents were provided by Invitrogen (Life Technologies, Milan, Italy).
7 The employed method is reported in detail in the Supplementary material. Briefly, a total of
8 5×10^5 cells J774A.1 cells were seeded in 12-well culture plates, and when semi-confluent, the
9 cells were exposed to 100 µM AZT and 10 or 100 µM UDCA-AZT in growth medium for 30
10 min. At the end of the treatment, the cells were washed and lysed. The cell lysates were dried
11 under a nitrogen stream, resuspended in methanol and centrifuged to remove cell debris. The
12 supernatant (10 µl) was used to measure the levels of AZT and UDCA-AZT via HPLC
13 analysis. All of the values obtained from experiments with J774A.1 cells are the mean of three
14 independent experiments.

16 2.3. Toxicity texts (MTT assay)

18 J774A.1 cells were seeded in 96-well plates at a density of 8,000 cells per well and
19 reached an optimal population density within 48-72 h. The cells were then incubated for 15,
20 30 and 60 min in 200 µL of culture medium in the presence and absence of 100 µM UDCA-
21 AZT. At the end of the time-course, the incubation medium was removed and replaced with
22 200 µL of fresh culture medium, and 20 µL of 3-(4,5-dimethylthiazol-2-yl)-2,5-
23 diphenyltetrazolium bromide (MTT) stock solution (5 mg/mL) were added to each well and
24 incubated at 37 °C for 4 h. A negative control of 20 µL of the MTT stock solution added to
25 200 µL of medium alone was included. Then, 100 µL of DMSO were added to each well and
26 incubated at 37 °C for 2 h in an orbital shaker incubator. Finally, the absorbance of each well

1 was measured at 570 nm using a microtiter plate reader.

2 3 4 5 2.4. *Preparation of loaded and unloaded chitosan microspheres*

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10 Chitosan microspheres containing UDCA–AZT (named CP) were prepared using the
11 spray-drying method. Chitosan chloride (400 mg) was dissolved in water (15 mL), whereas
12 UDCA–AZT (100 mg) was dissolved in methanol (35 ml). The drug solution was dispersed
13 into chitosan one (solid concentration: 1% w/v of UDCA–AZT and chitosan). Due to the very
14 low solubility of the drug, a fine suspension was obtained and spray-dried using a mini spray-
15 dryer equipped with a high performance cyclone (Büchi B-191, Büchi Labortechnik AG,
16 Flawil, Switzerland) and with a 0.7 mm two-fluid nozzle. The following standard operating
17 conditions were utilized: inlet and outlet temperature, 100 °C and 73 °C, respectively; spray
18 flow rate, 500 L/h; pump setting, 8% (2.00 mL/min); and aspirator setting, 90%. Aqueous
19 solutions of chitosan chloride (0.8 % w/v) were sprayed under the same conditions to obtain
20 drug-free microspheres (CH). The microparticles were stored in a desiccator at room
21 temperature.
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41 2.5. *HPLC analysis*

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46 The prodrug UDCA–AZT and its hydrolysis product AZT were quantified using HPLC.
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48 The chromatographic apparatus consisted of a modular system (model LC-10 AD VD pump
49 and model SPD-10A VP variable wavelength UV–vis detector; Shimadzu, Kyoto, Japan) and
50 an injection valve with a 20 µL sample loop (model 7725; Rheodyne, IDEX, Torrance, CA,
51 USA). Separations were performed at room temperature on a 5 µm Hypersil BDS C-18
52 column (150 mm × 4.6 mm i.d.; Alltech Italia Srl, Milan, Italy) equipped with a guard
53 column. Data acquisition and processing were accomplished with a personal computer using
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1 CLASS-VP Software, version 7.2.1 (Shimadzu Italia, Milan, Italy). The detector was set to
2 260 nm. The mobile phase consisted of a mixture of water and methanol that was regulated by
3 a gradient profile programmed as follows: isocratic elution with 20% (v/v) MeOH in H₂O for
4 10 min; a 1-min linear gradient to 75% (v/v) MeOH in H₂O; and the mobile phase
5 composition was then maintained at 75% MeOH for 10 min. After each cycle, the column was
6 conditioned with 20% (v/v) MeOH in H₂O for 10 min. The flow rate was 1 mL/min. The
7 xanthine derivative 7n-PX was employed as an internal standard for the analysis of rat blood
8 (see below). The retention times for 7n-PX, AZT and the prodrug UDCA-AZT were 6.5, 8.4,
9 and 19.6 min, respectively. The HPLC assay of UDCA-AZT alone was performed
10 isocratically with 80% (v/v) MeOH in H₂O. In this case, the retention time of UDCA-AZT
11 was 4.8 min. The chromatographic precision was evaluated (see Supplementary material).

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13 *2.6. Characterization of microspheres*

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15 The prepared microspheres were characterized in terms of production yield, drug
16 content and encapsulation efficiency, particle size and particle size distribution, particle
17 morphology, true density and water uptake capacity.

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19 *2.6.1. Production yields*

20 The production yields (YP) were calculated as a percent weight of microspheres
21 obtained with respect to the initial amounts of UDCA-AZT and chitosan that were employed
22 for the preparation of the feed suspension.

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24 *2.6.2. UDCA-AZT content in CP*

25 The UDCA-AZT content in the microparticulate powders was determined. The
26 microparticles (approximately 0.95 mg) were accurately weighed and dissolved in 3 ml of

1 water to which 300 μL of 0.2% H_3PO_4 was added; the final volume of the solution was
2 adjusted to 10 mL with methanol. Then, 10 μL of the filtered solutions (0.45 μm) were
3 injected into the HPLC system for the UDCA-AZT assay. The drug loading and entrapment
4 efficiency were calculated according to equations 1 and 2 (Rassu et al., 2014; see
5 Supplementary material).

6 All of the obtained values are the mean of four independent experiments.

7 2.6.3. Particle size measurement

8 The size and size distribution of the microspheres were analyzed via the light diffraction
9 method using a Coulter LS 100Q (Beckman Coulter Particle Characterization, Miami, FL,
10 USA). A sample (2 mg) of unloaded and drug-loaded microspheres was suspended in silicon
11 oil, sonicated for approximately 3 s and analyzed. The reported results are the averages of
12 triplicate averages. For comparison, a test was also performed on the physical mixture
13 containing UDCA–AZT and chitosan chloride (87:13 w/w) (MIX) that was prepared by
14 geometric dilution using an agate mortar pestle, as well as on chitosan chloride and UDCA-
15 AZT alone, before and after grinding.

16 2.6.4. Particle morphology

17 The shape and surface characteristics of the powders were studied using scanning
18 electron microscopy (VP-SEM; Zeiss EVO40XVP, Arese, Milan, Italy). The samples were
19 placed on double-sided tape that had previously been secured to aluminum stubs and then
20 analyzed under an argon atmosphere at an 18 kV acceleration voltage after gold sputtering.

21 2.6.5. True Density

22 The true density of the microspheres was measured via helium pycnometry
23 (Micromeritics Accupyc II 1340 Analysis system, Peschiera Borromeo, Italy) at 21 $^{\circ}\text{C}$

1 (Gavini et al., 2012). The density (ρ) of the powder was determined in triplicate for each
2 batch.

3 4 2.6.6. *Determination of water uptake capacity*

5 Investigations of the water uptake capability of CP and CH were carried out using a
6 modified Enslin apparatus, as described previously by Rassu et al., 2009. Briefly, the dried
7 microspheres were spread uniformly on the filter paper, which was previously soaked with
8 phosphate buffer, pH 6.5. The volume of fluid absorbed from the microspheres during the
9 time of the experiment (0 to 60 min) was recorded. The values (mean of at least 3
10 experiments) were expressed as μl of fluid absorbed per gram of microspheres during the time
11 of the experiment. For comparison, the test was also performed using MIX.

12 13 2.7. *In vitro release and dissolution studies*

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15 The release tests were performed using the flow-through dissolution method (European
16 Pharmacopoeia, Apparatus 4), with a modified glass cell that is useful for organic solvents
17 (described in the Supplementary material). The CP formulation (20 mg) was placed into the
18 glass cell. The dissolution medium was warmed to 37 ± 0.1 °C and introduced through the
19 bottom of the cell to obtain a suitable continuous flow. A methanol-water blend (70-30 v/v)
20 was used as the dissolution medium. The test solution (10 μL) was analyzed via HPLC to
21 evaluate the amount of UDCA-AZT released from the microspheres, which was calculated as
22 a percentage of the initial amount of UDCA-AZT incorporated into the microspheres prior to
23 the dissolution test. For comparison, the test was also performed using MIX. All experiments
24 were performed with at least three replicates.

25 26 2.8. *In vivo UDCA-AZT administration and quantification*

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2 Male Wistar rats (200–250 g) were anesthetized during the experimental period and
3 received a femoral intravenous infusion of 0.1 mg/mL of UDCA-AZT dissolved in a medium
4 constituted by 20% (v/v) DMSO and 80% (v/v) physiologic solution, at a rate of 0.2 mL/min
5 for 10 min. At the end of the infusion and at fixed time points, blood samples (100 μ L) were
6 collected and CSF samples (50 μ L) were withdrawn using the cisternal puncture method
7 described by van den Berg et al. (2002), which requires a single needle stick and allows the
8 collection of serial (40–50 μ L) CSF samples that are virtually blood-free (Dalpiaz et al.,
9 2014). A total volume of approximately 150 μ L of CSF was collected during the experimental
10 session. Four rats were employed for femoral intravenous infusions. The CSF samples (10
11 μ L) were immediately injected into the HPLC system for AZT and UDCA-AZT detection.
12 The blood samples were hemolysed immediately after their collection with 500 μ L of ice cold
13 water, and 50 μ L of 10% sulfosalicylic acid and 100 μ L of internal standard (30 μ M 7n-PX)
14 were then added. The samples were extracted twice with 1 mL of water-saturated ethyl
15 acetate, and after centrifugation, the organic layer was reduced to dryness under a nitrogen
16 stream. Two hundred microliters of a water-methanol mixture (70:30 v/v) were added, and
17 after centrifugation, 10 μ L were injected into the HPLC system for AZT and UDCA-AZT
18 detection.

19 The precision of the analytical method was determined (see Supplementary material).
20 The *in vivo* half-life of AZT in the blood was calculated by nonlinear regression (exponential
21 decay) of concentration values in the time range within 3 h after infusion and confirmed by
22 linear regression of the log concentration values versus time.

23 Nasal administration of UDCA-AZT was performed on anesthetized rats laid on their
24 backs, following two main procedures. The first procedure consisted of the introduction of 50
25 μ L of an aqueous suspension of UDCA-AZT (2 mg/mL) into each nostril, as described

1 previously (Dalpiaz et al., 2014). After administration, blood (100 μ L) and CSF samples (50
2 μ L) were collected at fixed time points and analyzed using the same procedures described
3 above. Four rats were employed for nasal administration of the UDCA-AZT suspension.

4 The second procedure was based on the insufflation of CP microparticles into each
5 nostril of anaesthetized rats using single dose Monopowder P[®] insufflators (Valois Dispray
6 SA, Mezzovico, Switzerland). These devices were constituted by a pump, a nasal adapter and
7 a solid formulation reservoir (Sacchetti et al., 2002). The insufflators were loaded with
8 approximately 0.8 mg of UDCA-AZT-loaded microparticles (corresponding to approximately
9 100 μ g of UDCA-AZT), or with approximately 0.8 mg of the corresponding physical mixture
10 (MIX). The rats received this amount in each nostril. The amount of powder emitted during
11 administration was determined based on the difference between the insufflator weights before
12 and after each insufflation. After administration, blood (100 μ L) and CSF samples (50 μ L)
13 were collected at fixed time points and analyzed using the same procedures described above.
14 Four rats were employed for nasal administration of the microparticulate powder.

15 All *in vivo* experiments were performed in accordance with the guidelines issued by the
16 Italian Ministry of Health (D.L. 116/92 and D.L. 111/94-B), the Declaration of Helsinki, and
17 the Guide for the Care and Use of Laboratory Animals as adopted and promulgated by the
18 National Institute of Health (Bethesda, Maryland, USA). The protocols for all *in vivo*
19 experiments were approved by the Local Ethics Committee (University of Ferrara, Ferrara,
20 Italy). All efforts were made to reduce the number of animals and their suffering.

21 The areas under the concentration curves of UDCA-AZT in the CSF (AUC, μ g mL⁻¹
22 min) were calculated using the trapezoidal method. The *in vivo* half-life of UDCA-AZT in the
23 CSF was calculated by linear regression of the log concentration values versus time between 1
24 and 3 h after nasal administration of the loaded microparticles. All calculations were
25 performed using the computer program Graph Pad Prism (GraphPad Software, Incorporated,
26 La Jolla, CA, USA).

2.9. Statistical analysis

Statistical comparisons of the amounts of water uptake of the powder samples and the UDCA-AZT C_{\max} and AUC values obtained in the CSF of rats were made using the Student's t test or one-way ANOVA (GraphPad Prism). P values <0.05 were considered to be statistically significant.

3. Results

3.1. Uptake of AZT and UDCA-AZT in macrophages

The incubation of 100 μM zidovudine or its prodrug with the macrophages for 30 min allowed us to detect 0.054 ± 0.007 nmoles of AZT and 1.12 ± 0.16 nmoles of UDCA-AZT in 10^6 cells, as reported in Fig. 1. The amount of AZT detected in the cells after incubation with UDCA-AZT for 30 min was 0.081 ± 0.009 nmoles/ 10^6 cells (Fig. 1). The cell counter measured the volume of macrophages as 1.7 ± 0.1 pL; as a consequence, the amount of AZT released by the macrophages after incubation with 100 μM UDCA-AZT incubation corresponds to a concentration of 47 ± 5 μM . No toxicity was observed for cells incubated with 100 μM UDCA-AZT. These data indicate not only that the prodrug is able to permeate and remain in the macrophages with an efficiency twenty times higher than that of its parent drug ($P < 0.001$) but also that the macrophages hydrolyze the prodrug, facilitating intracellular AZT release (Fig 1). Moreover, the incubation of macrophages with 10 μM UDCA-AZT allowed us to detect 0.21 ± 0.03 nmoles of the prodrug in 10^6 cells (Fig 1). In this case, the AZT amounts were lower than the limit of detection of the analytical system. Therefore, the

1 ability of UDCA-AZT to elude the AET systems (Dalpiaz et al., 2012) appears to be useful
2 for inducing uptake into macrophages. According to the data reported in Fig. 1, the presence
3 of the prodrug in the CSF should allow its permeation and permanence in the macrophages,
4 therefore making the prodrug an efficient carrier for AZT.

6 *3.2. Characterization of microspheres*

8 *3.2.1. Production yields*

9 The spray-drying technique proved to be suitable for preparing loaded and unloaded
10 chitosan microparticles; in fact, these microparticles were obtained with good production
11 yields of between 61.7% (CP) and 64.7% (CH). Generally, the yields obtained with the spray-
12 drying at laboratory scale with conventional spray-dryers are in the range of 20–70% due to
13 the loss of product on the walls of the drying chamber and the low capacity of the cyclone to
14 separate fine particles (Giunchedi et al., 2000; Sosnik and Seremeta, 2015). However, the
15 yield strongly depends on the work scale; the yield is high in larger scale setups because
16 large batch sizes and separators (e.g., filter systems) are used.

18 *3.2.2. UDCA–AZT content in CP*

19 The amount of encapsulated UDCA–AZT in the CP sample was found to be $13\pm 0.3\%$
20 (w/w), showing that chitosan, which is characterized by hydrophilic properties, was able to
21 encapsulate the lipophilic prodrug with a good encapsulation efficiency ($64.9\pm 1.5\%$).

23 *3.2.3. Particle size measurement*

24 Table 1 reports the volume-surface diameters (d_{vs}) obtained for the loaded (CP) and
25 unloaded microparticles (CH), for MIX and for chitosan chloride and UDCA-AZT before and
26 after grinding using an agate mortar and pestle. The loading of UDCA–AZT into chitosan

1 microparticles caused an increase in the particle size: CP microspheres had a d_{vs} of 3.59 ± 0.1
2 μm in comparison to a d_{vs} of $2.32\pm 0.01 \mu\text{m}$ for CH ($P<0.0001$). Furthermore, the presence of
3 the drug resulted in a change in the particle size distribution: loaded microparticles showed a
4 larger curve than drug-free microspheres (Fig. 2 up). In particular, CH showed a Gaussian
5 curve while the CP curve appeared to be leptokurtic, right skewed and wide, with the presence
6 of small shoulders.

7 MIX had a d_{vs} of $6.11\pm 0.022 \mu\text{m}$, which was significantly higher than the values
8 obtained for microspheres ($P<0.001$) and UDCA-AZT alone but significantly lower than the
9 value obtained for chitosan chloride alone, which were both analyzed after grinding. The
10 grinding of the drug and the polymer in an agate mortar reduced the particle size and shifted
11 the size distributions to lower values (Fig. 2, lower panel). In fact, the d_{vs} of chitosan chloride
12 changed from $19.98\pm 0.24 \mu\text{m}$ to $12.67\pm 0.819 \mu\text{m}$ due to aggregate breaking. The size
13 reduction of UDCA-AZT crystals was even more pronounced: the d_{vs} decreased from
14 $23.33\pm 0.06 \mu\text{m}$ to $5.12\pm 0.018 \mu\text{m}$ after grinding.

16 *3.2.4. Particle morphology*

17 Fig. 3A presents an SEM image of MIX obtained by geometric dilution using an agate
18 mortar and pestle. This picture reveals not only particles characterized by a round morphology
19 and a relatively smooth surface but also the presence of fragments characterized by an
20 irregular shape. For comparison, Figs. 3B and 3C present SEM images of chitosan chloride
21 and UDCA-AZT, respectively, which were ground using an agate mortar and pestle. Fig. 3B
22 shows particles characterized by a round morphology and a smooth surface. These
23 morphologic characteristics did not appear to be significantly different from those of chitosan
24 chloride before grinding, as evidenced by the SEM image presented in Fig. 3E. The SEM
25 picture of ground UDCA-AZT, Fig. 3C, showed fragments characterized by an irregular
26 shape (details are evident in the SEM image reported in Fig. 3D) and a high aptitude to

1 aggregate. These morphologic characteristics appeared to be significantly different from those
2 of the prodrug UDCA-AZT before grinding, as evidenced in Fig. 3F, where fragments of an
3 irregular shape are shown to be characterized by diameters at least one order of magnitude
4 higher than those of the ground UDCA-AZT particles. Taking into account these aspects, we
5 can conclude that the SEM image of the physical mixture of chitosan chloride and UDCA-
6 AZT (Fig. 3A) allows us to discriminate its two components; in particular, the particles with a
7 round morphology can be attributed to chitosan chloride, whereas the fragments of irregular
8 shape can be attributed to UDCA-AZT.

9 Figs. 4A and 4B present SEM images of CP, which is constituted by the loaded UDCA-
10 AZT microparticles based on chitosan chloride. The pictures reveal the presence of spherical
11 particles characterized by a smooth surface and several fragments characterized by an
12 irregular shape and high porosity, similar to spongy balls, with some degree of aggregation.
13 For comparison, a SEM image of the unloaded microparticles (CH) (Dalpiaz et al., 2008)
14 reveals their round shape and smooth surfaces. Taking into account these aspects, we can
15 hypothesize that the spherical particles presented in images 3A and 3B are constituted by
16 unloaded chitosan chloride, whereas the spongy balls were obtained from the interaction
17 between chitosan chloride and UDCA-AZT, which gave rise to the highly porous structures.

18 19 *3.2.5. True density*

20 The true density of the microspheres was $1.48 \pm 0.02 \text{ g/cm}^3$, which is comparable to the ρ
21 of nasal microspheres (Gavini et al., 2012).

22 23 *3.2.6. Determination of water uptake capacity*

24 Fig. 5 shows the water uptake of both loaded and unloaded formulations. The drug-free
25 formulations absorbed a greater amount of water than the loaded microparticles. Drug loading
26 decreased the water absorption capability of chitosan microspheres ($P < 0.05$). UDCA-AZT

1 and the chitosan chloride physical mixture also exhibited water uptake capabilities stronger
2 than that of CP but weaker than that of CH after 15 to 60 min ($P<0.05$).

3.3. *In vitro* release and dissolution studies

6 The dissolution and release studies of UDCA–AZT were performed in a MeOH/H₂O
7 (70:30, v/v) mixture. The employment of methanol as a cosolvent was necessary to increase
8 the low solubility of the prodrug in water (0.0030 ± 0.0001 mg/ml; Dalpiaz et al., 2012), thus
9 ensuring sink conditions.

10 Fig. 6 reports the release profile of UDCA–AZT from the loaded CP sample. The
11 release pattern is compared with that of UDCA-AZT dissolution from its physical mixture
12 with chitosan chloride. The dissolution rate of the prodrug included in MIX appeared to be
13 significantly lower than that of UDCA-AZT loaded in the CP sample. Indeed, after 2 h of
14 incubation, $38.4\pm 1.6\%$ of the prodrug loaded in the dissolution apparatus appeared to have
15 dissolved, whereas in the case of CP loaded microparticles, the amount of dissolved UDCA-
16 AZT was $86.2\pm 3.4\%$. As a consequence, the dissolution half-life of UDCA-AZT loaded in CP
17 sample appeared to be approximately 15 min, whereas for the prodrug in the physical mixture
18 (MIX), the dissolution half-life was greater than 2 h. These profiles suggest that the
19 employment of the CP sample as a nasal formulation should be appropriate, as this
20 formulation is able to release the prodrug UDCA-AZT in a relatively short time, allowing
21 potentially rapid absorption of the prodrug after the administration of the microparticles.

3.4. *In vivo* UDCA-AZT administration

25 Taking into account the fact that the CP microparticles were characterized not only by a
26 satisfactory encapsulation efficiency but also by the ability to induce relatively rapid

1 dissolution of UDCA–AZT, we tested these microparticles for nasal administration of the
2 prodrug in order to verify potential uptake in the CNS. The nasal administration of
3 UDCA–AZT was performed using three different formulations: (i) a suspension of the raw
4 UDCA-AZT powder in water, (ii) the powder constituted by loaded CP microparticles and
5 (iii) for comparison, the parent physical mixture of chitosan chloride and UDCA-AZT (87:13
6 w/w). The results were compared with those obtained after the intravenous infusion of
7 UDCA–AZT into rats.

9 *3.4.1. Intravenous administration of UDCA–AZT*

10 The prodrug was undetectable in rat blood samples following intravenous infusion. This
11 result is in agreement with the very fast hydrolysis of UDCA–AZT in rat blood at 37 °C
12 (Dalpiaz et al., 2012). Significant amounts of AZT were therefore detected following the
13 intravenous administration of the prodrug, as evidenced in Fig. 7. In particular, after the
14 infusion of 0.200 mg of UDCA–AZT into the rats, the AZT concentration in the bloodstream
15 was 4.50 ± 0.31 µg/mL. This concentration decreased over time with apparent first order
16 kinetics, as confirmed by the linearity of the semilogarithmic plot reported in the inset of Fig.
17 7 ($n = 9$, $r = 0.990$, $P < 0.0001$), and a half-life of 60.4 ± 3.8 min. These data are in good
18 agreement with those obtained from previous studies of *in vivo* UDCA-AZT
19 pharmacokinetics (Dalpiaz et al., 2014). No AZT or UDCA–AZT was detected in the CSF
20 within 180 min after the intravenous administration of UDCA–AZT.

22 *3.4.2. Nasal Administration of UDCA–AZT*

23 The nasal administration of pure UDCA-AZT as a water suspension did not allow us to obtain
24 detectable amounts of AZT or the prodrug in the blood or in the CSF, respectively, within 180
25 min of administration, as reported previously (Dalpiaz et al., 2014). In contrast, the nasal
26 administration of the powders constituted by MIX or by the CP microparticles (0.8 mg,

1 approximately 100 μg of UDCA–AZT in each nostril) produced detectable amounts of
2 UDCA–AZT in the CSF of the rats, as reported in Fig. 8. In particular, the UDCA–AZT C_{max}
3 was obtained after 60 min, with a value of $1.96 \pm 0.29 \mu\text{g/mL}$ ($3.1 \pm 0.4 \mu\text{M}$), after nasal
4 administration of the mixture.

5 The nasal administration of the same amount of CP microparticles increased the rate of
6 uptake of the prodrug into the CSF of rats. Indeed, the UDCA–AZT C_{max} was obtained at 60
7 min, with a value of $2.99 \pm 0.31 \mu\text{g/mL}$ ($4.7 \pm 0.5 \mu\text{M}$), which was significantly higher than the
8 C_{max} value obtained for the MIX sample ($P < 0.001$).

9 No AZT was detected in the bloodstream or in the CSF within 180 min after nasal
10 administration of the powders constituted by MIX or the CP microparticles.

11 The areas under the concentration (AUC) curve values obtained for UDCA–AZT in the
12 CSF after the nasal administration of MIX and CP microparticles (Fig. 8) were 201.39 ± 13.51
13 $\mu\text{g mL}^{-1} \text{ min}$ and $354.4 \pm 13.3 \mu\text{g mL}^{-1} \text{ min}$, respectively. The ratio between the AUC of the
14 CP sample and the AUC of the mixture was 1.76, indicating that the nasal formulation
15 constituted by the CP microparticles allowed an uptake of UDCA–AZT into the CSF that was
16 76% higher than that obtained after the administration of the parent mixture ($P < 0.001$).

17 The UDCA–AZT concentration in the CSF after nasal administration of CP
18 microparticles decreased over time with apparent first order kinetics, as confirmed by the
19 linearity of the semilogarithmic plot reported in the inset of Fig. 8 ($n = 4$, $r = 0.998$, $P < 0.05$),
20 and a half-life of $80.4 \pm 9.2 \text{ min}$.

23 4. Discussion

25 Although antiretroviral nucleoside derivatives are largely used against HIV infection
26 for their efficacy at the peripheral level, the total eradication of this virus from the body is

1 currently difficult to achieve. Indeed, antiretroviral drugs are unable to reach the CNS
2 (Kaufmann and Cooper, 2000; Strazielle, 2003); in contrast, the CNS is easily reached by
3 HIV through infected monocytes (Davis et al., 1992; Gray et al., 1996) that differentiate into
4 macrophages and microglia in the brain (Rausch and Stover, 2001). Therefore, the CNS and
5 those macrophages constitute sanctuaries for HIV, where drug resistance is induced and from
6 which the periphery can be re-infected (Cunningham et al., 2000; Kolson and Gonzalez-
7 Scarano, 2000). These phenomena are attributed primarily to the expression of AETs on the
8 membranes of macrophages (Chaudhary et al., 1992; Neyfakh et al., 1989) and blood brain
9 barrier (BBB) cells (Namanja et al., 2012; Pavan and Dalpiaz 2011), whose activity prevents
10 the penetration of antiretroviral drugs into HIV sanctuaries.

11 Among numerous nucleoside antivirals, AZT was the first to be approved and remains
12 the most widely used for the treatment and prophylaxis of HIV/AIDS (Hughes, 2015).
13 Unfortunately, long-term clinical use of AZT is associated with significant side effects, but
14 despite these drawbacks, AZT is considered to be one of the more potent inhibitors of the HIV
15 DNA polymerase (reverse transcriptase) (Arts and Hazuda, 2012; Golan et al., 2011).
16 Currently, tenofovir (TDF) is known to be a more potent and less toxic antiretroviral (ARV)
17 drug than AZT; however, a higher virological failure rate was observed after a clinical TDF-
18 containing ARV regimen in comparison to AZT-containing ARV therapy (Tang et al., 2012).

19 AZT activity in the CNS appears to be necessary not only in the brain tissue but also
20 in the CSF subarachnoid spaces that contain macrophages and constitute the only site of HIV
21 replication in the brain (Cunningham et al., 1997; Ghersi-Egea et al., 1996). It is therefore
22 important to obtain formulations that are able to target AZT to the CSF, but it is equally
23 important to induce the ability of AZT to elude the AET systems in order to avoid its
24 extrusion from the CNS and the associated macrophages. In this regard, we know that the
25 prodrug UDCA-AZT, which was obtained by the conjugation of AZT to ursodeoxycholic acid
26 (UDCA), is able to elude the AET systems (Dalpiaz et al., 2012). Moreover, the results

1 reported in this paper demonstrate for the first time that the prodrug UDCA-AZT is more
2 efficient than AZT in permeating and remaining inside the murine macrophage-like J774A.1
3 cell line, which is commonly used as model system for studying the internalization process of
4 macrophages (Wang et al., 2012). Human cells can be used for *in vitro* studies of
5 inflammation and infection, and many similarities between immortalized human monocytic
6 cell lines (e.g., U937 and THP-1) and primary human blood monocytes have been reported
7 (Yagil-Kelmer et al., 2004). However, these immature cells often require the addition of
8 stimulant factors to promote their differentiation to a mature, adherent phenotype (Shelley et
9 al., 2002). This requirement complicates the study of inflammatory outcomes related to these
10 cell types, as stimulant factors exert specific effects on cellular behavior. Additionally,
11 maturation protocols vary across studies. In contrast, murine cell lines offer the advantage of
12 an immortalized, relatively stable, mature, adherent macrophage phenotype. The murine cell
13 lines J774A.1, RAW 264.7 and IC-21 exhibit the maturity markers F4/80 and Mac-1,
14 indicating their macrophage-like phenotypes, and have been used as models of macrophage
15 activation in numerous studies (Chamberlain et al., 2009). As an example, J774 mouse
16 macrophages have been successfully used to demonstrate the uptake of azidothymidine
17 triphosphate (AZT-TP)-loaded nanoparticles and the related efficient delivery of AZT-TP into
18 the cell cytoplasm (Giacalone et al., 2013).

19 Moreover, our results demonstrate that the prodrug UDCA-AZT is hydrolyzed in
20 macrophages, facilitating intracellular AZT release. Interestingly, no toxicity was observed in
21 the macrophages during their incubation with UDCA-AZT. We demonstrated previously that
22 this prodrug is not a substrate of human AETs (Dalpiaz et al., 2012). This result suggests that
23 UDCA-AZT should not be subject to efflux from human macrophages. Furthermore, AZT is
24 known to induce anti-HIV activity in the HTLV-1-transformed cell line MT-4, which is
25 highly susceptible to and permissive for HIV infection (Magnani et al., 1996), thus suggesting
26 that AZT is able to induce anti-HIV activity in human macrophages.

1 It is important to note that the prodrug UDCA-AZT is quickly hydrolyzed in whole
2 blood, so intravenous administration is not suitable for its permeation across the BBB. Indeed,
3 following this type of administration, we detected only AZT, which exhibited concentration
4 values that decreased over time with a half-life of approximately 1 h. No AZT or
5 UDCA-AZT were detected in the CSF of rats after intravenous administration. In contrast,
6 the nasal administration of this prodrug appears promising for achieving uptake into the CNS
7 and particularly the CSF. We demonstrated that the nasal administration of chitosan-based
8 microparticles can induce the selective uptake of neuroactive agents into the CSF of rats,
9 probably by promoting drug permeation across the olfactory nasal mucosa (Dalpiaz et al.,
10 2008; Gavini et al., 2011; Rassa et al., 2015a). Very recently, the poor water solubility of
11 UDCA-AZT encouraged us to encapsulate this prodrug in solid lipid microparticles (Dalpiaz
12 et al., 2014). In particular, the nasal administration of UDCA-AZT loaded in stearic acid-
13 based microparticles induced the selective uptake of the prodrug into the CSF of rats, with
14 amounts of up to approximately 0.4 $\mu\text{g/ml}$ within 60 min after administration (Dalpiaz et al.,
15 2014). These amounts increased (up to approximately 1.50 $\mu\text{g/mL}$ within 120 min after
16 administration) when the same quantity of microparticles were administered nasally as a
17 water suspension in the presence of chitosan chloride (Dalpiaz et al., 2014). This result was
18 attributed to the good mucoadhesive properties of chitosan (Dalpiaz et al., 2008) and to its
19 ability to transiently open the tight junctions in the epithelial membranes (Dodane et al, 1999;
20 Illum et al., 1994).

21 Nasal formulations obtained in the presence of water can be associated with the risk of
22 chemical and physical instability and microbiological growth. Moreover, in the nasal cavity,
23 liquid formulations are often rinsed into the GIT or out of the nose, so their residence time is
24 generally short (Kublik and Vidgren, 1998). These disadvantages can be overcome by using
25 powder-based formulations (Marttin et al., 1997; Rassa et al., 2015b). For this reason, we
26 prepared microspheres based on chitosan chloride in order to increase the nose-to-brain

1 delivery of UDCA-AZT. The efficacy of the microparticulate system for the brain delivery of
2 the prodrug was verified by comparing the properties of the microparticles with those of their
3 parent physical mixture, which was obtained by geometrical dilution of the components using
4 an agate mortar and pestle. The grinding process induced a reduction of the d_{vs} diameters of
5 chitosan chloride and the UDCA-AZT particles. The chitosan chloride particles were
6 characterized by a round morphology and a relatively smooth surface before and after
7 grinding, whereas UDCA-AZT appeared as fragments with an irregular shape. These two
8 different morphological characteristics were distinctly apparent in the SEM picture of the
9 physical mixture, where small irregular fragments of UDCA-AZT appeared on the surface of
10 larger and rounder chitosan chloride microspheres.

11 The spray-drying process allowed us to obtain unloaded microparticles of chitosan
12 chloride (CH) with the “classical” round morphology and a small d_{vs} diameter. The loaded
13 microparticles (CP) evidenced a slight increase in size and a morphology characterized by
14 spherical particles mixed with fragments of irregular shape and high porosity, similar to
15 spongy balls, with some degree of aggregation. We can conclude that the spherical particles
16 are attributable to pure chitosan chloride that has not interacted with UDCA-AZT during the
17 spray-drying process, whereas the “spongy balls” were obtained by a combination of the
18 prodrug with polymer, inducing the formation of the highly porous structures. The different
19 degrees of water solubility between UDCA-AZT and chitosan can justify these characteristics
20 of the CP formulation. The description of the CP formulation’s morphology appears to be
21 consistent with the observed dissolution and water uptake profiles. Indeed, we observed that
22 the dissolution rate of UDCA-AZT from the CP sample was significantly higher than that of
23 the parent physical mixture. This phenomenon can be attributed to the highly porous
24 structures observed in the CP sample, which allows a faster dissolution of UDCA-AZT with
25 respect to its fragments included in MIX. Moreover, as demonstrated previously by Gavini et
26 al. (2011), water soluble chitosans, such as salts, are more able to completely amorphize

1 poorly soluble drugs into the polymer matrix compared to the chitosan base; as a
2 consequence, the dissolution rate and bioavailability of poorly soluble drugs improve.

3 Moreover, we observed that the water uptake rate of the CH sample was significantly
4 reduced if chitosan chloride was mixed with UDCA-AZT, but a drastic decrease was
5 observed in the case of the CP sample. The presence of UDCA-AZT, which is a very poorly
6 water soluble molecule, in the physical mixture can explain its reduction of water uptake with
7 respect to the CH sample, but the drastic reduction observed for the CP microparticles can be
8 attributed to their porous structures, in which the presence of the prodrug and chitosan
9 chloride together contributed to dramatically reducing the water uptake aptitude of the
10 polymer.

11 Therefore, the CP sample appeared to be a valuable formulation for the nasal
12 administration of UDCA-AZT. In particular, we hypothesized that the size and density of the
13 microparticles should induce their deposition on the nasal mucosa, which should not be
14 dehydrated by their presence, given the poor the water uptake of this sample. Moreover, the
15 ability of the loaded microparticles to increase the dissolution rate of UDCA-AZT suggested
16 their potential aptitude for inducing prodrug permeation across the nasal mucosa, a
17 phenomenon that is probably potentiated by the ability of chitosan to transiently open tight
18 junctions (Illum et al., 2015).

19 The nasal administration of raw UDCA-AZT to rats did not produce any detectable
20 levels of UDCA-AZT or AZT in the bloodstream or the CSF, whereas the nasal
21 administration of the CP sample and its parent physical mixture allowed us to obtain
22 relatively high levels of UDCA-AZT in the CSF of rats 60 min after administration. The
23 relative bioavailability of the CP sample was 176% of that of its parent physical mixture
24 ($P < 0.001$). No AZT was detected in the bloodstream of rats after nasal administration of the
25 CP sample and its parent physical mixture, confirming the existence of a direct nose-CNS
26 pathway for this prodrug (Dalpiaz et al., 2014). Our results demonstrate that the role of

1 chitosan in inducing selective UDCA-AZT uptake into the CSF is potentiated when the
2 prodrug is formulated as a microparticulate system by spray-drying. This phenomenon is
3 illustrated in Fig. 9, which presents a comparison of the AUC values obtained in the CSF after
4 nasal administration of the same dose of UDCA-AZT (200 μ g) to rats using different
5 formulations (i.e., the solid lipid microparticles (SLMs), their water dispersion in the presence
6 of chitosan (SLMs + Ch), which was obtained previously by Dalpiaz et al. (2014), MIX and
7 CP, which was described here). In particular, CP was able to induce an uptake of UDCA-AZT
8 into the CSF of rats that was 1.8, 3.2 and 18.6 times greater than that of its parent physical
9 mixture (MIX), the dispersion of solid lipid particles in the presence of chitosan (SLMs + Ch)
10 and the SLMs in solid form, respectively.

11 To summarize, after nasal administration, chitosan microspheres induce the efficient
12 uptake of UDCA-AZT into the CSF. We demonstrated previously that this prodrug is able to
13 elude the AETs and avoid their inactivation (Dalpiaz et al., 2012); as a consequence, the
14 ability of the prodrug to elude the AET systems can prevent its extrusion into the bloodstream
15 and induce AZT transport in the macrophages located in the subarachnoid spaces of the CSF.
16 Consistent with these findings, it was demonstrated previously that the nasal co-
17 administration of AZT and probenecid (an inhibitor of AETs) increased AZT uptake into the
18 CSF of rats (Seki et al., 1994).

19 Taking into account the observations that the UDCA-AZT C_{\max} obtained in the CSF
20 after the nasal administration of the sample CP was approximately 5 μ M and that the half-life
21 of the prodrug in the CSF was approximately 90 min, we can hypothesize that multiple nasal
22 administrations every 90 min will allow rats to obtain UDCA-AZT concentrations ranging
23 between approximately 5 and 10 μ M in their CSF (Bourne and Dittert, 1990). We
24 demonstrated that the incubation of macrophages with 10 μ M and 100 μ M UDCA-AZT
25 allowed us to detect prodrug amounts ranging from approximately 0.2 to 1 nmoles in 10^6
26 cells. We also demonstrated that the AZT concentration in macrophages after incubation

1 with UDCA-AZT was 47 μ M. AZT exhibits anti-HIV activity in the HTLV-1-transformed
2 cell line MT4, with an IC₅₀ value of approximately 30 nM (Magnani et al., 1996), which is
3 more than three orders of magnitude lower than the concentration of AZT that we detected in
4 the macrophages after incubation with 100 μ M UDCA-AZT. Therefore, it seems reasonable
5 to deduce that incubation with 5 or 10 μ M UDCA-AZT can allow the macrophages to attain
6 AZT concentrations sufficient to induce anti-HIV activity. As a consequence, taking into
7 account the aspects we described above, we predict that the amount of UDCA-AZT taken up
8 in the CSF following the nasal administration of the loaded microparticles might be sufficient
9 to achieve anti-HIV activity in the CNS.

11 **Conflicts of interest**

12 The authors declare that they have nothing to disclose regarding funding or conflicts
13 of interest relating to this manuscript.

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1 **Figure Captions**

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5 **Fig. 1.** Intracellular levels of AZT and its prodrug UDCA-AZT after incubation (30 min) with
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7 J774A.1 murine macrophages. The incubation concentrations were 100 μ M for AZT and 10
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9 and 100 μ M for UDCA-AZT. The graph also reports the intracellular levels of AZT after the
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11 incubation of the macrophages with 100 μ M UDCA-AZT. The data are reported as the
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13 mean \pm S.D. of three independent experiments. *: P<0.001 versus 100 μ M AZT.
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19 **Fig. 2.** Size distribution of loaded (CP) and unloaded (CH) chitosan microspheres (up), MIX
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21 and chitosan chloride and UDCA-AZT alone, before and after grinding (down). The particle
22
23 size distribution was graphically expressed as curves obtained by plotting the volume of the
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25 particles as a percentage *versus* size (μ m), shown on a logarithmic scale
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31 **Fig. 3.** Scanning electron microscopy (SEM) micrographs of the mixture of chitosan chloride
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33 and UDCA-AZT (87:13 w/w, magnification of 3,610 times) [A]; ground chitosan chloride
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35 (magnification of 3,140 times) [B]; ground UDCA-AZT with magnifications of 512 times [C]
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37 and 10,000 times [D]; chitosan chloride before grinding (magnification of 2,740 times) [E];
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39 and UDCA-AZT before grinding (magnification of 505 times) [F].
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46 **Fig. 4.** Scanning electron microscopy (SEM) micrographs of the loaded microparticles
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48 (sample CP) with magnifications of 11,250 times [A] and 3,620 times [B].
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53 **Fig. 5.** Water uptake profiles of loaded (CP) and unloaded (CH) microspheres over 60 min in
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55 comparison to that of the physical mixture (MIX, chitosan chloride and UDCA-AZT (87:13
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57 w/w). The data are reported as the mean \pm SD of three independent experiments.
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1 **Fig. 6.** *In vitro* release of UDCA–AZT from CP microparticles based on chitosan chloride.

2 The release profile is compared with the dissolution profile obtained for the parent physical
3 mixture of chitosan chloride and UDCA-AZT (87:13 w/w) (MIX). The data are reported as
4 the mean±SD of three independent experiments.

6 **Fig. 7.** Elimination profile of AZT after the infusion 0.200 mg of UDCA–AZT into rats. The
7 data are expressed as the mean±SD of four independent experiments. The elimination
8 followed apparent first order kinetics, as confirmed by the semilogarithmic plot reported in
9 the inset (n =9, r = 0.990, P<0.0001). The half-life of AZT was calculated to be 60.4±3.8 min.

11 **Fig. 8.** UDCA–AZT concentrations (µg/mL) detected in the CSF after the nasal
12 administration of dry powders constituted by loaded CP microparticles and its parent mixture
13 (MIX) of chitosan chloride and UDCA-AZT. Each dose contained 200 µg of the prodrug. The
14 inset reports the semilogarithmic plot of the UDCA-AZT concentrations (µM) in the CSF
15 after the nasal administration of the CP sample. The elimination of UDCA-AZT from the CSF
16 followed apparent first order kinetics, as confirmed by the linear regression of the points
17 ranging from 60 to 180 min (n =4, r = 0.988, p<0.05). The data are expressed as the mean±SD
18 of four independent experiments.

20 **Fig. 9.** A comparison among the AUC values obtained in the CSF of rats after the nasal
21 administration of 200 µg of UDCA-AZT using different formulations (i.e., loaded solid lipid
22 microparticles (SLMs), their water dispersion in the presence of chitosan (SLMs + Ch), the
23 physical mixture of chitosan and prodrug (MIX) and the loaded microparticles based on
24 chitosan chloride (CP)). The data pertaining to SLMs and the SLM + Ch formulations were
25 reported previously (Dalpiaz et al., 2014). The data are expressed as the mean±SD of four
26 independent experiments.

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Table 1.

Volume-surface diameters (d_{vs} , μm) of unloaded (CH) and loaded (CP) microparticles together with d_{vs} values of the parent mixture (MIX) of chitosan chloride and UDA-AZT and its single components before and after grinding by agate mortar and pestle. Data are reported as the mean \pm SD of three independent experiments.

Sample	d_{vs} (μm)
CP	3.59 ± 0.14
CH	2.32 ± 0.11
MIX	6.11 ± 0.02
Grinded Chitosan HCl	12.67 ± 0.82
Grinded UDCA-AZT	5.12 ± 0.02
Chitosan chloride before grinding	19.98 ± 0.06
UDCA-AZT before grinding	23.33 ± 0.06

Figure 1

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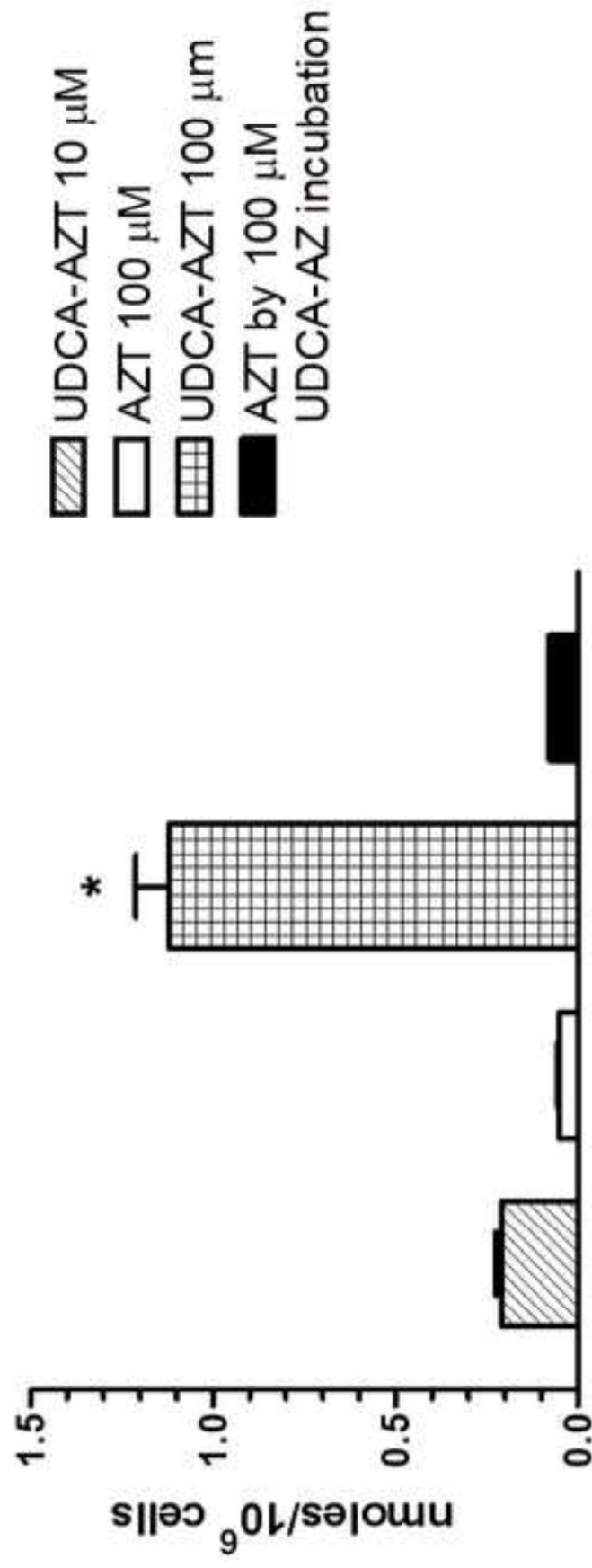


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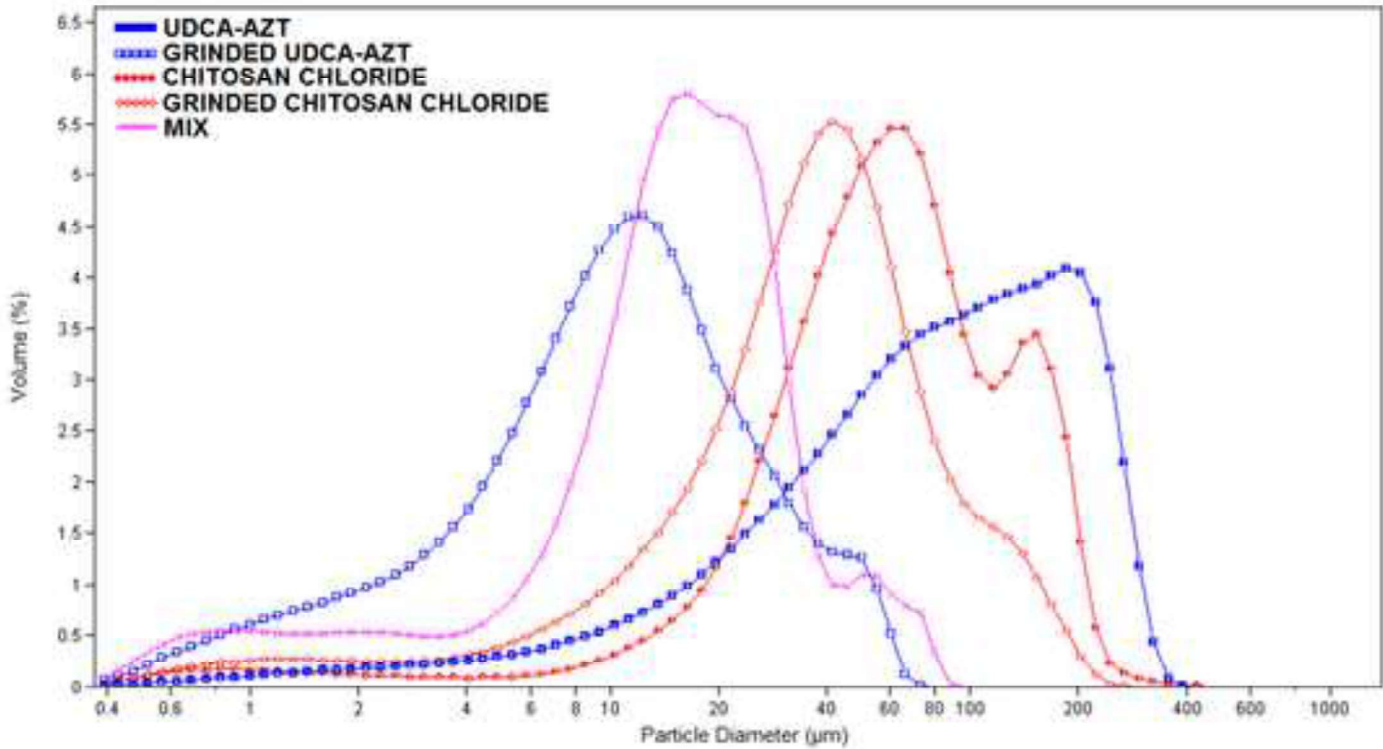
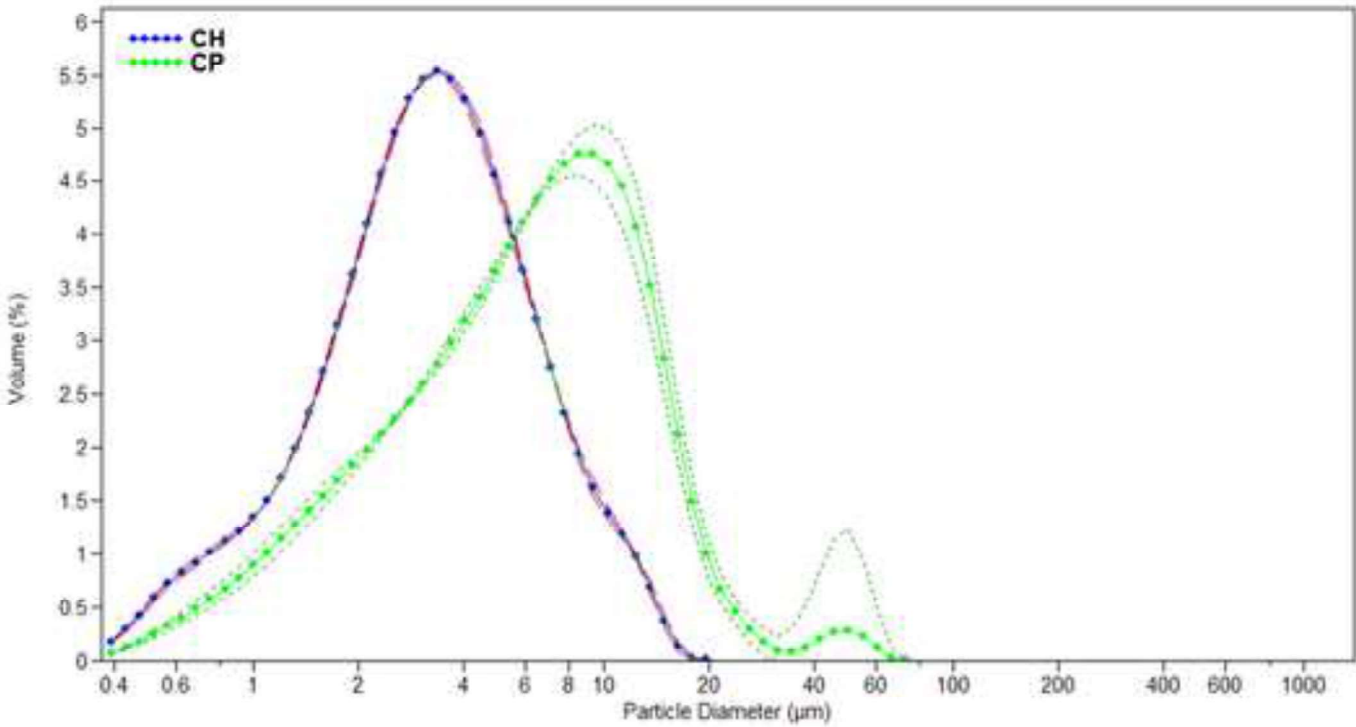


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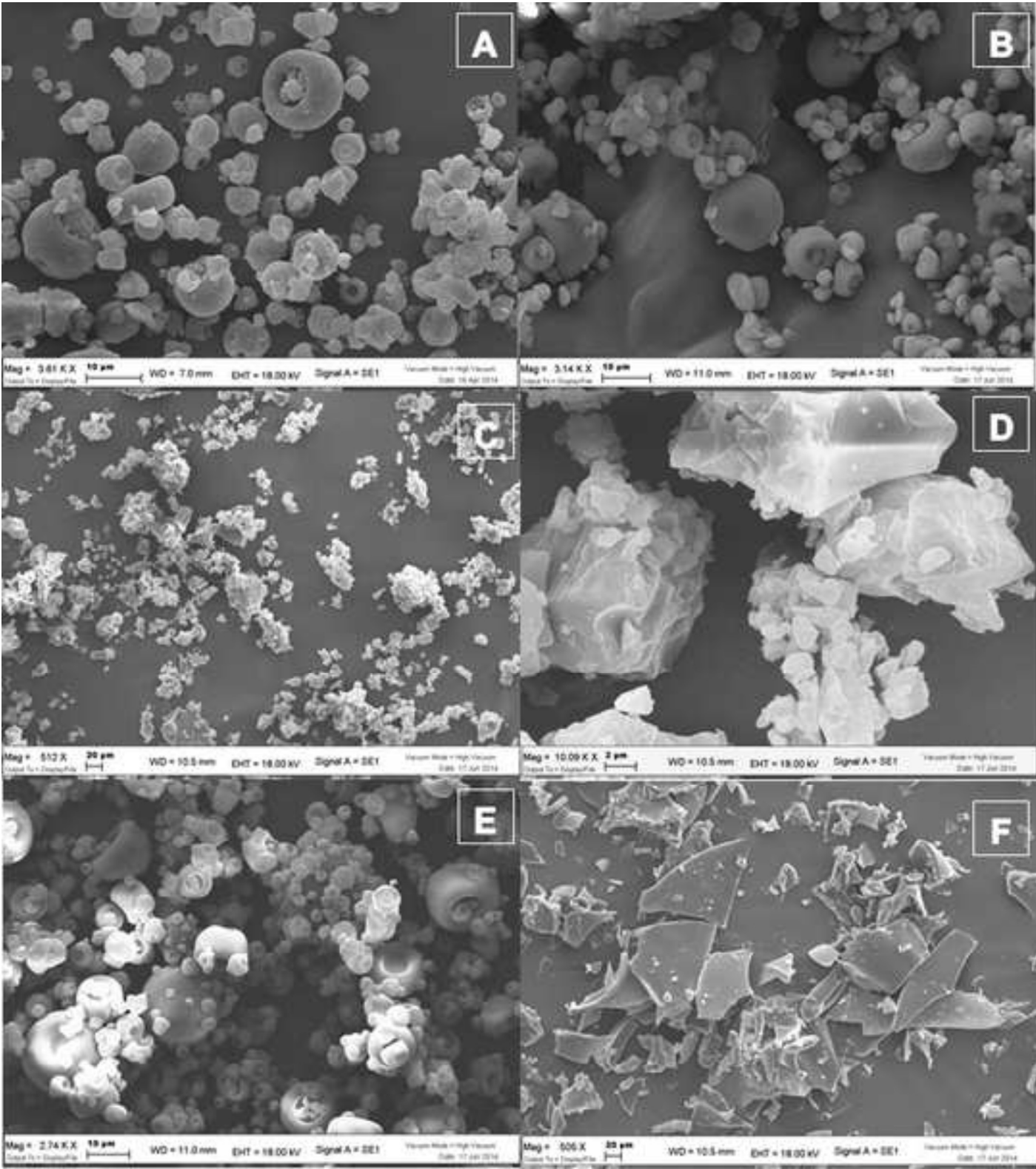


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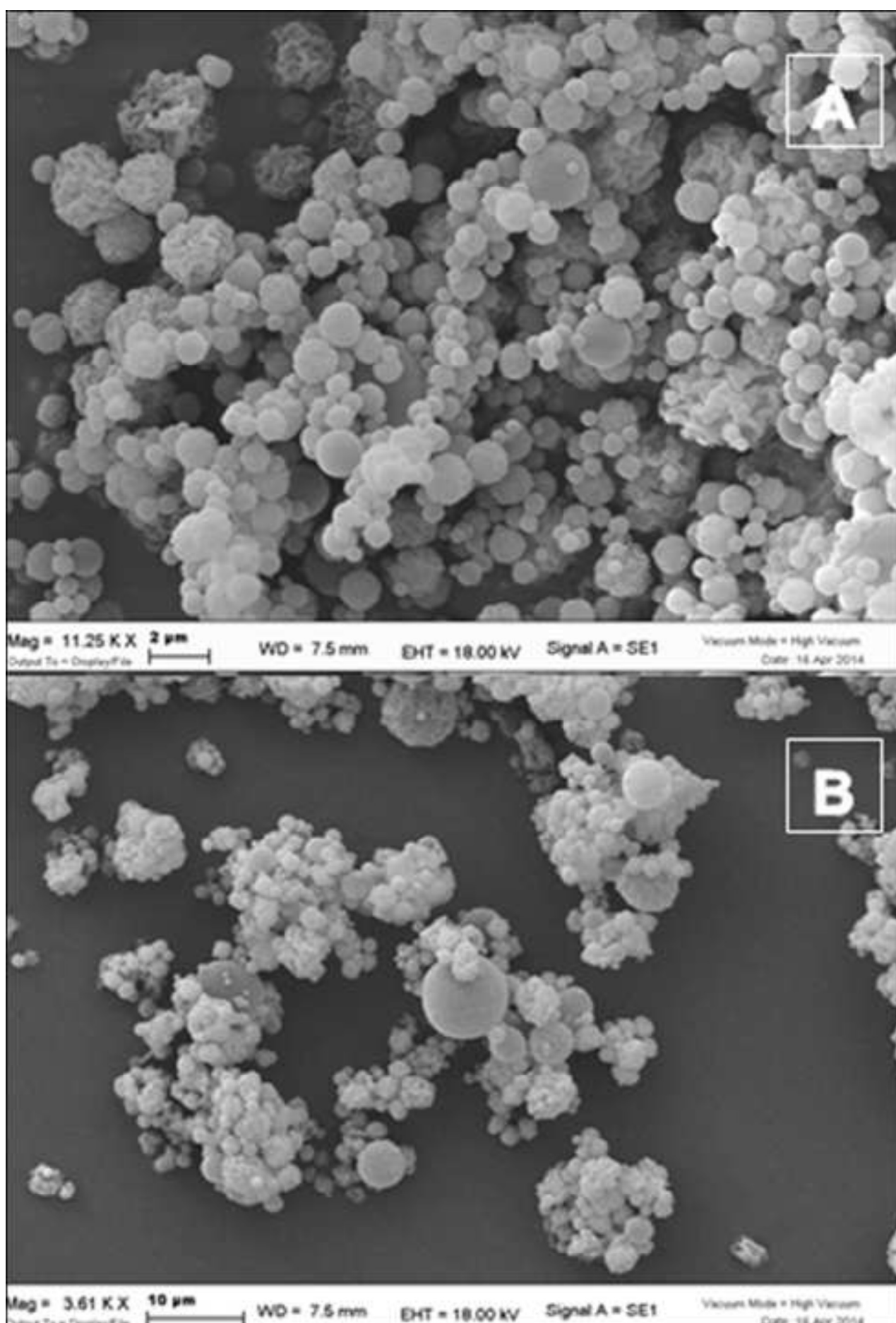


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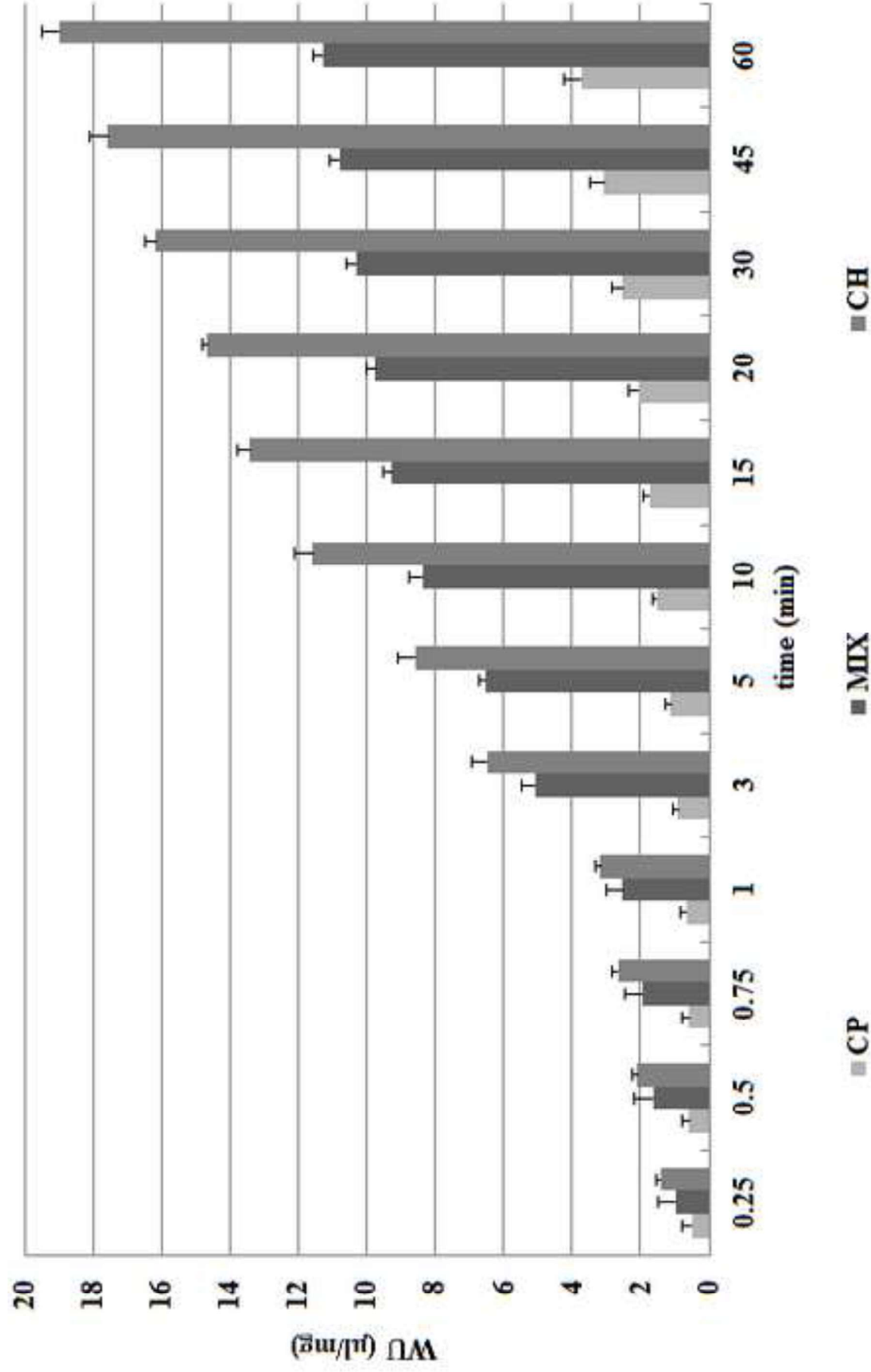


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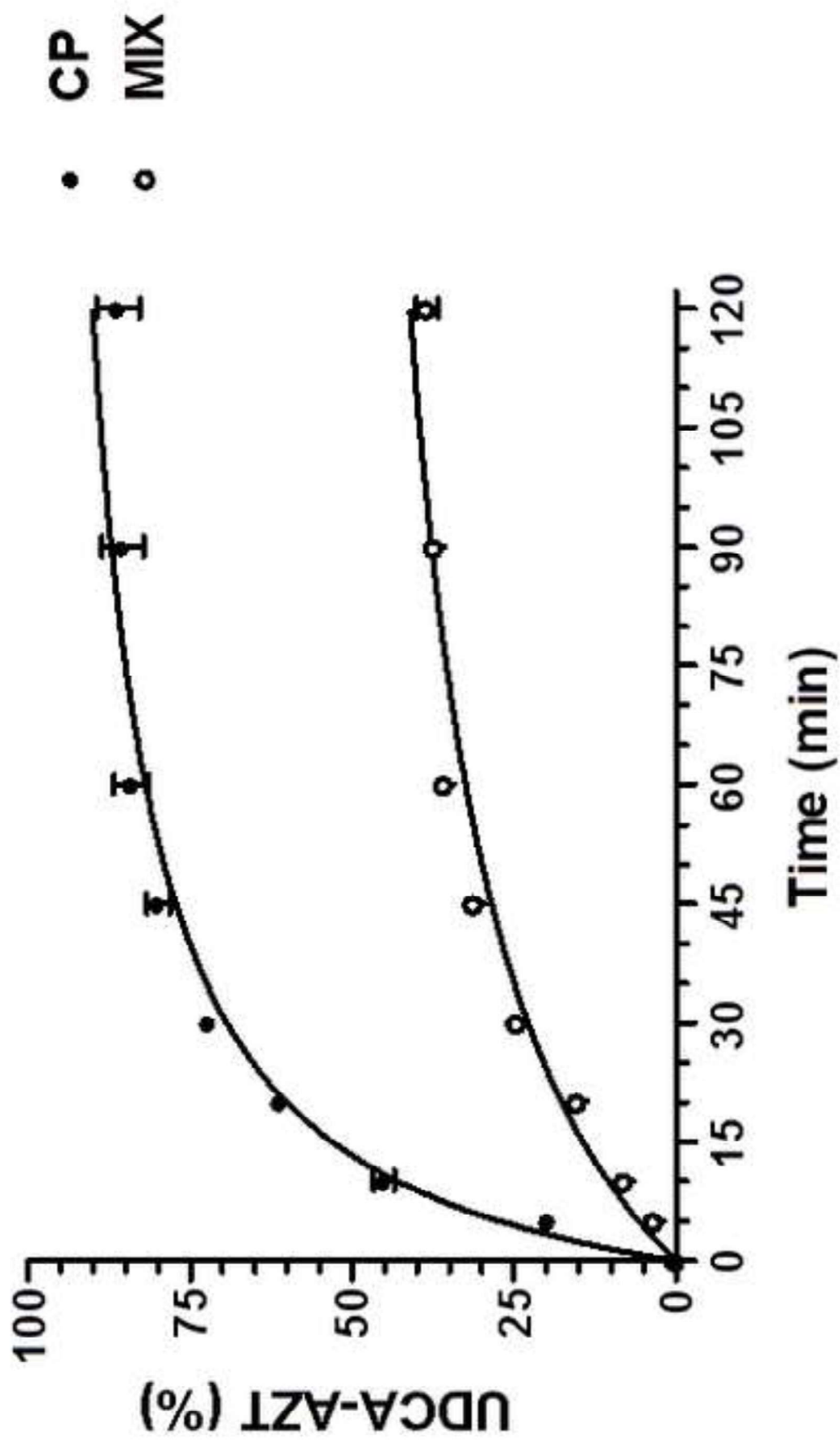


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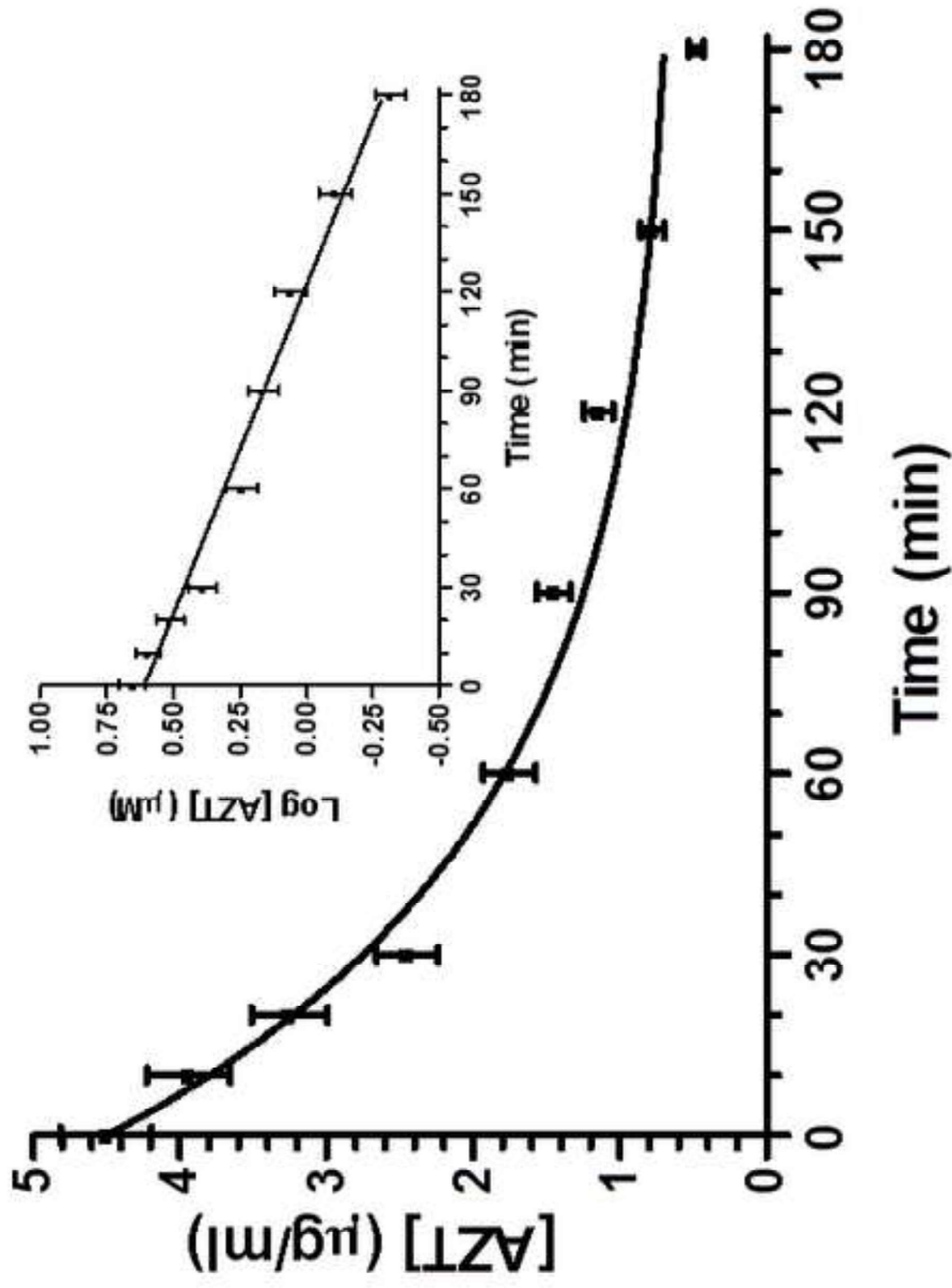


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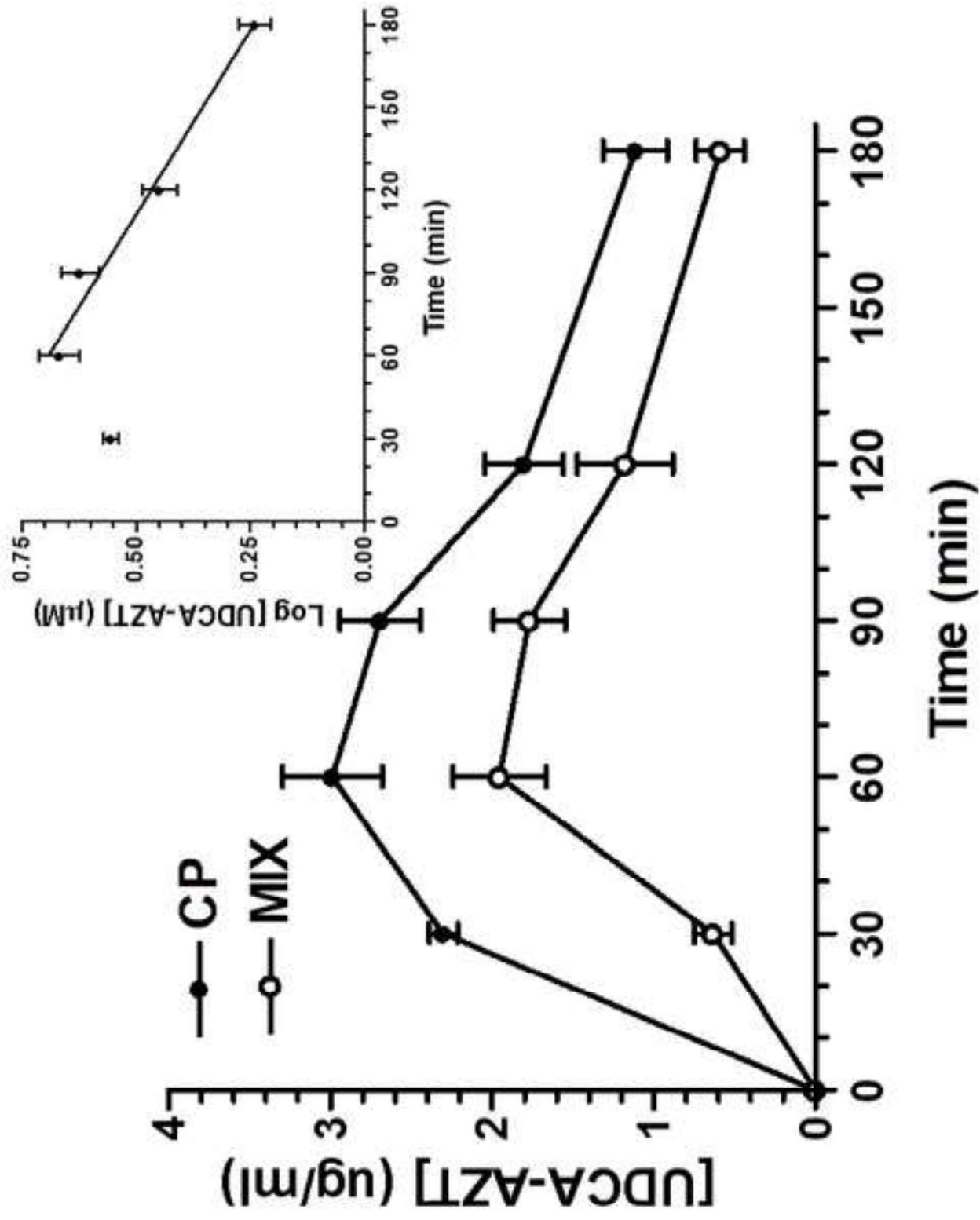


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