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1 **SURGERY ON CERVICAL FOLDS FOR TRANSCERVICAL INTRAUTERINE**
2 **ARTIFICIAL INSEMINATION WITH FROZEN-THAWED SEMEN ENHANCES**
3 **PREGNANCY RATES IN THE SHEEP.**

4
5 Pau Salvatore^{a,b}, Falchi Laura^{a*}, Ledda Mauro^a, Bogliolo Luisa^a, Ariu Federica^a, Zedda Maria
6 Teresa^a.

7 ^aSection of Obstetrics and Gynecology, Department of Veterinary Medicine, University of Sassari,
8 Sassari, Italy.

9 ^bCentro di Competenza Biodiversità Animale, Sassari, Italy.

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11 *Corresponding author: Falchi Laura. lfalchi@uniss.it

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15 **ABSTRACT**

16 In sheep industry, genetic progress rate achieved by artificial insemination (AI) is limited by the
17 convoluted anatomy of the cervix, which does not allow the passage of an insemination catheter for
18 uterine semen deposition. The aim of this study was to test, in 98 pregnant at term Sarda ewes, the
19 effects of: Experiment 1) total or partial ablation of cervical folds and Experiment 2) 4 or 2
20 incisions of cervical folds, on the passage of an insemination catheter, deposition of frozen-thawed
21 semen and pregnancy rates. Surgical procedures were performed within 24h from parturition
22 providing deep sedation and epidural anaesthesia. Duration of surgeries and post-operative recovery
23 were carefully monitored. For both experiments, 5 months since surgery, independently of the stage
24 of oestrus cycle, cervical patency was tested through the transcervical passage of a palpation probe.
25 Six months since surgery, in Experiment 1, ewes were naturally mated with fertile rams. In
26 Experiment 2, ewes submitted to incisions of the cervical folds and a control group underwent
27 synchronization of oestrus and transcervical AI with frozen-thawed semen. Thirty days later, for
28 both experiments, pregnancy rates were assessed by ultrasonography and lambing rates were
29 recorded. Five months after surgery, in Experiment 1, transcervical passage of a palpation probe to
30 reach the uterine lumen was possible in all ewes submitted to total and partial ablation of folds. In
31 Experiment 2, this was achievable in 90.5% ewes with 4 incisions of the folds and in 89.6% ewes
32 with 2 incisions with no significant differences among groups ($P=0.44$). In Experiment 1,
33 pregnancy rates in ewes mated to rams after total or partial ablation of the cervical folds was 100%.
34 In Experiment 2, following transcervical AI, pregnancy rates were higher in groups submitted to 4

35 (63.7%) or 2 (41.4%) incisions of the cervical folds compared to the control group (8%; $P < 0.05$).
36 These data were confirmed at lambing with rates of 56.8% and 41.4% in ewes submitted to 4 or 2
37 incisions respectively, significantly higher than the control group (4%; $P < 0.05$). Surgical ablation or
38 incision of the cervical folds in post-partum ewes represent valid procedures for transcervical
39 intrauterine deposition of semen for AI, obtaining satisfactory pregnancy rates. These procedures
40 might be useful in programs of genetic selection and MOET.

41

42 Key words: cervical surgery, fertility, frozen-thawed semen, sheep, transcervical insemination,
43 lambing.

44

45 **1. Introduction**

46 Programs of genetic improvement are the base for progress in farm animal breeding. In the sheep
47 industry this could be easily accomplished by a method of artificial insemination (AI) that is
48 reliable and economically sustainable. However, in this species it has a poor uptake, the main
49 reason being the poor quality and short life of frozen-thawed semen caused by damage to the
50 spermatozoa associated with cryopreservation and thawing [1]. The impaired ability of frozen-
51 thawed spermatozoa to move through the female reproductive tract and to reach the site of
52 fertilization is one of the major problems of AI in the sheep. This is summed up to the impossibility
53 to deposit the semen directly in the uterine lumen, because of the convoluted anatomy of the sheep
54 cervix. This structure is in fact characterized by a series of funnel-shaped folds that protrude
55 caudally and are often misaligned, precluding the transcervical passage and intrauterine delivery of
56 semen by using conventional AI catheters. The anatomy of the sheep cervix is also highly variable
57 among individuals. Breed, age, parity and physiological state [2-4] might influence its shape and
58 degree of relaxation explaining the variability in the success of transcervical AI. These limiting
59 factors explain the reason why, in the ovine species, AI is mostly performed using fresh semen
60 deposited in the external *os* of the cervix (cervical insemination)[5].

61 Many attempts, mainly mechanical and hormonal, have been made in the past to overcome this
62 anatomical barrier. Some studies focused on the design of new insemination catheters [6-10], but
63 their successful passage through the cervix and consequent deposition of semen in the uterus was
64 strongly influenced by the above mentioned differences in the breed and age of the animals [3, 4].

65 Another approach has been the use of hormonal treatments to enhance the dilation of cervical canal,
66 mimicking the pathway that involves the oxytocin-mediated synthesis of PGE_2 enhanced by
67 gonadotropins and oestrogens. Prostaglandins E_2 act on both cervical extra-cellular matrix and
68 smooth muscle layers leading respectively to re-arrangement of collagen bundles and relaxation

69 [11]. Among others, 17β -oestradiol [12, 13], oxytocin[13-15], FSH [4, 16] and PGE₂ analogues [4,
70 16-18] were tested. Other studies investigated the effects of myorelaxing substances [19] or
71 cytokines [20]. All of these methods have been, at best, only partially successful and in some cases
72 completely unsuccessful with respect to pregnancy rates.

73 To our knowledge, no attempt has been made to enhance cervical patency and transcervical passage
74 of insemination catheters by a surgical approach. Therefore, the aim of this study was to test, in
75 pluriparous Sarda ewes: Experiment 1) if total or partial surgical ablation of cervical folds would
76 allow the passage of an insemination catheter through the cervix up to the uterine lumen and would
77 affect pregnancy and lambing rates after natural mating; Experiment 2) if a less invasive surgical
78 procedure based on 4 or 2 incisions of cervical folds could enhance cervical patency and allow
79 uterine deposition of frozen-thawed semen with satisfactory pregnancy rates.

80

81 **2. Materials and methods**

82

83 **2.1 Animal management and experimental plan**

84 The study started during lambing season (October-November 2015-2016) in Sardinia, Italy, and all
85 the procedures were carried out under the European regulations on the Care and Welfare of Animals
86 in Research. The experiments were performed on a total number of 98 multiparous Sarda ewes aged
87 between 3 and 4 years old, all pregnant at term, randomly assigned to one of the 4 different surgical
88 procedures on cervical folds, carried out in 2 different experiments. The experimental plan is shown
89 in Fig.1.

90 **2.2 Surgical procedures**

91 Surgery was performed within 24h from parturition taking care that expulsion of fetal membranes
92 had occurred. All animals were initially submitted to mild sedation with acepromazine maleate
93 (0.5mL/50Kg BW, IM, Prequillan, Fatro S.p.A., Italy) and, after careful trichotomy and disinfection
94 of the sacrococcygeal area, epidural anaesthesia was achieved by injection of Lidocaine 2%
95 (30mg/10kg BW, Esteve S.p.A., Italy). The ewes were then placed in a cradle in dorsal recumbency
96 with the hindquarters slightly elevated (Trendelenburg position). The perineal area and vulva were
97 carefully cleaned with an antiseptic solution of 10% povidone iodide and after setting up the
98 surgical field, a lubricated speculum was gently inserted in the vagina in order to locate the external
99 os of the cervix and its folds. The most caudal fold was then grasped with Duval forceps and the
100 cervix was gently retracted up to the vulva (Fig.2a). With the aid of Duval forceps, the remaining
101 folds, up to the most cranial one, were progressively grasped and retracted (Fig.2b), exteriorizing

102 them completely (Fig.2c). All surgical procedures were performed under sterile conditions and their
103 duration was recorded.

104 At this point, cervical folds were either completely (n.ewes=5) or partially removed (n.ewes = 20;
105 Experiment 1) or incised in 4 sites (dorsally, ventrally and 2 laterally; n.ewes = 44) or in 2 sites
106 (dorsally and ventrally; n.ewes = 29; Experiment 2).

107 **2.2.1 Experiment 1: Total or partial ablation of cervical folds**

108 Total ablation was performed excising each fold from the most cranial one at 2-3mm from the base
109 with Metzenbaum scissors (Fig. 3a). The edges of the wound were immediately sutured with a
110 Schmieden suture (monofilament polyglecaprone 25,USP 5/0, Vetsuture®, Paris, France) that was
111 interrupted and restarted in 4 points (dorsal, ventral and laterals).

112 Partial ablation was performed excising from each fold 2 trapezoid-shape pieces of tissue dorsally
113 and ventrally at 2-3 mm from the base of the fold (Fig. 3b). These portions were removed with
114 electrocautery.

115

116 **2.2.2 Experiment 2: 4 or 2 incisions of cervical folds**

117 A schematic representation of the sites of incision of the cervical folds is given in Fig. 3. For every
118 fold, after exteriorisation and distension of the tissue, either 4 (dorsal, ventral and 2 lateral) or 2
119 (dorsal and ventral) incision areas were delimited by 2 Dandy forceps (Fig.4a-5a) and cut by
120 electrocautery (Fig.4b-5b).

121 **2.2.3 Post operatory care**

122 After surgery, a topical antibiotic treatment (Orbenin; cloxacillin suspension, Pfizer Italy Srl) was
123 applied and the cervix was repositioned. All animals were kept under careful post operatory
124 observation for 24h, and afterwards reintroduced in the flock. Milk production performances were
125 monitored by the farmer during lactation.

126 **2.3 Assessment of cervical patency after surgery**

127 Five months since surgery, all females in both experiments were evaluated for patency of the
128 cervical canal and easiness in transcervical passage of a probe up to the uterine lumen. In detail,
129 ewes were restrained in a cradle in Trendelenburg recumbency and after cleaning the vulvar area
130 and inserting a vaginal speculum, the cervix was gently retracted caudally up to the vulva with the
131 aid of Bozeman forceps. A palpation probe (commonly used in laparoscopic procedures, 3.5mm in
132 Ø, Richard Wolf, USA), was inserted through the cervical canal. The patency test was carried out
133 without considering the stage of oestrus cycle of the ewes. The ability and the time taken in
134 reaching the uterine lumen was recorded and the easiness in the passage of the probe was scored
135 from I to IV(I= very easy; II= easy; III= moderately difficult; IV=difficult).

136 **2.4 Natural mating in ewes with total or partial ablation of the cervical folds**
137 **(Experiment 1)**

138 In order to test if total or partial ablation of cervical folds affected oestrus behaviour, mating and,
139 finally, pregnancy rates, one month after the assessment of cervical patency, all ewes surgically
140 treated with either total (n=5) or partial (n= 20) ablation of cervical folds were synchronised using
141 intravaginal progestagen sponges (Crono-gest 20mg, Intervet Italia S.r.l, Italy) for 14 days. On the
142 day of sponge removal, 300 IU of PMSG (Folligon, Intervet Italia S.r.l, Italy) were injected IM. The
143 ewes were then allowed to mate to 3 adult rams of proven fertility for 2 consecutive cycles.

144 **2.5 Transcervical artificial insemination with frozen-thawed semen in ewes with 4 or 2**
145 **incisions of cervical folds (Experiment 2)**

146 **2.5.1 Semen preparation**

147 Briefly, semen was collected by artificial vagina from 3 rams of proven fertility and only ejaculates
148 with a score of mass motility ≥ 3 (scale of 0-5; 0= no motility, 5 = vigorous swirling waves of
149 movements) and $\geq 3 \times 10^9$ spz/mL were further processed. Semen was pooled (to avoid individual
150 variability) diluted in home-made Tris-EY (Egg Yolk) based extender with 6% glycerol to reach a
151 concentration of 1.6×10^9 spz/mL (400×10^6 spz/straw), cooled to 4°C and loaded into 0.25mL straws
152 (IMV technologies, France). The straws were submitted to LN₂ vapors and then plunged and stored
153 in LN₂ until the day of insemination. Straws were then thawed warming them at 37°C for 30 sec.
154 An aliquot of thawed semen (5µL) was collected and assessed for motility parameters through
155 CASA (computers assisted sperm analysis; Ivos, Hamilton Thorne, Biosciences). Total and
156 progressive motility were 65 and 45% respectively.

157 **2.5.2 Artificial insemination with frozen-thawed semen**

158 Six months after surgery, in order to assess pregnancy rates, the ewes that underwent incisions of
159 the cervical folds (4 incisions, n=44; 2 incisions, n=29), and a control group of 25 animals (no
160 surgery) were synchronised using intravaginal progestagen sponges (Crono-gest 20mg, Intervet
161 Italia S.r.l, Italy) for 14 days. On the day of sponge removal, 300 IU of PMSG (Folligon, Intervet
162 Italia S.r.l, Italy) were injected IM. At 56-58h from sponge removal, transcervical artificial
163 insemination was performed using frozen-thawed semen. Ewes were placed in dorsal recumbence
164 in a cradle, the perineal and vulvar area were carefully cleaned with an antiseptic solution and a
165 lubricated speculum was gently inserted in the vagina. The fold of the external os of the cervix was
166 localised and gently extruded using Bozeman forceps up to the vulvar vestibulum (Fig.6). The
167 insemination catheter (Cassou mini-pistolet for ovine-caprine; IMV technologies, France), loaded
168 with thawed semen, was then inserted through the cervical canal and the semen was deposited,

169 when possible, directly in the uterine lumen. The animals in which passing the cervix to reach the
170 uterus was not possible were recorded and semen was deposited in the cervix as deep as possible.

171 **2.6 Pregnancy detection**

172 For both experiments, return to oestrus was checked by introducing teaser rams wearing harnesses
173 with crayons in the experimental groups from 15 to 20 days after artificial insemination. Pregnancy
174 rate (pregnant ewes/ inseminated ewes) was determined at 30 days after insemination by transrectal
175 ultrasonography (MyLab One, Esaote, Italy). Lambing rate was also recorded.

176 **2.7 Statistical analyses**

177 Statistical analysis was performed using Stata 11.2/IC (StataCorp LP, USA). Continuous data
178 regarding the duration of surgery, the time taken to pass the cervix with the palpation probe during
179 the assessment of post-surgery cervical patency were not normally distributed and were analysed by
180 non-parametric Kruskal-Wallis test followed by two-samples Wilcoxon rank-sum test for pairwise
181 comparisons with Bonferroni's correction. Categorical data regarding the ability to reach the uterus
182 with the probe, the easiness in passing through the cervical canal, pregnancy rates and lambing rates
183 were analysed by χ^2 -test. The significance level was defined for $P < 0.05$.

184 **3. Results**

185 **3.1 Surgical procedures**

186 The mean duration of surgery was, for partial ablation (Exp.1) and 4 or 2 incisions of cervical folds
187 (Exp.2), 28 ± 6 min and no difference among these procedures was observed ($P > 0.05$). Total
188 ablation of cervical folds took around 30 additional minutes due to suturing time.

189 After the 24h of post operatory observation, all animals submitted to surgery were in good health
190 conditions and were reintroduced in the flock. Milk production was not affected by the surgical
191 procedure.

192 **3.2 Post surgical assessment of cervical patency**

193 The results obtained from the post surgical assessment of cervical patency are summarised in Table
194 1.

195 **3.2.1 Experiment 1**

196 Four months after surgery, the passage of the probe through the cervical canal up to the uterine
197 lumen was allowed in all ewes submitted to total (5/5, 100%) and partial (20/20, 100%) ablation of
198 cervical folds.

199 **3.2.2 Experiment 2**

200 Reaching the uterus was achievable in 40/44 (90.9%) ewes that underwent 4 incisions of the folds
201 surgery and in 26/29 (89.6%) ewes that underwent 2 incisions surgery. The differences among
202 procedures were not statistically significant ($P > 0.05$). In those subjects in which the uterine lumen

203 was reachable, passing the cervical canal was easier and effortless in ewes submitted to ablation of
204 the folds compared to those that underwent incision ($P<0.01$). In the group submitted to 2 incisions,
205 the passage of the probe was easy but not effortless compared to the other groups ($P<0.001$).

206 In addition, the time spent in reaching the uterine lumen was significantly lower in ewes that
207 underwent total (2.4 ± 0.5 sec) or partial (4.9 ± 3 sec) ablation of cervical folds compared to those
208 that had 4 (21 ± 26 sec) or 2 (26.2 ± 21 sec) incisions ($P<0.05$); no significant difference was found
209 between the latter 2 groups ($P>0.05$).

210 **3.3 Pregnancy and lambing rates**

211 **3.3.1 Experiment 1**

212 Ultrasound check at 30 days after natural mating in ewes submitted to total or partial ablation of
213 cervical folds showed pregnancy rates of 100% in both groups at the first oestrus after
214 synchronisation. No relevant problems were reported during pregnancy and lambing.

215 **3.3.2 Experiment 2**

216 The site of deposition of frozen-thawed semen during transcervical AI in the control and in 4 or 2
217 incisions groups is reported in Table 2. Semen deposition in the uterus was possible in none of the
218 ewes of the control group. No return to oestrus at 15-20 days and the ultrasound scanning
219 performed at 30 days after AI with frozen-thawed semen, revealed that pregnancy rates were
220 significantly higher in the groups of ewes submitted to 4 (28/44; 63.7%) or 2 (12/29; 41.4%)
221 incisions of the cervical folds compared to the control untreated group (2/25; 8%; $P<0.001$). Among
222 the 8 ewes (4 in the 4 incisions group and 4 in the 2 incisions group) in which frozen-thawed semen
223 was deposited in the cervix, only 1 ewe with 4 incisions of the cervical folds was pregnant and
224 lambled regularly. Data on pregnancy rates were confirmed at lambing with rates of 56.8% and
225 41.4% in ewes submitted to 4 or 2 incisions respectively, significantly higher than the control group
226 (1/25; 4%; $P<0.001$; Table 2). Moreover, lambing occurred without relevant problems.

227

228 **4. Discussion**

229 The desired practical and commercial diffusion of intrauterine AI will be achieved if and when it
230 becomes possible to pass an insemination catheter through the cervix allowing uterine deposition of
231 semen without causing trauma to the cervix. Eppleston et al. showed that, in the sheep, there was a
232 linear relationship between fertility and depth of deposition of frozen-thawed semen and that, when
233 inseminating into the uterus, the site of deposition did not affect fertility. This suggested that an
234 effective transcervical method of insemination would lead to good fertility rates of around 80%, a
235 similar figure to the one achieved using laparoscopic insemination [21]. The anatomy of the sheep
236 cervix represents a major limiting factor for intrauterine trans-cervical AI in the ovine species. In

237 fact, its lumen is highly convoluted due to the presence of 4-7 cervical folds [2] in 3 distinct
238 sections: the caudal section being the entrance to the external os with a large fold [whose shape has
239 been classified in several previous studies [2, 22]], the central section having the majority of the
240 larger folds, and the cranial section which meets the uterine body at the internal os and in which
241 folds are smaller and less well defined [22]. The cervical folds project caudally into the lumen and
242 are generally out of alignment with the first [23].

243 In the present experiments, we proposed a surgical approach for ablation/incision of cervical folds
244 that allowed the passage of a Cassou insemination catheter and the deposition of frozen semen
245 directly in the uterus. The surgery was performed within the 24h post-partum following expulsion
246 of foetal membranes because during this time the cervix can be easily manipulated reducing the risk
247 of traumas that could compromise the future reproductive ability. In the pre-partum period, the
248 cervix undergoes a series of modifications that results in relaxation of smooth muscle layers and in
249 softening of the connective tissue. This multifactorial event is controlled by reproductive hormones
250 and is characterised by increase in inflammatory cells, in the amount of extra-cellular water and in
251 dispersion of collagen fibrils [24]. The remodelling of cervical tissue provided, in our experiment,
252 the optimal conditions to perform the surgery, limiting the effects of this invasive procedure. In the
253 post- partum ewes submitted to surgery, cervical folds were in fact hypertrophic, softened and could
254 be easily exteriorised from the vulva and manipulated to perform ablation or incision. Moreover,
255 since in the peri-partum period, nearby tissues and ligaments are relaxed, excessive stretching that
256 could result in traumas and potential fibrosis was avoided. This was confirmed, in ewes submitted
257 to the surgery, by the post-operative full recovery and by the full ability to carry the pregnancy at
258 term and lamb with no complications.

259 Moreover, it is worth considering the important role of the cervix as a barrier protecting the
260 endometrium and the conceptus from pathogens and that surgical ablation or incisions of cervical
261 folds do not compromise this function. In fact, no infections were reported in the follow-up of the
262 surgery nor during successive pregnancies. Providing that all animal welfare criteria are met and
263 that sterility conditions are maintained during surgery, we can therefore propose this approach as a
264 safe procedure with no risk for the animal health.

265 Pregnancy and lambing rates achieved after 4 or 2 incisions of cervical folds using frozen-thawed
266 semen were as high as 63% and 56% respectively. This result is similar to those achieved by
267 laparoscopic AI. With the laparoscopic technique, pregnancy rates using frozen-thawed semen are
268 satisfactory and range from 43% [25] to around 72% [26]. However, it is a surgical procedure that
269 cannot be used repeatedly on the same animal for problems of post-manipulation adhesences in the
270 abdominal cavity and ethical issues on animal welfare [27]. Moreover it requires trained personnel

271 and expensive and delicate instruments. Previously reported pregnancy rates obtained by
272 transcervical AI, if fresh semen is used, are satisfactory, ranging around 50-60% [25, 28]. Using
273 frozen-thawed semen, rates range from 30- 32% [25, 29] to less than 10% [30], far below those
274 obtained in this study. The surgical procedures we proposed in the present study are “once in a
275 lifetime” procedures, that are not repeated on the same subject and that allow transcervical
276 intrauterine insemination for the entire reproductive career of the animal (unpublished data). These
277 findings are supported by the observations of the condition of cervical folds after post-surgical
278 lambing (Fig.7).

279 Pregnancy rates were satisfactory in all 4 groups submitted to surgery. However, total ablation of
280 cervical folds requires longer execution times that are inadvisable under field conditions. Pregnancy
281 rates in animals submitted to 4 incisions of cervical folds were numerically but not statistically
282 higher compared to the group submitted to 2 incision (63.7% vs 41.4% respectively). However, we
283 can speculate that, increasing the size of the groups this difference would result statistically
284 significant and therefore the technique of 4 incisions would be preferable. For what concerns partial
285 ablation, the execution times are similar to those of the incisions of cervical folds and the
286 penetration times are very low. This suggests that the above technique, together with the 4 incisions
287 technique, are advisable in in field transcervical AI programs. These two procedures could provide
288 consistent benefit in MOET (multiple ovulation embryo transfer) programs in the ovine species
289 (unpublished data). In conclusion, surgical ablation or incision of the cervical folds in post-partum
290 ewes represent valid procedures for trans-cervical intrauterine deposition of semen for AI, obtaining
291 satisfactory pregnancy rates. Although it is a surgical procedure, animals recovered soon and their
292 productive and reproductive careers were not compromised. Therefore we propose these techniques
293 as useful tools for successful spreading of superior genotypes in selected animals.

294

295 **Conflict of interest**

296 The authors declare no conflict of interest.

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300 Italy.

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302 **References**

303 [1] Gillan L, Maxwell WM. The functional integrity and fate of cryopreserved ram spermatozoa in
304 the female tract. J Reprod Fertil Suppl. 1999;54:271-83.

- 305 [2] Kershaw CM, Khalid M, McGowan MR, Ingram K, Leethongdee S, Wax G, et al. The anatomy
306 of the sheep cervix and its influence on the transcervical passage of an inseminating pipette into the
307 uterine lumen. *Theriogenology*. 2005;64:1225-35.
- 308 [3] Kaabi M, Alvarez M, Anel E, Chamorro CA, Boixo JC, de Paz P, et al. Influence of breed and
309 age on morphometry and depth of inseminating catheter penetration in the ewe cervix: a
310 postmortem study. *Theriogenology*. 2006;66:1876-83.
- 311 [4] Falchi L, Taema M, La Clanche S, Scaramuzzi RJ. The pattern of cervical penetration and the
312 effect of topical treatment with prostaglandin and/or FSH and oxytocin on the depth of cervical
313 penetration in the ewe during the peri-ovulatory period. *Theriogenology*. 2012;78:376-84.
- 314 [5] Cappai P, Sanna SR, Branca A, Fraghì A, Bomboi G. Comparison of laparoscopic and
315 transcervical insemination with frozen semen in Sarda dairy ewes. *Animal Science*. 1998;66:369-
316 73.
- 317 [6] Wulster-Radcliffe MC, Lewis GS. Development of a new transcervical artificial insemination
318 method for sheep: effects of a new transcervical artificial insemination catheter and traversing the
319 cervix on semen quality and fertility. *Theriogenology*. 2002;58:1361-71.
- 320 [7] Wulster-Radcliffe MC, Wang S, Lewis GS. Transcervical artificial insemination in sheep:
321 effects of a new transcervical artificial insemination instrument and traversing the cervix on
322 pregnancy and lambing rates. *Theriogenology*. 2004;62:990-1002.
- 323 [8] Macías A, Ferrer LM, Ramos JJ, Lidón I, Rebollar R, Lacasta D, et al. Technical Note: A new
324 device for cervical insemination of sheep – design and field test1. *Journal of Animal Science*.
325 2017;95:5263-9.
- 326 [9] Álvarez M, Chamorro CA, Kaabi M, Anel-López L, Boixo JC, Anel E, et al. Design and “in
327 vivo” evaluation of two adapted catheters for intrauterine transcervical insemination in sheep.
328 *Animal Reproduction Science*. 2012;131:153-9.
- 329 [10] Halbert GW, Dobson H, Walton JS, Buckrell BC. A technique for transcervical intrauterine
330 insemination of ewes. *Theriogenology*. 1990;33:993-1010.
- 331 [11] Kershaw CM, Scaramuzzi RJ, McGowan MR, Wheeler-Jones CP, Khalid M. The expression
332 of prostaglandin endoperoxide synthase 2 messenger RNA and the proportion of smooth muscle
333 and collagen in the sheep cervix during the estrous cycle. *Biol Reprod*. 2007;76:124-9.
- 334 [12] Akinbami MA, Meredith S, Warren JE, Anthony RV, Day BN. Cervical dilation, conception
335 rate, and concentrations of progesterone and estradiol-17B in postpartum ewes treated with porcine
336 relaxin. *Theriogenology*. 1990;34:927-40.
- 337 [13] Wulster-Radcliffe MC, Costine BA, Lewis GS. Estradiol-17 beta-oxytocin-induced cervical
338 dilation in sheep: application to transcervical embryo transfer. *J Anim Sci*. 1999;77:2587-93.
- 339 [14] Stellflug JN, Wulster-Radcliffe MC, Hensley EL, Cowardin EA, Seals RC, Lewis GS.
340 Oxytocin-induced cervical dilation and cervical manipulation in sheep: effects on laparoscopic
341 artificial insemination. *J Anim Sci*. 2001;79:568-73.
- 342 [15] Khalifa RM, Sayre BL, Lewis GS. Exogenous oxytocin dilates the cervix in ewes. *J Anim Sci*.
343 1992;70:38-42.
- 344 [16] Leethongdee S, Khalid M, Bhatti A, Ponglowhapan S, Kershaw CM, Scaramuzzi RJ. The
345 effects of the prostaglandin E analogue Misoprostol and follicle-stimulating hormone on cervical
346 penetrability in ewes during the peri-ovulatory period. *Theriogenology*. 2007;67:767-77.
- 347 [17] Candappa IB, Bainbridge HC, Price NT, Hourigan KR, Bartlewski PM. A preliminary study on
348 the suitability of Cervidil to induce cervical dilation for artificial insemination in ewes. *Res Vet Sci*.
349 2009;87:204-6.

- 350 [18] Bartlewski PM, Candappa IB. Assessing the usefulness of prostaglandin E2 (Cervidil) for
351 transcervical artificial insemination in ewes. *Theriogenology*. 2015;84:1594-602.
- 352 [19] Santos KC, Monte AP, Lima JT, Ribeiro LA, Palheta Junior RC. Role of NO-cGMP pathway
353 in ovine cervical relaxation induced by *Erythroxyllum caatingae* Plowman. *Anim Reprod Sci*.
354 2016;164:23-30.
- 355 [20] Croy BA, Prudencio J, Minhas K, Ashkar AA, Galligan C, Foster RA, et al. A preliminary
356 study on the usefulness of huIL-8 in cervical relaxation of the ewe for artificial insemination and for
357 embryo transfer. *Theriogenology*. 1999;52:271-87.
- 358 [21] Eppleston J, Salamon S, Moore NW, Evans G. The depth of cervical insemination and site of
359 intrauterine insemination and their relationship to the fertility of frozen-thawed ram semen. *Animal*
360 *Reproduction Science*. 1994;36:211-25.
- 361 [22] Halbert GW, Dobson H, Walton JS, Buckrell BC. The structure of the cervical canal of the
362 ewe. *Theriogenology*. 1990;33:977-92.
- 363 [23] More J. Anatomy and histology of the cervix uteri of the ewe: new insights. *Acta Anat (Basel)*.
364 1984;120:156-9.
- 365 [24] Fosang AJ, Handley CJ. Connective tissue remodelling in the ovine cervix during pregnancy
366 and at term. *Connect Tissue Res*. 1988;17:277-85.
- 367 [25] Masoudi R, Zare Shahneh A, Towhidi A, Kohram H, Akbarisharif A, Sharafi M. Fertility
368 response of artificial insemination methods in sheep with fresh and frozen-thawed semen.
369 *Cryobiology*. 2017;74:77-80.
- 370 [26] Hiwasa M, Kohno H, Togari T, Okabe K, Fukui Y. Fertility after different artificial
371 insemination methods using a synthetic semen extender in sheep. *J Reprod Dev*. 2009;55:50-4.
- 372 [27] Stafford KJ, Chambers JP, Sylvester SP, Kenyon PR, Morris ST, Lizarraga I, et al. The stress
373 caused by laparoscopy in sheep and its alleviation. *N Z Vet J*. 2006;54:109-13.
- 374 [28] Casali R, Pinczak A, Cuadro F, Guillen-Munoz JM, Mezzalira A, Menchaca A. Semen
375 deposition by cervical, transcervical and intrauterine route for fixed-time artificial insemination
376 (FTAI) in the ewe. *Theriogenology*. 2017;103:30-5.
- 377 [29] Windsor DP, Szell AZ, Buschbeck C, Edward AY, Milton JT, Buckrell BC. Transcervical
378 artificial insemination of Australian Merino ewes with frozen-thawed semen. *Theriogenology*.
379 1994;42:147-57.
- 380 [30] Sanchez-Partida LG, Windsor DP, Eppleston J, Setchell BP, Maxwell WMC. Fertility and Its
381 Relationship to Motility Characteristics of Spermatozoa in Ewes After Cervical, Transcervical, and
382 Intrauterine Insemination With Frozen-Thawed Ram Semen. *Journal of Andrology*. 1999;20:280-8.
383

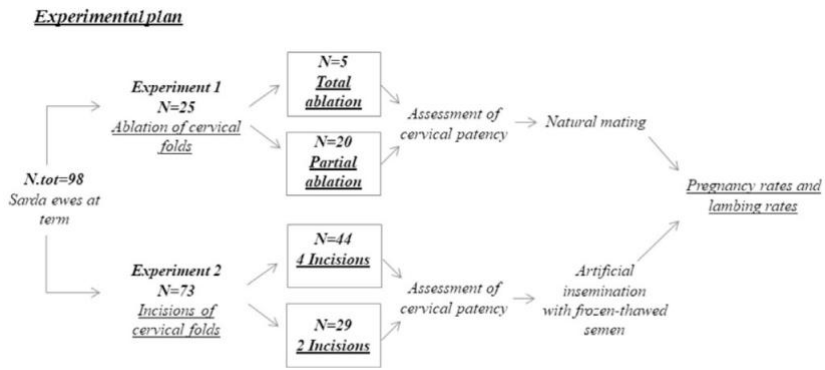


Fig. 1. Experimental design to test the effects of four different surgical procedures on cervical folds on fertility and lambing rates in pluriparous Sarda ewes.

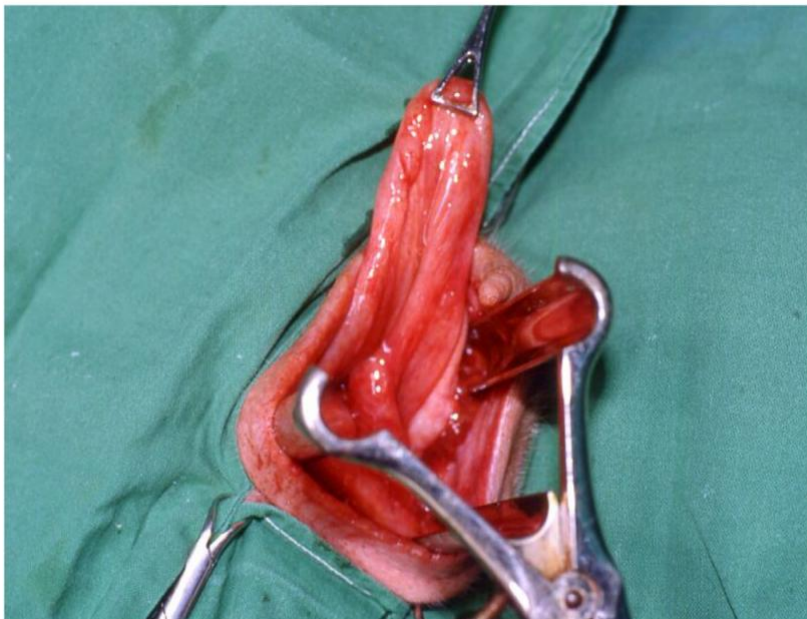


Fig. 2a. Procedure for the exteriorisation of cervical folds: a) the most caudal fold of the external os (arrow) was grasped with Duval forceps and the cervix was gently retracted up to the vulvar vestibulum.

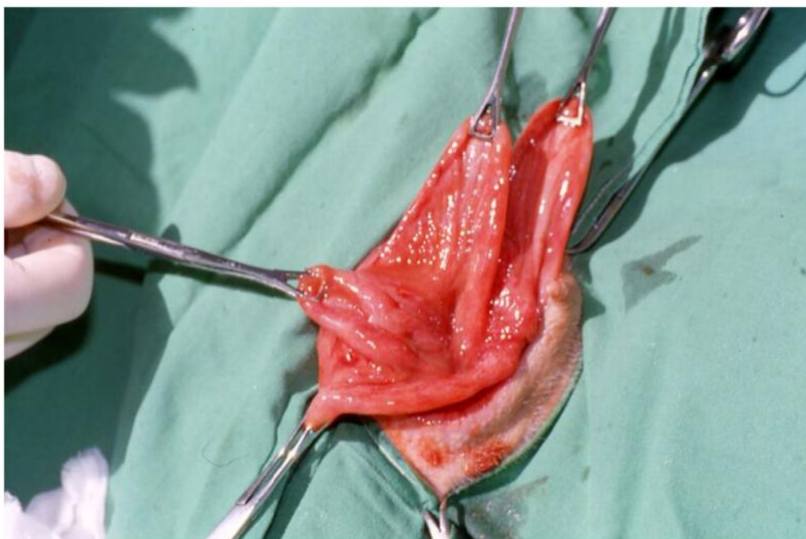


Fig. 2b. Procedure for the exteriorisation of cervical folds: b) progressively, with the aid of other Duval forceps.



Fig. 2c. Procedure for the exteriorisation of cervical folds: c) the cervical canal was completely exteriorised.

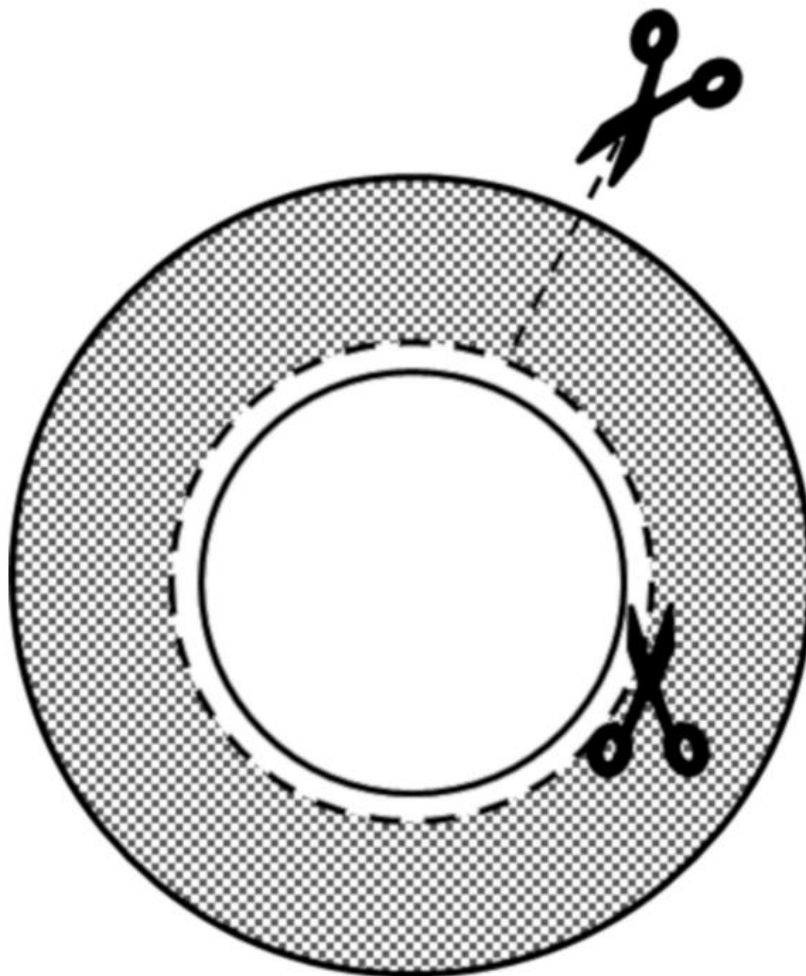


Fig. 3a. Schematic representation of total (a) and partial (b) ablation of cervical folds. The grey area represents the tissue removed from each cervical fold and the dotted lines represent the sites of incision.

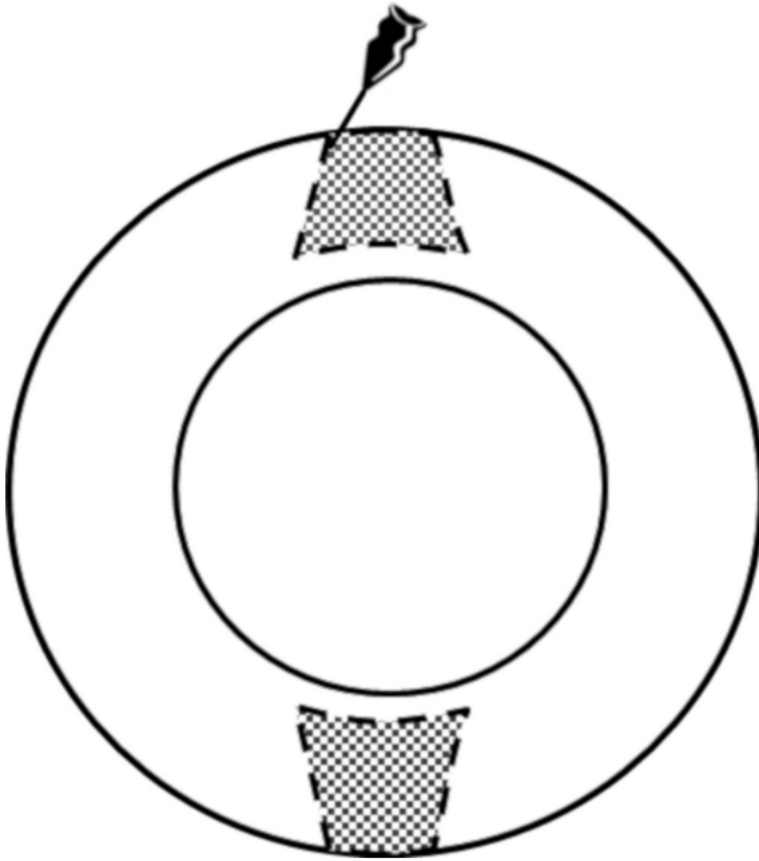


Fig. 3b. Schematic representation of total (a) and partial (b) ablation of cervical folds. The grey area represents the tissue removed from each cervical fold and the dotted lines represent the sites of incision.

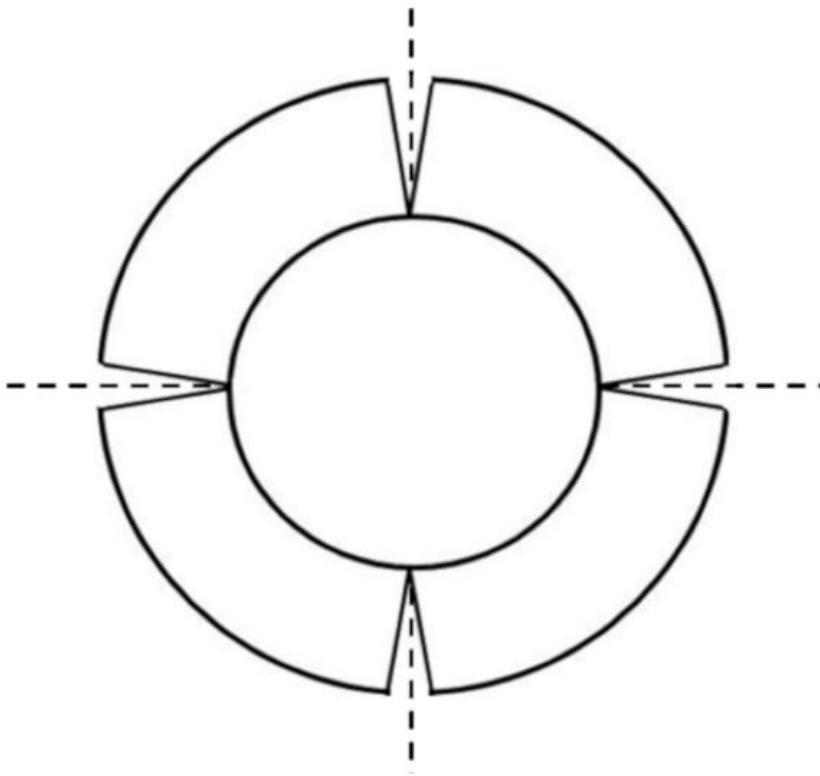


Fig. 4a. Schematic representation of the sites of incision of the cervical folds, indicated by dotted lines: a) 4 incisions (dorsal, ventral and 2 laterals).

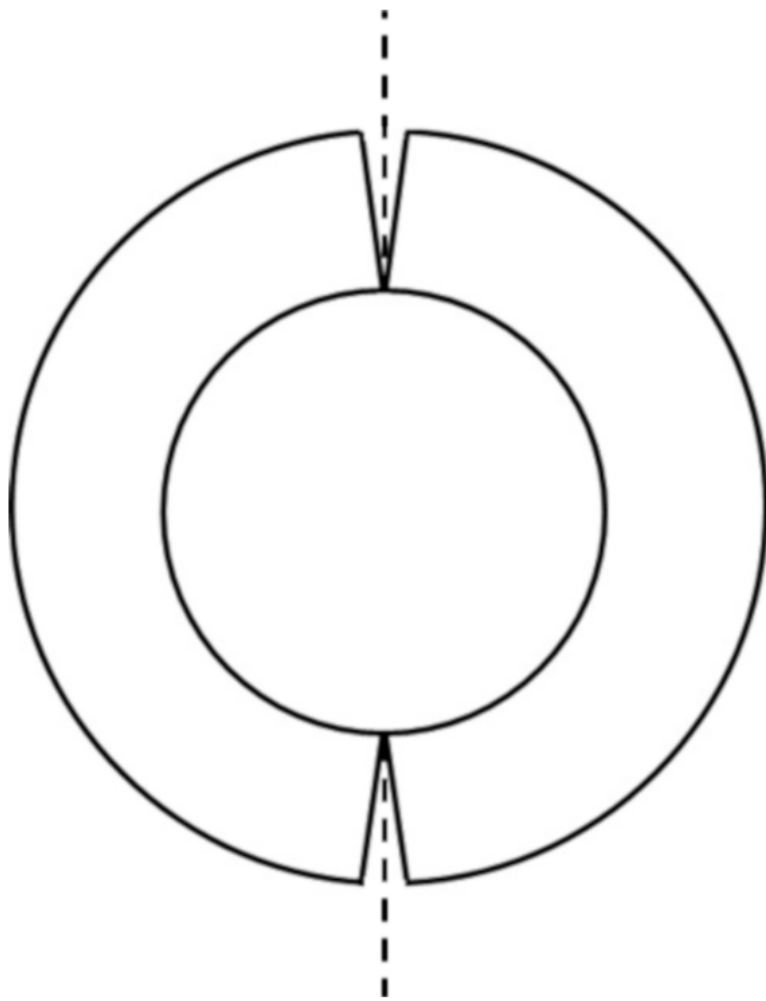


Fig. 4b. Schematic representation of the sites of incision of the cervical folds, indicated by dotted lines: b) 2 incisions (dorsal and ventral).

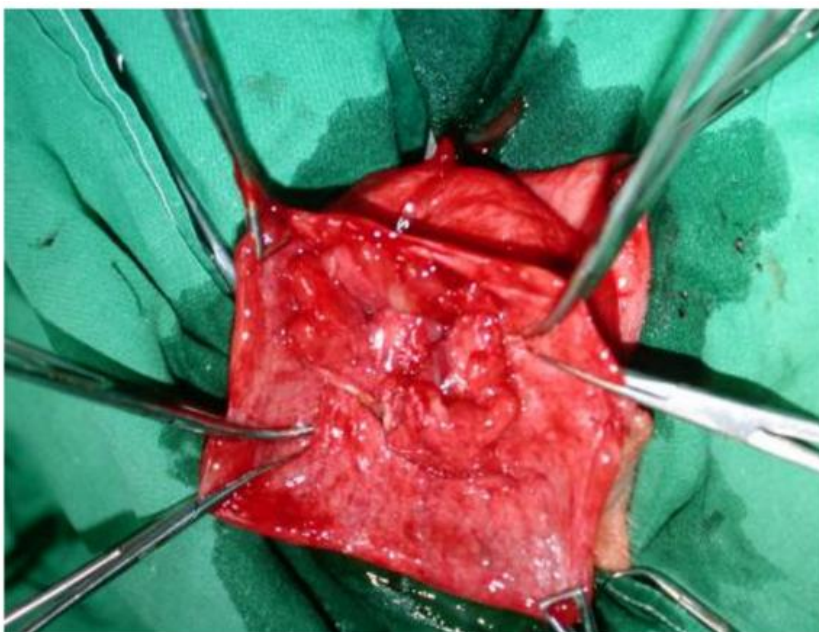


Fig. 5a. Incision of cervical folds: a) areas were delimited by Dandy forceps (arrows).

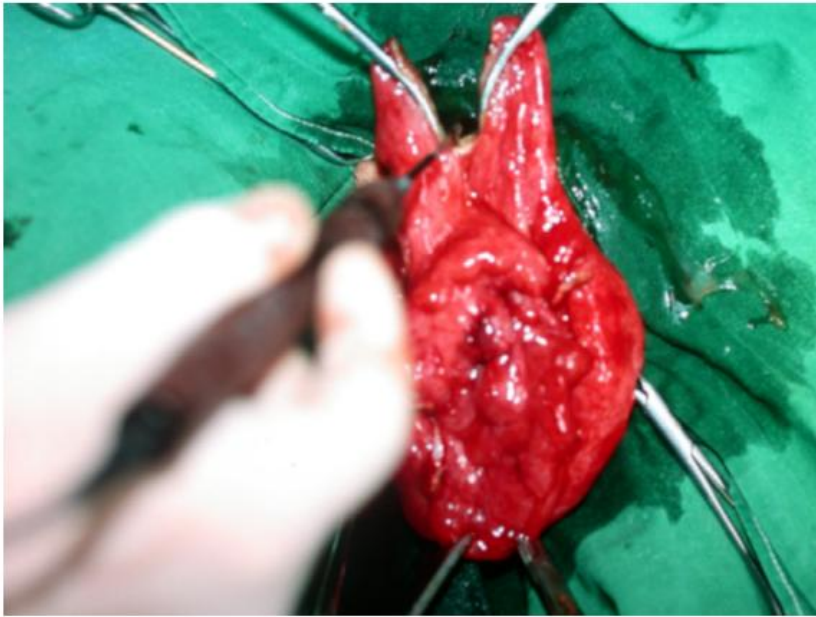


Fig. 5b. Incision of cervical folds: b) cut by electrocautery.

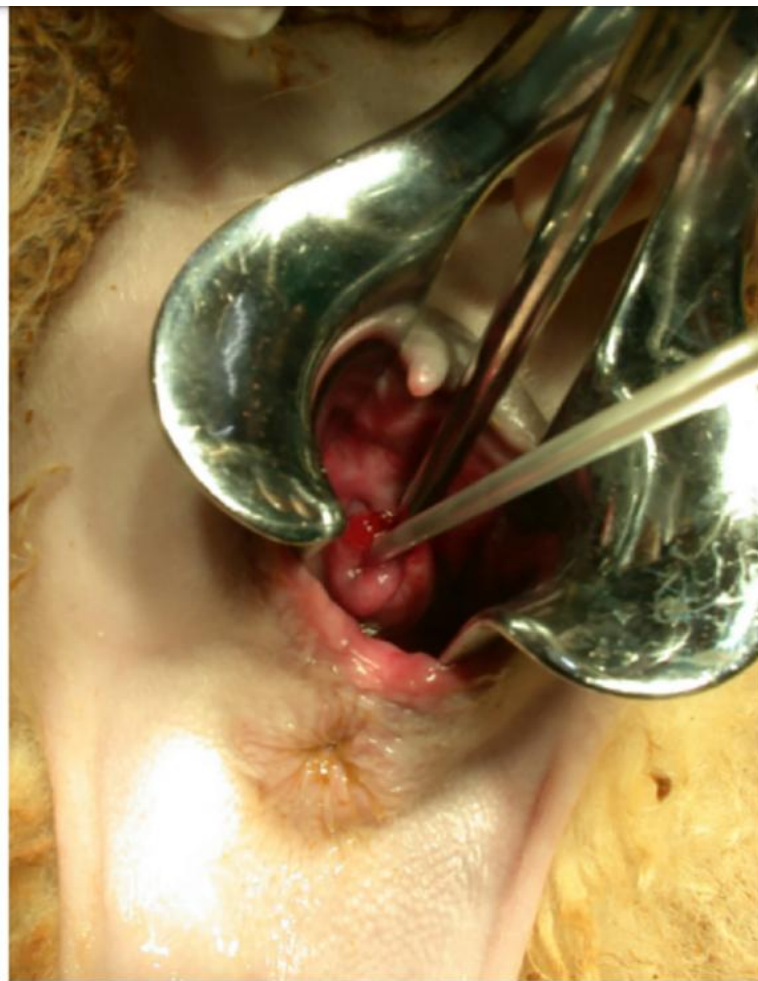


Fig. 6. Transcervical artificial insemination with frozen-thawed semen after incision of cervical folds: the fold of the external os of the cervix was gently extruded using Bozeman forceps up to the vulvar vestibulum and the insemination catheter was inserted in the cervical lumen and semen was deposited, when possible, directly in the uterus.

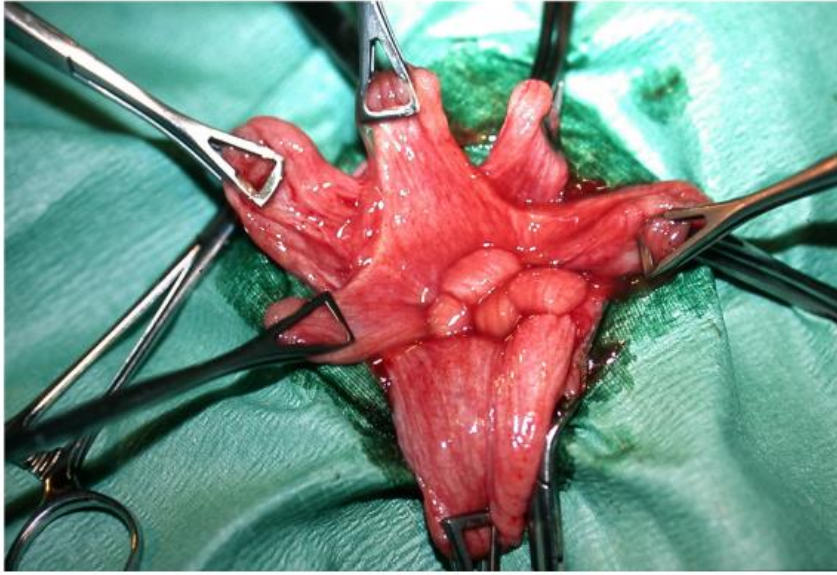


Fig. 7. Cervix of a ewe submitted to 4 incisions of cervical folds after post-surgical lambing.

Table 1
Cervical patency following 4 different surgical procedures on cervical folds.

	Surgical procedure on cervical folds	Ability to reach the uterus		Easiness in reaching the uterus (% score)				Time to reach the uterus (sec. \pm SD)
			%	I	II	III	IV	
Exp. 1	Total ablation	5/5	100	100^a	0^a	0	0	2.4 \pm 0.5^a
	Partial ablation	20/20	100	100^a	0^a	0	0	4.9 \pm 3^a
Exp. 2	4 incisions	40/44	90.9	65^{ab}	25^{ab}	7.5	2.5	21 \pm 26^b
	2 incisions	26/29	89.6	38.5^b	46.1^b	15.4	0	26.2 \pm 21^b

The easiness in passing through the cervical canal refers to ewes in which reaching the uterine lumen was possible. Scores: I) very easy; II) easy; III) moderately difficult; IV) difficult. The time taken to reach the uterus is expressed in mean \pm SEM. Different superscripts indicate significant differences among procedures for $P < 0.05$.

Table 2
Site of deposition of frozen-thawed semen, pregnancy and lambing rates in control group and in ewes submitted to 2 or 4 incisions of the cervical folds.

	N° tot	Site of deposition of semen		Pregnant ewes	Pregnancy rates (%)	Ewes at lambing	Lambing rates(%)
		Uterus	Cervix				
Control	25	0	25	2/25	8^a	1/25	4^a
4 Incisions of cervical folds	44	40	4	28/44	63.7^b	25/44	56.8^b
2 Incisions of cervical folds	29	25	4	12/29	41.4^b	12/29	41.4^b

Different superscripts (^a, ^b) indicate significant differences within column for $P < 0.05$.